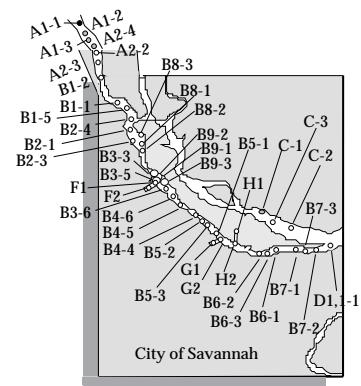
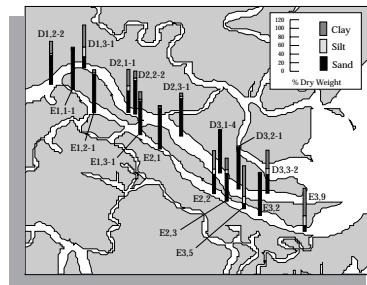
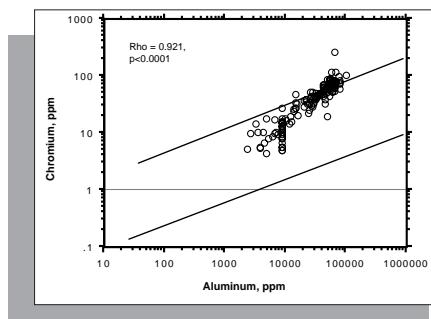
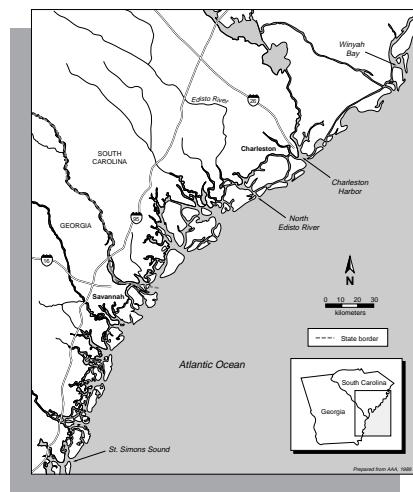


**National Status and Trends Program
for Marine Environmental Quality**

Magnitude and Extent of Sediment Toxicity in Selected Estuaries of South Carolina and Georgia



Silver Spring, Maryland
April 1998

US Department of Commerce

noaa National Oceanic and Atmospheric Administration

Coastal Monitoring and Bioeffects Assessment Division
Office of Ocean Resources Conservation and Assessment
National Ocean Service

Coastal Monitoring and Bioeffects Assessment Division
Office of Ocean Resources Conservation and Assessment
National Ocean Service
National Oceanic and Atmospheric Administration
U.S. Department of Commerce
N/ORCA2, SSMC4
1305 East-West Highway
Silver Spring, MD 20910

Notice

This report has been reviewed by the National Ocean Service of the National Oceanic and Atmospheric Administration (NOAA) and approved for publication. Such approval does not signify that the contents of this report necessarily represents the official position of NOAA or of the Government of the United States, nor does mention of trade names or commercial products constitute endorsement or recommendation for their use.

Magnitude and Extent of Sediment Toxicity in Selected Estuaries of South Carolina and Georgia

Edward R. Long

*National Oceanic and Atmospheric Administration,
National Ocean Service, Office of Ocean Resources Conservation
and Assessment*

Geoffrey I. Scott, John Kucklick, Michael Fulton, Brian Thompson

*National Oceanic and Atmospheric Administration,
National Marine Fisheries Service, Southeast Fisheries Science Center*

R. Scott Carr, James Biedenbach

U. S. Geological Service/National Biological Service

K. John Scott , Glen B. Thursby

Science Applications International Corporation

G. Thomas Chandler

University of South Carolina

Jack W. Anderson

Columbia Analytical Services

Gail M. Sloane

Florida Department of Environmental Protection



Silver Spring, Maryland
April 1998

United States
Department of Commerce

William M. Daley
Secretary

National Oceanic and
Atmospheric Administration

D. James Baker
Under Secretary

National Ocean Service

Nancy Foster
Assistant Administrator

INTRODUCTION

Background. Toxic chemicals can enter the marine environment through numerous routes: stormwater runoff, industrial point source discharges, municipal wastewater discharges, atmospheric deposition, accidental spills, illegal dumping, pesticide applications and agricultural practices. Once they enter a receiving system, toxicants often become bound to suspended particles and increase in density sufficiently to sink to the bottom. Sediments are one of the major repositories of contaminants in aquatic environments. Furthermore, if they become sufficiently contaminated sediments can act as sources of toxicants to important biota. Sediment quality data are direct indicators of the health of coastal aquatic habitats.

Sediment quality investigations conducted by the National Oceanic and Atmospheric Administration (NOAA) and others have indicated that toxic chemicals are found in the sediments and biota of some estuaries in South Carolina and Georgia (NOAA, 1992). This report documents the toxicity of sediments collected within five selected estuaries: Savannah River, Winyah Bay, Charleston Harbor, St. Simons Sound, and Leadenwah Creek (Figure 1).

As a component of its National Status and Trends (NS&T) Program, NOAA monitors toxicant concentrations in selected locations throughout the nation and surveys the biological significance of toxicant accumulations in selected regions. In the monitoring component of the program, mollusks and demersal fishes are captured annually for chemical analyses of their tissues. Sediments are collected and analyzed for a suite of metals and organic parameters. Spatial patterns and temporal trends in chemical concentrations are determined from the data (O'Connor and Ehler, 1991; O'Connor, 1991). Chemical analyses of sediments collected at each sampling site were performed at many of the sites the first year that each site was sampled. The monitoring activities were initiated in 1983 and have continued each year to the present time.

The NS&T Program has analyzed oyster and sediment samples from the lower Winyah Bay, the Santee River, the lower Savannah River, and two sites in Charleston Harbor along with fishes and sediments from Charleston Harbor and lower Savannah River (Lauenstein et al., 1993). These data revealed that the concentrations of a number of chemicals were significantly elevated in sediments relative to background conditions. Detectable concentrations of PCBs, DDTs, chlordane, mirex, and polynuclear aromatic hydrocarbons (PAHs) occurred in sediment samples from these sites (NOAA, 1991). Many chlorinated pesticides also occurred in the tissues of resident oysters from the sites in South Carolina and Georgia (NOAA, 1989). The concentrations of some trace metals in some samples exceeded the concentrations expected in reference areas based upon normalization to aluminum content (Hanson and Evans, 1991). Concentrations of arsenic, cadmium, selenium, total PAHs, and tributyltins were particularly high in some areas and years (O'Connor and Baliaeff, 1995).

Furthermore, scattered among the estuaries of South Carolina and Georgia are a number of hazardous waste sites with very high chemical concentrations. One site adjacent to Purvis Creek in St. Simons Sound that was near a defunct chemical manufacturer had extremely high PCB and mercury concentrations in sediments and fish (Bronstein, 1995). Sediment and pore water samples from this area were toxic in laboratory tests and the toxicity was attributable, at least in part, to high concentrations of methylmercury and PCBs in the sediments (Winger et al., 1993). Several small sites in and adjacent to Charleston Harbor have had a history of elevated concentrations of different chemicals and have been the focus of remedial investigations (Heyward Robinson, Charleston Harbor Project, personal communication).

Analyses of age-dated sediment cores from the Savannah River estuary have shown a history of contamination by anthropogenic chemicals, including mercury, chromium, lead, PAHs, dieldrin, DDT and PCBs during the 1950s and 1960s (Alexander et al., 1994). Concentrations have gradually decreased during the past 20-30 years and recently-deposited, surficial sediments have lower chemical concentrations.

Sources of pollution in Charleston Harbor include industrial and municipal point sources, urban and suburban nonpoint sources, septic tank overflows, and runoff from forested urban and agricultural watersheds (Matthews et al., 1980). The number of permitted point sources in Charleston Harbor, however, have decreased from 115 in 1969 to 78 in 1986 (SCSGC, 1992; Davis and Van Dolah, 1992). Portions of Charleston Harbor, including the Ashley and Cooper rivers, have had relatively high concentrations of anthropogenic trace metals, PCBs and pesticides in the sediments.

There are 13 current point source discharges to Winyah Bay, including the discharges from the International Paper Mill which contained the highest concentrations of dioxins among mills surveyed in 1989 (DHEC, unpublished data). A health advisory that warned people not to eat fish and shellfish because of high dioxin concentrations in the Sampit River, a tributary to the Winyah Bay, was lifted in 1992 (South Carolina Department of Health and Environmental Control). Pesticide use in the Winyah Bay watershed has been very high relative to the size of the watershed (South Carolina Sea Grant Consortium, 1992). Kucklick and Bidleman (1994) reported high concentrations of several pesticides and PAHs in Winyah Bay.

Leadenwah Creek, a small tributary to the North Edisto River estuary, receives considerable agricultural runoff (Scott et al., 1990; 1993). Runoff of pesticides such as azinphosmethyl, endosulfan, and fenvalerate, from nearby vegetable farms has caused major fish kills, and other impacts to fish, oysters, and macropelagic fauna (Scott et al., 1988; Scott et al., 1993; Fulton, 1989). Toxicity of the sediment-associated pesticides fenvalerate and endosulfan to meiobenthic animals has been demonstrated in laboratory tests performed with samples collected from Leadenwah Creek. Toxicity tests of fenvalerate included measures of reproductive success, in which depressed egg production and mean clutch sizes were observed in copepods (Chandler, 1990). Endosulfan inhibited larval colonization and early juvenile growth among polychaetes but did not appear to affect meiobenthic copepods (Chandler and Scott, 1991).

Current Survey Rationale. Because of the presence of anthropogenic chemicals in sediments and biological tissues, the relatively high risks of adverse effects to living marine resources, the documented urban and industrial development of the estuaries and a lack of information on toxicant effects, NOAA elected to study this area. Sediments were collected throughout the area over a two-year period to determine if there was an effect on biota based on the use of a battery of laboratory toxicity tests.

The objectives of the survey were to:

- (1) determine the presence and severity of toxic responses;
- (2) estimate the spatial extent of toxicity;
- (3) identify spatial patterns of toxicity in each system; and
- (4) characterize the relationships between toxicity and the concentrations of potential toxicants in the sediments.

Sampling and testing methods used in previous surveys performed elsewhere in the USA were employed in this survey. A wide variety of candidate measures of toxicant effects were evaluated and compared to determine which would be most useful in NOAA's surveys (Wolfe, 1992; Long and Buchman, 1989). Batteries of assays performed with sediments, bivalve molluscs, and demersal fishes in selected regions have been used to form a weight of evidence with regard to the presence and incidence of toxicant-associated bioeffects. Analyses of sediment toxicity have been included in these regional assessments to provide an estimate of potential effects of sediment contaminants on resident benthic populations. Batteries of toxicity tests appropriate for analyzing sediment toxicity were selected following evaluations of a number of candidates (Long and Buchman, 1989).

Thus far, sediment toxicity surveys have been performed by NOAA in San Francisco Bay (Long and Markel, 1992); Tampa Bay (Long et al., 1994); Long Island Sound (Wolfe et al., 1994); Hudson-Raritan estuary (Long et al., 1995); Boston Harbor (Long et al., in press); Los Angeles/Long Beach Harbor (Sapudar et al., 1994); San Diego Bay (Fairey et al., 1996); western Florida panhandle (Sloane et al., in press); and in several other areas in which the surveys.

Study Area. The study area included Winyah Bay, Charleston Harbor, Leadenwah Creek, Savannah River, and St. Simons Sound (Figure 1). In Winyah Bay the study area included the lower Sampit River, Georgetown Harbor, and lower Winyah Bay seaward to approximately the junction with the Intracoastal Waterway (Figure 2). The Charleston Harbor survey area included the Ashley River below I-95, the Cooper River from approximately Goose Creek, and Wando River from approximately Nowell Creek, Charleston Harbor, and stretched seaward to the mouth of the harbor at Fort Sumter (Figure 3). In Leadenwah Creek, samples were collected from the head of the creek to its junction with the North Edisto River estuary (Figure 4). In the Savannah River the study area extended from approximately Interstate - 95 to the mouth of the river and included several industrial harbors and the south river channel that parallels the Savannah River seaward to approximately Cockspur Island (Figures 5). In St. Simons Sound the study area included the lower Turtle River, Brunswick Harbor (East River), Brunswick River, the Back River, Terry Creek, the lower estuary seaward to the mouth of the bay (Figure 6). As described below in Methods, each of the five study areas was stratified (sub-divided) into approximately equal strata and samples were collected from randomly-chosen locations.

METHODS

Sampling Design. Five estuaries were investigated during 1993 and 1994: Charleston Harbor, Winyah Bay, and Leadenwah Creek were sampled during May-June, 1993, and St. Simons Sound and Savannah River were sampled in June-July, 1994. The same sampling procedures, boat, and crew were used in both years.

The study area included saltwater portions of these five estuaries. Stratified, random sampling designs patterned after those of the EMAP-Estuaries surveys (Schimmel et al., 1994) were used in each area during the selection of sampling stations. Each bay was subdivided into irregular-shaped strata. Large strata were established in the open waters of the bays

where toxicant concentrations were expected to be uniformly low. This approach provided the least intense sampling effort in areas known or suspected to be relatively homogenous in sediment type and water depth and relatively distant from contaminant sources. In contrast, relatively small strata were established in urban harbors, bayous and tributaries nearer suspected sources in which conditions were expected to be heterogeneous or transitional. Sampling effort was more intense in the small strata than in the large strata. The large strata were roughly equivalent in size to each other small strata were roughly equivalent in size to each other.

This approach combines the strengths of a stratified design with the random-probabilistic selection of sampling locations. Data generated within each stratum can be attributed to the dimensions of the stratum. Therefore, these data can be used to estimate the spatial extent of toxicity with a quantifiable degree of confidence (Heimbuch, et al., 1995). Strata boundaries were established to coincide with the dimensions of major basins, bayous, waterways, etc. in which hydrographic, bathymetric and sedimentological conditions were expected to be relatively homogeneous.

The locations of individual sampling stations within each strata were chosen randomly using the EMAP computer software and hardware (Dr. Kevin Summers, U. S. EPA, Environmental Research Laboratory, Gulf Breeze, FL). One to three samples were collected within each stratum. Usually, four alternate locations were provided for each station in a numbered sequence. The coordinates for each alternate were provided in tables and were plotted on the appropriate navigation chart. In a few cases the coordinates provided were inaccessible. They were rejected and the vessel was moved to the next alternate.

Sample Collection. At each station the sampling vessel was piloted to the first alternate location for the sample collection. If the station was inaccessible or if the material at the location was only coarse sand and gravel with no mud (silt+clay) component, that alternate location was abandoned and the second (third, or fourth, if needed) alternate was sampled. In almost all cases the first or second alternates were acceptable and were sampled.

Vessel positioning and navigation were aided with a Trimble NavGraphic XL Global Positioning System (GPS) unit and a compensated LORAN C unit. Both systems generally agreed very well with each other when both were operational. Both were calibrated and their accuracy verified each morning at a channel marker within the study area.

Samples were collected with a Kynar-lined 0.1m² modified van Veen grab sampler (also, known as a Young grab) deployed with an electric windless aboard the state of Florida R/V *Raja*. The grab sampler and sampling utensils were acid washed with 10% HCl at the beginning of each survey, and thoroughly cleaned with site water and acetone before each sample collection. Usually 3 or 4 deployments of the sampler were required to provide a sufficient volume of material for the toxicity tests and chemical analyses. The upper 2-3 cm. of the sediment were sampled to ensure the collection of recently-arrived materials. Sediments were removed with a plastic scoop and accumulated in a stainless steel pot. The pot was covered with a Teflon plate between deployments of the sampler to minimize sample oxidation and exposure to shipboard contamination. The material was carefully homogenized in the field with a stainless steel spoon before it was distributed to prepared containers for each analysis.

The portions of samples to be tested for amphipod survival and sea urchin bioassays were shipped in 2 gal. polyethylene jugs. Portions for Microtox™ tests were shipped in 100 mL glass jars and portions for chemical analyses were shipped in pre-cleaned, 250 mL I-Chem glass jars with Teflon lids. Jars and lids were labelled with information on sample and station numbers.

Samples were shipped in ice chests packed with water ice or blue ice to the testing laboratories by overnight courier. They were accompanied by chain of custody forms and station labels.

Locations of the individual sampling stations in each area are illustrated in Figures 7-13, and coordinates for each are listed in Appendix A. Field log notes containing information on depth and sediment characteristics at each station are also listed in Appendix A. A total of 162 samples were tested for toxicity in this survey. In Charleston Harbor 63 samples were collected, 52 were chosen randomly by NOAA and 9 were chosen for specific locations by the Charleston Harbor Project (CHP) (Figure 7). Samples were collected along the Ashley, Cooper, and Wando rivers, throughout Charleston Harbor, and seaward through the lower harbor to Fort Sumter (Figure 7). Nine samples were collected in Leadenwah Creek, a tributary to the North Edisto River previously impacted by agricultural runoff (Figure 8). In Winyah Bay nine samples were collected in the lower Sampit River, Georgetown Harbor, and upper Winyah Bay (Figure 9). In St. Simons Sound 20 samples were collected from locations in the Turtle River, Brunswick Harbor, Terry Creek, Back River, and St. Simons Sound (Figure 10). In the Savannah River 60 samples were collected throughout three different segments of the system: from the mouth of the river to Fort Johnson (Figure 11), from Fort Johnson to Port Wentworth (Figure 12), and upstream above Port Wentworth (Figure 13). One sample was collected in a reference area, North Inlet-Oyster Landing, north of Charleston.

Multiple toxicity tests were performed on all sediment samples. Chemical analyses were performed on a subset of samples from each estuary for trace metals, butyl tins, polynuclear aromatic hydrocarbons, chlorinated pesticides and PCBs following a review and evaluation of the toxicity test results. Amphipod survival tests were performed by the National Biological Service (now U.S. Geological Survey) laboratory in Corpus Christi, TX., and Science Applications International Corporation in Narragansett, RI. Microtox™ bioluminescence tests and chemical analyses were performed by the National Marine Fisheries Service laboratory of NOAA in Charleston, SC. Sea urchin fertilization tests were performed by the National Biological Service (now U.S. Geological Survey) laboratory in Corpus Christi, TX. Copepod reproduction tests were performed by the University of South Carolina in Columbia, SC. Cytochrome P-450 RGS bioassays were performed by Columbia Analytical Services in Carlsbad, CA.

Amphipod Survival Test. The amphipod tests are the most widely and frequently used assays in sediment evaluations performed in North America. They are performed with adult crustaceans exposed to relatively unaltered, bulk sediments. *Ampelisca abdita* has shown relatively little sensitivity to nuisance factors such as grain size, ammonia, and organic carbon in previous surveys. In previous surveys, the NS&T Program has observed wide ranges in responses among samples, strong statistical associations with toxicants, and small within-sample variability (Long et al., 1994; Wolfe et al., 1994; Long et al., 1995).

Ampelisca abdita is a euryhaline benthic amphipod that ranges from Newfoundland to south-central Florida, and along the eastern Gulf of Mexico. The amphipod test with *A. abdita* has been routinely used for sediment toxicity tests in support of numerous EPA programs, including EMAP in the Virginian, Louisianian, and Carolinian provinces (Schimmel et al., 1994). Amphipod toxicity tests followed ASTM protocols (ASTM, 1990, 1992). In the first year, amphipod tests of samples from Charleston Harbor, Winyah Bay, and Leadenwah Creek were conducted by the National Biological Service (NBS) laboratory. In the second year amphipod assays were conducted by Science Applications International Corporation, (SAIC).

In year one, test animals were purchased from Brezina and Associates of Dillon Beach, California. Amphipods were packed in native sediment with 8-10 liters of seawater in doubled plastic bags. Oxygen was injected into the bags and shipped via overnight courier to the testing lab at Port Aransas. Upon arrival, amphipods were acclimated and maintained at 20°C for one day prior to the initiation of the test.

Control sediments for year one testing included sediment collected from the natural habitat of the amphipods in California, and a reference sediment from Redfish Bay, Texas. The Redfish Bay sediments had been used in previous sediment quality assessment studies by the U.S. Fish & Wildlife Service (then, NBS, now USGS). Both the control and reference sediments were handled in the same manner as the samples from South Carolina.

For year two testing, amphipods were collected by SAIC from tidal flats in the Pettaquamscutt (Narrow) River, a small estuary flowing into Narragansett Bay, Rhode Island. Animals were held in the laboratory in pre-sieved uncontaminated ("home") sediments under static conditions. Fifty percent of the water in the holding containers was replaced every second day when the amphipods were fed. During holding, *A. abdita* were fed laboratory cultured diatoms (*Phaeodactylum tricornutum*).

Control sediments were collected by SAIC from the Central Long Island Sound (CLIS) reference station of the U.S Army Corps of Engineers, New England Division. These sediments have been tested repeatedly with the amphipod survival test and other assays and found to be non-toxic (amphipod survival has exceeded 90% in 85% of the tests) and un-contaminated (Wolfe et al., 1994; Long et al., 1995). Sub-samples of the CLIS sediments were tested along with each series of samples from Savannah River and St. Simons Sound.

Amphipod testing performed by both laboratories followed the procedures detailed in the Standard Guide for conducting 10 day Static Sediment Toxicity Tests with Marine and Estuarine Amphipods (ASTM, 1990, 1992). Briefly, amphipods were exposed to test and negative control sediments for 10 days with 5 replicates of 20 animals each under static conditions using filtered seawater. For the NBS test, 250 mL of test or control sediments were delivered into 1-liter glass jars, and 700 mL of seawater were added to each jar. The SAIC procedure differed somewhat: 200 mLs of test or control sediments were placed in the bottom of the test chamber, and covered with approximately 600 mL of filtered seawater (28-30 ppt). For both sets of tests, air was provided by air pumps and delivered into the water column through a pipette to ensure acceptable oxygen concentrations, but suspended to ensure that the sediments would not be disturbed. Temperature was maintained at ~20°C by either incubator (NBS) or water bath (SAIC). Lighting was continuous during the 10 day exposure period to inhibit the swimming behavior of the amphipods. Constant light inhibited emergence of the

organisms from the sediment, thereby maximizing the amphipod's exposure to the test sediments.

Twenty healthy, active animals were placed into each test chamber, and monitored to ensure they burrowed into sediments. Non-burrowing animals were replaced, and the test initiated. The jars were checked daily, and records kept for dead animals, and animals on the water surface, emerged on the sediment surface, or in the water column. Those on the water surface were gently freed from the surface film to enable them to burrow, and dead amphipods were removed.

Tests were terminated after ten days. Contents of each of the test chambers were sieved through a 0.5 mm mesh screen. The animals and any other material retained on the screen were examined under a stereomicroscope for the presence of amphipods. Total amphipod mortality was recorded for each test replicate.

During year 1 the NBS laboratory ran tests in two batches of 41 and 42 samples each with holding times of <10 days and <15 days, respectively. Sample holding times for most tests in year 2 were <8 days; however, due to poor performance in negative controls some samples were re-tested and holding times ranged up to 38 days. A positive control test was used to document the sensitivity of each batch of test organisms. The positive control consisted of 96 hr water-only exposures to sodium dodecyl sulfate (SDS). LC50 values were calculated for each test run.

Sea Urchin Fertilization and Embryological Tests. Tests of sea urchin fertilization and embryo development have been used in assessments of ambient water and effluents and in previous NS&T Program surveys of sediment toxicity (Long et al., 1994; Carr et al., 1996). Test results have shown wide ranges in responses among test samples, excellent within-sample homogeneity, and strong associations with the concentrations of toxicants in the sediments. The tests, performed with the early life stages of sea urchins, have demonstrated high sensitivity.

Toxicity of sediment pore waters were conducted with the sea urchin *Arbacia punctulata*. These tests were performed during both years by the National Biological Service (NBS), National Fisheries Contaminant Research Center in Corpus Christi, Texas at their laboratory located in Port Aransas. Sea urchins used in this study were obtained either from jetties at Port Aransas, Texas, or from Gulf Specimen Company, Inc. (Panacea, Florida), and were acclimated to Port Aransas seawater before gametes were collected for testing.

Pore water was extracted from sediments with a pressurized squeeze extraction device (Carr and Chapman, 1992). Sediment samples were held refrigerated (at 4° C) until pore water was extracted. Pore water was extracted as soon as possible after receipt of the samples, but in no event were sediments held longer than 7 days from the time of collection before they were processed. After extraction, porewater samples were centrifuged in polycarbonate bottles (at 4200 g for 15 minutes in year 1, and in year 2 using a new centrifuge at 1200 g for 15 minutes was adequate) to remove any particulate matter, and were then frozen. Two days before the start of a toxicity test, samples were moved from a freezer to a refrigerator at 4° C, and one day prior to testing, thawed in a tepid water bath. Experiments performed by NBS have demonstrated no effects upon toxicity attributable to freezing of the pore water samples.

Sample temperatures were maintained at $20\pm 1^\circ \text{C}$. Sample salinity was measured and adjusted to 30 ± 1 ppt, if necessary, using ultrapure sterile water or concentrated brine. Other water quality measurements were made for dissolved oxygen, pH, sulfide and total ammonia. Temperature and dissolved oxygen were measured with YSI meters; salinity was measured with Reichert or American Optical refractometers; pH, sulfide and total ammonia (expressed as nitrogen, TAN) were measured with Orion meters and their respective probes. The concentrations of un-ionized ammonia (UAN) were calculated using respective TAN, salinity, temperature, and pH values.

Each of the porewater samples was tested in a dilution series of 100%, 50%, and 25% of the water quality adjusted sample with 5 replicates per treatment. Dilutions were made with clean, filtered ($0.45 \mu\text{m}$), Port Aransas laboratory seawater. Tests followed the methods of Carr and Chapman (1992). Pore water from a reference area in Redfish Bay, Texas, an area located near the testing facility and in which sediment porewaters have been determined to be non-toxic in this test (e. g., Long et al., 1994), was included with each toxicity test as a negative (non-toxic) control. Adult male and female urchins were stimulated to spawn with a mild electric shock, and gametes collected separately.

For each test, $50 \mu\text{L}$ of 1:5000 diluted sperm were added to each vial with 5 mL of porewater sample and incubated at $20\pm 2^\circ \text{C}$ for 30 minutes. One mL of a well mixed dilute egg suspension was added to each vial, and incubated an additional 30 minutes at $20\pm 2^\circ \text{C}$. Two mLs of a 10% solution of buffered formalin solution was added to stop the test. Fertilization membranes were counted after the 1-hour exposures and fertilization percentages calculated for each replicate test. In the embryological tests, 100 embryos were examined in each replicate after 48-hour exposures to pore water. Both percent normal and percent abnormal embryological development were recorded for each porewater concentration (100%, 50%, and 25%) following methods of Carr et al., 1996.

Microbial bioluminescence (Microtox™) tests. This is a test of the relative toxicity of extracts of the sediments prepared with an organic solvent, and, therefore, it is immune to the effects of nuisance environmental factors, such as grain size, ammonia and organic carbon. Organic toxicants, and to a lesser degree trace metals, that may or may not be readily bioavailable are extracted with the organic solvent. Therefore, this test can be considered as a test of potential toxicity. In previous NS&T Program surveys, the results of Microtox™ tests have shown extremely high correlations with the concentrations of mixtures of organic compounds (Long et al., 1994; Long et al., 1995; Wolfe et al., 1994).

The Microtox™ assay was performed with dichloromethane (DCM) extracts of sediments following the basic procedures used in testing Puget Sound sediments (U.S. EPA, 1986, 1990, 1994) and San Francisco Bay sediments (Long and Markel, 1992). All sediment samples were stored in the dark at 4°C for 5-10 days before processing was initiated. A 3-4 g. sediment sample from each station was weighed, recorded, and placed into a DCM rinsed 50 mL centrifuge tube. A 15 g portion of sodium sulfate was added to each sample and mixed. Pesticide grade DCM (30 mL) was added and mixed. The mixture was shaken for 10 seconds, vented and tumbled overnight.

Sediment samples were allowed to warm to room temperature and the overlying water discarded. Samples were then homogenized with a stainless steel spatula, and 15-25 grams of

sediment were transferred to a centrifuge tube. The tubes were spun at 1000 g for 5 min. and the pore water was removed using a Pasteur pipette. Three replicate 3-4 g sediment subsamples from each station were placed in mortars containing a 15 g portion of sodium sulfate and mixed. After 30 min., subsamples were ground with a pestle until dry. Subsamples were added to 50 mL centrifuge tubes. Then, 30 mL of DCM were added to each tube and shaken to dislodge sediments. Tubes were then shaken overnight on an orbital shaker at a moderate speed. Tubes were then centrifuged at 500 g for 5 min and the sediment extracts transferred to TurbovapTM tubes. Then, 20 mL of DCM was added to sediment, shaken by hand for 10 sec and spun at 500 g for 5 min. The previous step was repeated once more and all three extracts were combined in the TurbovapTM tube. Sample extracts were then placed in the TurbovapTM and reduced to a volume of 0.5 mL. The sides of the TurbovapTM tubes were then rinsed down with methylene chloride and again reduced to 0.5 mL. Then, 2.5 mL of dimethylsulfoxide (DMSO) were added to the tubes which were returned to the TurbovapTM for an additional 15 min. Sample extracts were then placed in clean vials and 2.5 mL of DMSO were added to obtain a final volume of 5 mL DMSO.

A suspension of luminescent bacteria, *Vibrio fischeri*, (Azur Environmental, Inc.) was thawed and hydrated with toxicant-free distilled water, covered and stored in a 4°C well on the MicrotoxTM analyzer. To determine toxicity, each sample was diluted into four test concentrations. Percent decrease in luminescence of each cuvette relative to the reagent blank was calculated. Based upon these data, the sediment concentrations that caused a 50% decrease in light production (EC50's) were reported.

A negative control (extraction blank) was prepared using DMSO, the test carrier solvent. A phenol standard (45 mg/L phenol) was run after re-constitution of each vial of freeze-dried *V. fischeri*. In addition, a reference sediment was tested from North Inlet - an area shown to be non-toxic in sensitive laboratory tests in previous studies.

Copepod Reproduction Tests. Fourteen-day, chronic tests of reproductive success of the meiobenthic copepod *Amphiascus tenuiremis* were performed on 14 selected samples. The fourteen samples were selected to represent a presumed pollution gradient within Charleston Harbor. Analyses followed the standard protocols of Chandler (1990), Chandler and Scott (1991) and Strawbridge et al. (1992). Samples were press-seived (0.125 mm) to remove meiofauna and large particles; 12 gram sieved aliquots were extruded into triplicate beakers filled with clean sterile-filtered artificial seawater. Then, 35 barren females and 15 males were removed from stock cultures and added to each beaker. Flow-through exposures were conducted for 14 days. Test animals were fed phytoplankton (*Isochrysis galbana* and *Dunaliella tertiolecta*) on days 3, 6, 9, and 12. Barriers consisting of 0.045 mm mesh screens prevented animal losses. After 14 days all males, females, clutch sizes and offspring were counted and compared with North Inlet negative controls.

Toxicological end-points included survival of adults at the end of 14 days, naupliar production (no. nauplia per sample), copepodite production (no. copepodites per sample), clutch size (no. eggs per gravid female per sample), and total production (total no. nauplii + copepodites per sample). Results were initially analyzed using SAS ANOVA/GLM (F statistic) and Tukey's Studentized Range Test (p<0.05).

Cytochrome P-450 RGS Assays. This assay of the light produced by luciferase in a reporter gene system (RGS) of cultured human liver cells was conducted on selected samples from Charleston Harbor and Winyah Bay. The assay has been responsive to the presence of mixed-function oxidase inducers such as dioxins, furans, high molecular weight PAHs, and coplanar PCBs in tissues and sediments (Anderson et al., 1995). The induction of cytochrome P-450 RGS by sediment extracts was determined in 29 samples selected to represent a gradient in response in the amphipod, Microtox™ and urchin tests.

In these tests, standard protocols (Anderson et al., 1995, 1996; ASTM, 1997; APHA, 1996) were followed to ensure comparability with data derived for other areas. Approximately, 20 g of sediment from each station were extracted by EPA method 3540 to produce 1-2 mL of DCM/ extract mixture. Small portions of these samples (2-10 uL) were applied to approximately one million human liver cells contained in three replicate wells with 2 mL of culture medium. After 18 hours of incubation, the cells were washed, then lysed, then the solution was centrifuged following EPA method 3540 to produce 2 mL extracts of dichloromethane. Small portions (10 mL) were used in measures of luminescence. Solvent blanks and the reference toxicant (2, 3, 7, 8 - dioxin) were tested with each batch of samples. The relative light unit (RLU) from the solvent blank was set equal to unity and all other RLUs were divided by (normalized to) that of the blank. The running average fold induction for 10nM dioxin is approximately 140 and the induction from 1 mg/mL of benzo(a)pyrene (b[a]p) was 60 fold. Data were converted to mg of b(a)p equivalents per g of sediment by multiplying the response (fold induction) from 10 mL of extract by a factor (200) to represent the total of inducing substances in the 2 mL extract, and then dividing by the factor of 60 and the dry wt. of the sample.

Chemical Analyses. Sediment samples were chosen for chemical analyses based upon an examination of the toxicity test results. Samples were chosen that represented gradients in the toxicity results and that also represented contiguous geographic stations. Chemical analyses were performed by the National Marine Fisheries Service laboratory, Charleston, SC. Analytical methods followed performance-based analytical protocols and employed quality-assurance steps of the NS&T Program (Lauenstein and Cantillo, 1993).

Extraction for organic analyses: The methods for extraction and sample preparation are similar to those of Krahn *et al.* (1988) with few modifications. Approximately 8.5 g of sediment was dried by mixing with 100 g of Na₂SO₄, which had been ashed for 16 h at 700°C. The dried sample was transferred to a Pyrex Soxhlet thimble and the internal standards D₈-naphthalene (200 ng), D₁₀-acenaphthalene (200 ng), D₁₀-phenanthrene (502 ng), D₁₀-fluoranthene (497 ng), D₁₂-perylene (102 ng), dibromooctafluorobiphenyl (DBOFPB; 20 ng), 2,2',4,5',6-pentachlorobiphenyl (PCB-103; 20 ng) and 2,2',3,3',4,5,5',6-octachlorobiphenyl (PCB-198; 20 ng) were added. The sample was then extracted in a Soxhlet apparatus with 250 mL of CH₂Cl₂ for 18 h. Sample extracts were reduced in volume by a stream of purified nitrogen using a nitrogen blow-down concentrator (Turbo Vap, Zymark Instruments) to ca. 0.5 mL. Lipids and other high molecular weight compounds were removed from the sample by gel permeation chromatography. The liquid chromatograph consisted of an autosampler (Gilson Model 231), a Waters HPLC pump (model 501), two 22.5 x 500 mm gel permeation columns in series (Phenomenex Phenogel, 100 Å pore size), a UV detector (Linear model UV-106) and a fraction collector (Gilson model 201). The mobile phase was CH₂Cl₂ at a flow rate of 7 mL/min. 400 mL of the sample was injected into the system with lipids and other high molecular

weight compounds eluting in the first 14 minutes. The fraction of interest was collected beginning 1 min before the retention time of DBOFBP and ending 2 min after perylene. The resulting fraction was reduced in volume as above and the CH_2Cl_2 was replaced with hexane and concentrated to a final volume of ca. 0.5 mL. At this point, elemental sulfur was removed from the sample by treatment with activated copper. To remove remaining polar interferences, the sample was then transferred to a 6 g cyanopropyl solid phase extraction cartridge (Varian, prerinsed with 6 mL of hexane) and eluted with 12 mL of hexane.

Polycyclic aromatic hydrocarbon (PAH) analysis: PAHs were quantified by two methods, capillary GC-ITMS and HPLC with fluorescence detection. Details of the analytes measured, internal standards, quantitation ions (GC-ITMS) and fluorescence excitation and emission wavelengths (HPLC-fluorescence) are available from NOAA. The instrument used for the GC-ITMS analysis was a Finnigan MAT Magnum Ion Trap Mass Spectrometer equipped with a Varian 3400 gas chromatograph and Varian 8200 autosampler. The column was a 30 m x 0.25 mm (i.d.) DB-5ms (J&W Scientific) with a film thickness of 0.25 mm. The carrier gas was helium at a linear velocity of 33 cm/sec at 300 °C. The temperatures were 280°C, 220°C and 280°C for the injection, ion source and transfer line, respectively. The acquisition scan range was from 50-285 amu with a scan rate of 0.6 sec/scan. The sample was injected (1 mL) using a splitless Grob technique (1 minute split time) at an initial oven temperature of 45°C. After a one minute hold, the oven was ramped to 110°C at 25°C/min (one minute hold), ramped to 300 °C at 10°C/min then finally ramped to 320 °C with a 3.5 minute hold. The instrument was calibrated using a mixed standard of the analytes with the internal standards injected at five concentrations. The calibration was verified at the start and end of each sample sequence and every 8 hours in between using a mid-level calibration standard and recalibrated as necessary. The target analytes were identified both by the GC retention time window (15 sec) and the presence and ratios of fragmentation ions relative to the molecular ion.

PAHs were additionally quantified using HPLC with fluorescence detection utilizing a method similar to Wise *et al.* (1988) and Schantz *et al.* (1990). The instrument consisted of two HPLC pumps (Waters 6000A), a 680 gradient controller (Waters model 680) and an autosampler (Waters WISP). The column dimensions were 6 mm X 25cm, with a 5 mm particle size (Supelco LC-PAH) and the column was heated to 30°C (Fiaton TC-50 column heater controller and a CH-30 column heater). The solvent was pumped at a constant flow rate of 1.5 mL/min with a gradient program that started with a two minute hold at 60% water:40% acetonitrile followed by a linear ramp to 55% water:45% acetonitrile in 15 minutes and a final ramp to 0 % water:100% acetonitrile in 35 minutes with a 10 minute hold. Fluorescence was monitored with two fluorescence detectors (Perkin Elmer LC-240 and LS-4) connected in series at wavelengths specific to individual PAHs. The separation between deuterated and nondeuterated PAHs was 0.44, 0.40 and 0.41 min for phenanthrene, fluoranthene and perylene, respectively. A NIST certified PAH standard solution and the deuterated PAH internal standards were used to calibrate the instrument. Sample peaks were identified by retention times and fluorescence specific wavelength.

Organochlorine and pesticide analysis: Chlorine-containing compounds were analyzed using gas chromatography with electron capture detection (GC-ECD; Hewett-Packard 5890 series II). The instrument was configured with two columns, a 30 m x 0.25 mm i.d. (0.25 mm film thickness) DB-5 (5% phenyl; J&W Scientific) and a column with the same dimensions and 50% phenyl (Rtx-50; Restek Corp.). The carrier gas was held at a constant average

linear velocity of 33 cm/sec by pressure programming the injector. The carrier and detector makeup gasses were helium and argon:methane (95%:5%), respectively. The injector and detector temperatures were 250 and 320°C, respectively. The sample was injected (2 mL) using a splitless Grob technique (1 min split time). The sample was then split such that nearly equal portions were sent to each column. The initial oven temperature was 50°C with a one minute hold, followed by a ramp to 170°C at 4°C/min, then from 170°C to 210°C at 1°C/min then from 210 to 310°C at 4°C/minute with a 10 min hold.

The instrument was calibrated using a mixed standard of the target analytes (chlorinated pesticides, NIST SRM 2261; and PCB congeners, NIST/NOAA intercalibration mixture) prepared with the internal standards (DBOFPB, PCB 103 and PCB 198). The standard was prepared in three concentrations that bracketed the sample concentrations. The calibration curve was verified at the beginning of each sample set by injecting the mid-level continuing calibration with a check standard which was required to be within $\pm 20\%$ of the known value for each analyte or the instrument was recalibrated. Retention data was simultaneously acquired from the two columns and was used to identify the analytes. On each column, unknown peaks were identified relative to that in the standard. If the unknown analyte peak fell within ± 0.05 minutes of the standard and was present on both columns, then the peak was determined to be authentic. The analyte amount was determined on each column and the lower amount was reported.

Extraction for metals analyses: From the original sample, 20 g was transferred to a 30 mL acid-washed plastic sample cup then the sediment and cup were weighed to 0.0001 g. The sample was then covered and dried at 70°C for 24 h. After drying, the sample was reweighed to determine percent moisture. The dried sediment was then ground with a mortar and pestle and transferred to a 20 mL plastic screw-top container.

Samples were extracted using a closed-vessel, concentrated acid microwave digestion technique. A 0.5 g subsample of the ground sediment was weighed (0.0001 g) into a Teflon-lined digestion vessel. To this, 10 mL of concentrated HNO₃ (Instra-analyzed) plus 0.5 mL deionized water was added to the vessel. The sample was then microwaved using a well ventilated, 600 watt corrosion-resistant digestion microwave (CEM Model MDS-2000) for 2 hours at full power and 120 psi. The sample was allowed to cool, then 2.0 mL of 30% H₂O₂ was added to the vessel, which was then microwaved for an additional 10 min at full power and 80 psi. After cooling, the digestate was filtered (#41 filter paper) into a 50 mL volumetric flask and brought to volume with deionized water. The sample was then transferred by pouring into a 50 mL polypropylene conical centrifuge tube for analysis.

A separate extraction procedure was used for Hg. From the dried sediment sample, 0.2 to 0.5 g of sediment was weighed (0.0001 g) then transferred to a 300 mL biological oxygen demand bottle (BOD). To the bottle, 5 mL, 1.25 mL and 3.75 mL of deionized water, HNO₃, and HCl were added, respectively. The bottles were then placed in a 95 °A 5 °C water bath for 2 min. The bottles were removed, cooled to room temperature then 50 mL deionized water and 15 mL 5% KMnO₄ (w/v) solution was added to the BOD bottle. The bottle was then returned to the water bath for an additional 30 min, removed, and allowed to cool for at least 1 h. The bottles were then uncapped and 6 mL of a NaCl-hydroxyamine hydrochloride solution (12 g NaCl + 12 g hydroxyamine hydrochloride) and 55 mL deionized water were added.

Metals Analysis: A suite of metals (Al, As, Cd, Cr, Cu, Fe, Pb, Mn, Ni, Sn, Zn) were analyzed by inductively coupled plasma spectroscopy (ICP). The instrument (Perkin Elmer Plasma 1000 with autosampler) was calibrated initially by developing a standard curve for each element. The concentrations used spanned two orders of magnitude, with the lowest concentration being the method blank. The response factor was determined as the slope of the standard curve line (absorbance/mg metal). Prior to samples analysis, the calibration standards were analyzed to verify the standard curve. The check standards were a solution of Zn (5 mg/L), Cd (2 mg/L), Cr, Cu, and Ni (1 mg/L each), a solution of Al (100 mg/L), Fe (50 mg/L) and individual standards of Mg (10 mg/L), As (1 mg/L) and Sn (100 mg/L). In addition to the above calibration standards, a solution was analyzed with the concentration equal to the midpoint of the calibration curve for each metal to evaluate drift from the original calibration curve. This same standard was diluted 5:1 with deionized water as an interference check solution. Lastly, a quality control sample (SRM Marine Sediment MESS-2 from the National Research Board of Canada) was analyzed. The samples extracts were then analyzed in duplicate and the results averaged.

The metals Ag, As, Cd, Pb, and Se were analyzed by graphite furnace atomic absorption (graphite furnace-AA). A Perkin Elmer 5100 Atomic Absorption Spectrometer with a Zeeman HGA 600 Graphite Furnace was used for analyses. Prior to sample analysis, the instrument was calibrated with five separate concentrations of each metal spanning approximately 1 order of magnitude. The modifier $Mg(NO_3)_2$ was used for Ag, As, and Cd; $Ni(NO_3)_2$ for Se, and $PO_4-Mg(NO_3)_2$ modifier for Pb. Samples were analyzed in duplicate and the results averaged.

Mercury was analyzed by cold-vapor atomic absorption using a Leeman Labs PS200 mercury analyzer at a wavelength of 253.7 nm. Prior to sample analysis, a five point calibration curve was constructed. Samples were analyzed in duplicate and the Hg concentration determined from the slope of the calibration curve and the sample absorption.

Acid Volatile Sulfide (AVS) and Simultaneously Extractable Metals (SEM) Analyses: The general procedure for measuring AVS and SEMs was based on Allen *et al.* (1991) with a few modifications. The analytical train for releasing sulfide and metals from sediment consisted of a N_2 gas supply segment, and a reaction/trap segment. Nitrogen was supplied by a tank of compressed high-purity nitrogen which flowed through two 500 mL gas-washing bottles and then to floating-ball flow meters positioned immediately before the 6 individual reaction/trap segments in series. The first gas washing bottle in the nitrogen segment contained a solution of vanadous chloride (VCl_2), while the second bottle contains de-aerated, deionized water. A third empty gas washing bottle was incorporated to restrict liquids from traveling down line. The reaction/trap portion consisted of a 500 mL round-bottom flask with a septum inlet and a glass tubing inlet/outlet and two traps (impingers). The round-bottom flask contained the sediment sample, water and 20 mL of 6 M HCl added to the flask to volatilize the sulfide and metals. The two impingers contained 0.5 M NaOH to trap the H_2S generated in the reaction in the round bottom flask after the HCl addition. The extraction line contained six sample trains (round bottom flask + 2 traps) each controlled by an independent gas flow meter. The recovery of the trap system of AVS was 85% based on blanks amended with standardized sodium sulfite solution. To measure AVS, ca. 5 g of wet homogenized sediment was weighted into the boiling flask. Deionized water (80 mL) was then added to

the flask with a small Teflon-coated stir bar. This was connected to the line and the HCl-addition port sealed with a rubber septum. The sediment-deionized water mixture was then purged with nitrogen for 10 minutes to remove residual oxygen. The flow was then stopped then 20 mL of 6M HCl was added to each flask through the septum via a syringe. The sample was stirred with a magnetic stirrer and the reaction was allowed to proceed for 1.5 h. After the reaction was complete, the solution in each boiling flask was filtered through a 0.45 mm membrane filter (Gelman Sciences), with the flask rinsed several times with deionized water and the rinses added to the filtrate. The total volume of the filtrate was measured and a 50.0 mL aliquot was removed for SEMs analysis. The NaOH traps were developed by adding 10 mL of a mixed diamine reagent and allowing the mixture to react for 30 minutes. The solution was quantitatively transferred to a 100 mL volumetric flask and brought to volume. Approximately 2 mL of solution was transferred to a cuvette and the absorbance at 670 nm was read using a spectrophotometer (Milton Roy Spectronic 601). To calibrate the AVS method, a standard sulfite solution was prepared by weighing 12 g of $\text{Na}_2\text{S}\cdot 9\text{H}_2\text{O}$ into 1.0 L of deionized water. The solution was standardized by the sodium thiosulfate titration procedure described in Allen *et al.* (1991) using a starch indicator. Absorbance was measured at 670 nm and used with solutions of known concentration to construct a standard curve. Simultaneously extracted metals were measured in the 50.0 mL aliquot removed from the sediment extract. The acid treatment removes metals which are weakly associated with the sediments and not incorporated in crystalline matrices. Samples were analyzed by ICP for the suite of metals described previously for that technique.

Grain Size and Total Organic Carbon Analysis: Grain size was measured using a sieving and pipette method as described in Plumb (1981). The mass of sediment used varied between 20 and 50 g of wet sediment depending on the texture (more for sandier sediments). Total organic carbon was determined by first weighing 10 mg of dried sample onto a pre-combusted glass fiber filter then analyzing with a Perkin Elmer 2400 Elemental Analyzer (950 °C combustion temperature). Cysteine was used as an external calibration standard.

Quality Controls for PAHs: To monitor for the efficiency of extraction and interferences that may be introduced in the sample preparation scheme, both spiked matrix samples and blanks were analyzed using both HPLC with fluorescence detection and GC-ITMS. A total of six spiked matrix samples were analyzed using NMFS sample 216. Amounts of each analyte spiked along with the percent recovery were recorded. A standard reference material (SRM 1941; Organics in Marine Sediments) was obtained from the National Institute of Standards and Technology (NIST) to evaluate the efficiency of our extraction methods for removing PAHs from sediment. From 0.47 to 0.50 g dry weight of SRM 1941 was extracted seven times and analyzed. In general, PAHs analyzed by either method were within the NIST upper and lower confidence limits for the certified PAHs. The largest deviation from the NIST SRM material was for benzo(k)fluoranthene, which was 68% less than the stated NIST mean value when analyzed by GC-ITMS. Overall, the average deviation from the mean NIST SRM values (both certified and non-certified) was only 1% (some observations were biased high and some low) by either HPLC or GC-ITMS. The method detection limit (MDL) was determined as three times the standard deviation of repeated matrix spike determinations according to the Environmental Protection Agency protocol stated in the CFR (1991). A series of six blanks were also analyzed to check for contamination introduced during analysis.

Quality Control for Organochlorines and pesticides: Similar to the analysis of PAHs, both spiked sediments and a NIST SRM were analyzed for organochlorine compounds (NIST SRM 1941). Sediments were amended with 21 PCB congeners and 15 organochlorine pesticides or metabolites at three levels ranging from 1.2 ng to 5 ng total. The overall recovery (mean \pm standard deviation) of organochlorines from amended sediments was 102 \pm 23% for PCBs and 89 \pm 32% for organochlorine pesticides plus metabolites. The results of the analysis of SRM 1941 are available from NOAA. The overall precision was not as good as for PAHs or metals and is evident by only three analytes falling between the NIST upper and lower confidence limits. In general, there is little apparent systematic bias in the deviation from the certified values (mean deviation = -1.6%).

Quality Control for Metals: Several precautions were taken to avoid contamination during metals analysis. Plastic containers and utensils were used wherever practical. All labware was washed thoroughly with soap and water, rinsed with tap water then three times with distilled water and then soaked in a 50% concentrated nitric acid bath. Following the cleaning procedure, the glassware was air dried and stored in covered plastic container. As a check for contamination, two types of blanks were analyzed. The first consisted of a 15% nitric acid solution and was used as the endpoint of the daily calibration curve. The second type was a solution that was processed using the extraction procedure to check for contamination that may arise during this process. A total of seven blanks of this type were analyzed. The limit of detection (LOD) was determined from the blank information as the mean blank plus three times the standard deviation. LODs ranged from 0.05 mg/kg for Cd by graphite furnace AA to 390 mg/kg for Pb by ICP. To evaluate the efficiency of the nitric acid microwave digestion procedure, a standard reference material was analyzed seven times. Recoveries ranged from 64% recovery for Al to 175% recovery for Cd by ICP and averaged (mean \pm standard deviation) 95 \pm 25%. All recoveries were within the acceptable confidence limits of the SRM material.

Quality Control for Acid Volatile Sulfide: To validate the AVS procedure, both an intercalibration exercise with Skidaway Institute of Oceanography (SKIO) and spike recoveries were conducted. A total of 10 spiked blanks were analyzed during two recovery experiments. The first experiment consisted of five blanks spiked with 0.5, 1.0, 1.5, 2.0 and 2.5 mmoles of AVS and resulted in a recovery (mean \pm standard deviation) of 85 \pm 2.7%. The second recovery experiment consisted of five blanks spiked with 3.01 mmoles of AVS and resulted in a recovery of 87 \pm 0.8%. The intercalibration exercise was the analysis by both NMFS-Charleston and SKIO of a field-collected sediment sample for AVS. The AVS measured in the sample were comparable with SKIO obtaining 2.64 \pm 0.09 mmoles AVS (n=3) and NMFS-Charleston obtaining 2.08 \pm 0.31 mmoles AVS (n=5). The results of the spike recoveries and the intercalibration exercises indicate that the AVS method used by NMFS-Charleston is reasonably precise and accurate.

Method limits of detection (MLODs) for these analyses are summarized in Appendix B. They generally corresponded with the target MLODs attained by other participants in the NS&T Program (Lauenstein and Cantillo, 1993). Chemical analyses were performed according to the quality control/quality assurance procedures of the NS&T Program, including instrument calibration, use of internal standards, replication of some analyses, percent recoveries of spiked blanks, and analyses of standard reference materials.

Statistical methods. Percent amphipod survival data from each station that had a mean survival less than that of the control was compared to the control using a one-way, un-paired t-test (alpha = 0.05) assuming unequal variance. Data were not transformed since an examination of data from previous tests have shown that *A. abdita* percentage survival data met the requirement for normality. A one-sample t-test was used to compare data from each sampling block with the mean performance control (CLIS) for each stratum.

Significant toxicity for *A. abdita* is defined here as survival statistically less than that in the performance control (alpha = 0.05). In addition, samples in which survival was significantly less than controls and less than 80% of control values were regarded as “highly toxic” or “numerically significant”. The 80% criterion is used by EPA as a critical statistical value for *A. abdita* test data in EMAP-Estuaries methods (Holland, 1990). Similarly, the EPA/COE dredged material guidance manual (the “green book”) also consider sediments toxic if survival relative to a reference sediment is less than 80% (U.S. EPA/U.S. ACOE, 1990). Furthermore, statistical power curves created from SAIC’s extensive testing database with *A. abdita* show that the power to detect a 20% difference from the control is 90%.

Microtox™ data were analyzed using the computer software package developed by the manufacturer to determine concentrations of the extract that inhibited luminescence by 50% (EC50). This value was then converted to mg dry wt. of sediment/mL of extract (where dry wt. was calculated as the weight of sediment after removal of porewater). To determine significant differences of samples from each station, pair-wise comparisons were made between contaminated samples and results from control sediments using three different analyses. Following an ANOVA test, a sequence of three increasingly conservative statistical tests were performed to determine significant differences from controls: Mann-Whitney, Dunnett’s, and distribution-free. Dunnett’s analyses were performed with log-transformed data.

For both the sea urchin fertilization and morphological development tests, statistical comparisons among treatments were made using ANOVA and Dunnett’s one-tailed *t*-test (which controls the experiment-wise error rate) on the arcsine square root transformed data with the aid of SAS (SAS, 1989). The trimmed Spearman-Karber method (Hamilton et al., 1977) with Abbott’s correction (Morgan, 1992) was used to calculate EC50 (50% effective concentration) values for dilution series tests. Prior to statistical analyses, the transformed data sets were screened for outliers (SAS, 1992). Outliers were detected by comparing the studentized residuals to a critical value from a *t*-distribution chosen using a Bonferroni-type adjustment. The adjustment is based on the number of observations (*n*) so that the overall probability of a type 1 error is at most 5%. The critical value (CV) is given by the following equation: $cv = t(dfError, .05/(2 \times n))$. After omitting outliers but prior to further analyses, the transformed data sets were tested for normality and for homogeneity of variance using SAS/LAB Software (SAS, 1992).

Chemical data from the sample analyses were plotted on base maps to identify spatial patterns, if any, in concentrations. Trace metal concentrations were plotted against aluminum concentrations and compared to expected ratios for uncontaminated sediments developed by Schropp et al., 1988.

Similarly, the spatial patterns in toxicity were estimated by plotting data on base maps of each bay. Estimates of the spatial extent of toxicity were determined with cumulative distri-

bution functions in which the toxicity results from each station were weighted to the dimensions (km^2) of the sampling stratum in which the samples were collected (Schimmel et al., 1994). The size of each stratum (km^2) was determined by use of a planimeter applied to navigation charts, upon which the boundaries of each stratum were outlined. A critical value of less than 80% of control response was used in the calculations of the spatial extent of toxicity for all tests.

Chemistry/toxicity relationships were determined in a multi-step sequence similar to that followed in previous studies (e.g., Long et al., 1994; 1995a; 1996) to ensure comparability. First, simple Spearman-rank correlations were determined for each toxicity test and each physical/chemical variable. The correlation coefficients and their statistical significance were recorded and compared among chemicals. Second, for those chemicals in which a significant correlation was observed, the data were examined in scatterplots to determine if there was a reasonable pattern of increasing toxicity with increasing chemical concentration and if any chemical in the toxic samples equalled or exceeded published numerical guidelines. Scatterplots were prepared with un-transformed bioassay data.

Chemical concentrations expressed in dry wt. were compared with the ERM values of Long et al. (1995b) developed for NOAA. Also, the concentrations of three PAHs (acenaphthene, fluoranthene, and phenanthrene) and two pesticides (dieldrin and endrin) expressed in units of organic carbon were compared to proposed National sediment quality criteria (SQCs) developed by U. S. EPA (1994). Finally, the concentrations of un-ionized ammonia were compared to LOEC concentrations determined for the sea urchin tests by the U. S. National Biological Service (Carr et al., 1996) and NOEC concentrations determined for amphipod survival tests published by Kohn et al. (1994). As a part of this step, trace metal:aluminum ratios were compared to those from reference areas in the southeastern U. S. (Schropp et al., 1990).

Third, the numbers of samples out of those that were analysed that exceeded the respective guidelines were determined. The combined results of these three steps were examined to determine which chemical(s), if any, may have contributed to the observed toxicity and which probably had a minor or no role in toxicity.

Correlations were determined for all the substances that were quantified in each bay, usually including total (bulk) trace metals, metalloids, trace metals simultaneously extracted (SEM) with acid volatile sulfides (AVS), un-ionized ammonia (UAN), percent fines, total organic carbon (toc), chlorinated organic hydrocarbons (COHs), and polynuclear aromatic hydrocarbons (PAHs). In addition, chemical indices calculated as the sums or averages of quotients formed by dividing the chemical concentrations in the samples by their respective ERM values (from Long et al., 1995b) were compared with measures of toxicity. Those substances that showed significant correlations were indicated with one, two, or three asterisks, depending upon the significance of the correlations. In correlation analyses involving a large number of variables, such as in this study, some correlations could appear to be significant by random chance alone. Adjustments often can be made to account for this possibility. Note that in the results tables only those correlations shown with two or three asterisks would remain significant if the number of variables were taken into account in these analyses.

RESULTS

Toxicant Concentrations among Regions of the Study Area. Field notes taken from each sampling station are shown in Appendices A1-A3. Results of all toxicity tests and chemical analyses of the sediment samples are listed in Appendices B1-B3. Average, minimum, and maximum concentrations of selected substances were compared among six different regions (Table 1). The upper Savannah River included stations upstream of Fort Johnson and the lower Savannah River area included stations seaward of Fort Johnson.

Generally, the concentrations of most trace metals differed very little among the different regions of the study area (Table 1). For example, the concentrations of silver ranged from 0.12 to 0.48 throughout all areas and the average concentrations in the regions were very similar. The concentrations of chromium, lead and zinc, however, were slightly higher in Winyah Bay as compared to the other regions. The average concentrations of the five summed simultaneously-extracted trace metals (SEM: Cd, Cu, Ni, Pb, Zn) also differed relatively little among regions; some samples from Charleston Harbor, upper Savannah River, and Winyah Bay had relatively high concentrations. The average ratios of SEM concentrations to AVS concentrations were most elevated in the upper Savannah River and lowest in the Winyah Bay samples. Numerous samples in all regions had SEM/AVS ratios of less than 1.0, suggesting that trace metals were not highly bioavailable in most samples.

Average concentrations of major organic compounds were often highest in St. Simons Sound (Table 1). The average concentrations of total analyzed PAHs quantified with GC/MS were similar in St. Simons Sound and Charleston Harbor (approximately 2000 ng/g) and both areas had samples with unusually high concentrations. The concentrations of total DDTs (sum of DDTs plus metabolites) were relatively low and similar in all regions. One sample from St. Simons Sound had the effect of elevating the average concentration for that area. The average concentrations of total PCBs (double the sum of 20 congeners) were elevated considerably in St. Simons Sound, again as the result of one sample with extremely high PCB concentrations (1776 ng/g).

Toxicant Concentrations in Charleston Harbor. The concentrations of individual chemical substances and classes of chemicals are listed for each sample in Appendix B. In addition, these data are available in spreadsheet format from NOAA.

In the Charleston Harbor area, sediment texture differed considerably among stations (Figure 14). Samples collected in the lower harbor (stations C1-C4) were predominantly sand. Samples from the Ashley River were predominantly sand in the lower and upper stretches of the river and finer-grained silts and clays in the mid-river reaches. Texture also was heterogeneous in the Cooper River; some samples were primarily sand and others were mainly silts and clays. In the Wando River most samples were primarily sand. As expected, based upon the texture, the total organic carbon (toc) content varied considerably among stations (Figure 15). TOC content was relatively low (less than 1%) in most samples from the lower harbor, Wando River, lower Ashley River and a few scattered stations in the Cooper River. Most of the samples from the upper Ashley and Cooper river stations had 2-5% TOC. Aluminum concentration, an indicator of the presence of fine-grained particles, was highly variable among stations; showing a distributional pattern similar to that of TOC (Figure 16).

The concentrations of many potentially toxic substances followed distributional patterns similar to those of texture, TOC, and aluminum. For example, cadmium concentrations were lowest in the lower harbor, lower Ashley River, and Wando River and highest in the upper Ashley and Cooper rivers (Figure 18). Cadmium concentrations also were relatively high in Shipyard Creek (G stations). However, none of the cadmium concentrations equalled or exceeded the ERL value of Long et al. (1995b).

All of the other trace metals generally followed patterns similar to that of cadmium (Figure 19-27). Copper concentrations were highest in the upper Ashley and Cooper rivers, Shipyard Creek, and station CHP 10, however, only one sample (CHP 10) exceeded the ERL value for copper (Figure 19). A sample collected in upper Shipyard Creek had a very high chromium concentration, exceeding the ERL value for chromium (Figure 20). Several samples from the upper Ashley River exceeded the ERL value for lead (Figure 21). One sample from the Cooper River exceeded the ERM value for mercury (Figure 22). Nickel concentrations were relatively high in some samples from Shipyard Creek and the Cooper River (Figure 23). The highest concentrations of silver occurred in samples from the upper Ashley River and Shipyard Creek, however, none equalled the ERL value for silver (Figure 24). The Shipyard Creek stations also had relatively high concentrations of tin (Figure 25); no numerical guidelines have been derived for total tin. The concentrations of zinc were highest in samples from the upper Ashley River and Shipyard Creek, however, in all stations these concentrations were below the ERL value (Figure 26). The total concentrations of five divalent simultaneously-extracted metals (SEM) were highest in a few stations from the Ashley and Cooper rivers and Shipyard Creek, and as discussed later, most concentrations exceeded the respective acid-volatile sulfides (AVS) concentrations in the samples (Figure 27).

The concentrations of total PAHs were highest in station CHP 3 near downtown Charleston and several stations in the lower Ashley River (Figure 28). Some of the samples from the lower Ashley River with elevated PAH concentrations had petroleum-like sheens when collected. Total PAH concentrations exceeded the ERL value, but were far below the ERM value for total PAHs (Figure 28). Total DDT concentrations were low relative to the ERL and ERM values and relatively uniform throughout the area (Figure 29). Similarly, the concentrations of total PCBs were relatively low in all stations except station H5-2 in the Cooper River in which the total PCB concentration (161 ppb) nearly equalled the ERM value of 180 ppb.

Based upon these chemical data, toxicity would be most expected in samples from the upper Ashley and Cooper rivers and Shipyard Creek and least likely in the Wando River, lower Ashley River and lower harbor where the sediments were primarily sandy with relatively low chemical concentrations. However, relative to applicable ERM values, most chemical concentrations were not very elevated and the higher TOC concentrations in these samples may minimize toxicity by inhibiting contaminant bioavailability.

Toxicant concentrations in Leadenwah Creek. Four samples from Leadenwah Creek were analyzed for chemical concentrations. At the uppermost station, the sediments were primarily sand and the concentrations of silt + clay increased down the estuary toward the confluence with the North Edisto River (Figure 31). Similarly, the concentrations of TOC increased down the estuary (Figure 32) from less than 1% to over 3%. The distribution of arsenic (Figure 33) exemplifies the patterns that were apparent for most substances, that is, concentrations gradually increased down the estuary with increasing fines and TOC concentrations. This overall

pattern was evident with most trace metals and organic compounds (Appendix B). The concentrations of mercury increased sharply at the most seaward station (Figure 34). The concentrations of total PAHs also increased slightly from the upper to the lower regions of the estuary (Figure 35). In contrast the uppermost station had the highest concentration of total DDTs (Figure 36). Most substances occurred in concentrations far below numerical guidelines; the exceptions being arsenic, mercury, and total DDTs which exceeded ERL values by slight amounts in one or a few samples. None of the substances, however, exceeded respective ERM concentrations.

Toxicant concentrations in Winyah Bay. Chemical analyses were performed on all nine samples collected in Winyah Bay and both reference samples collected in North Inlet. Samples collected in the Georgetown Harbor and entrance channel were primarily silts and clays while the three samples from Winyah Bay had some sand (Figure 37). Percent sand decreased down the bay from station B5 to station B7. As expected, the concentrations of aluminum also were high in the harbor stations and were relatively low in the bay stations (Figure 38). In following with the grain size, the TOC content was highest (3-5%) in the harbor stations and lowest in station B5 which had the lowest percent fines (Figure 39). Similarly, the concentrations of arsenic were highest in the inner harbor stations and lowest at the sandy sediments of station B5 (Figure 40); some concentrations exceeding the ERL value for arsenic. Concentrations of cadmium paralleled those of arsenic; however, none of the concentrations equalled or exceeded the ERL value for arsenic (Figure 41). The spatial patterns for chromium, copper, and lead concentrations were similar to those of the other metals, closely following the distribution of fine-grained materials and TOC (Figures 42-44).

None of the concentrations of total DDTs were particularly elevated, although station B1-3 had a total DDT concentration in excess of the ERL value (Figure 45). Also, none of the samples had high total PCB concentrations, although, again, station B1-3 had the highest concentration of the nine samples. Similarly, the concentrations of total PAHs were highest in the sample from station B1-3; however, none of the concentrations equalled or exceeded the ERL value for total PAHs (Figure 47).

The two reference stations in North Inlet had consistently low toxicant concentrations, usually near or below detection or quantification limits. In two different samples the concentrations of fines were 15.2% and 45.4%; the concentrations of sands were 84.8% and 54.6%; and the concentrations of TOC were 0.7% and 1.8%. The concentrations of naturally-occurring trace metals, as expected, were higher in the sample with the highest concentrations of fines and aluminum. None of the toxicant concentrations measured would be expected to pose a toxicological risk.

Toxicant concentrations in Savannah River. With some exceptions scattered through the area, most stations in the upper Savannah River strata had primarily fine-grained sediments (Figure 48). Sediments in many of the B, G, and H strata were sticky clay with silt and some sands. A few stations in the C and B, however, were primarily sand with little or no silts and clays. In the lower sections of Savannah River, in contrast, many samples were primarily sand (Figure 49). Stations in the D and E strata often were sandy with minor amounts of fines. However, sediments from stations E3-5 and E3-9 had only small amounts of sand and were primarily clay. Six samples from strata A, the most inland stratum (not shown), had variable amounts of sand, silt, and clay in mixed sediments. Total organic carbon content

ranged from less than 1% to nearly 8%. Most samples, however, had 2-4% TOC (Figures 50-51) with low concentrations in sandy samples and high concentrations in fine-grained samples. Station A2-4 (not shown) had an unusually high TOC content of nearly 8%.

Cadmium concentrations were highest in samples collected directly adjacent to the city of Savannah (strata B3, B9) and in a Back River (station C-1) station, exceeding the ERL value for cadmium of 1.2 ug/g (Figure 52). The different concentration scale in the lower reaches of the river (Figure 53) reflect the considerably lower concentrations observed below Fort Johnson. Cadmium concentrations were very low (<0.35 ug/g) in strata A1 and A2 (not shown).

As observed with cadmium, the concentrations of zinc were highest near the city of Savannah (often >100 ug/g) and diminished considerably downstream (Figures 54-55). Note again that the scales in Figures 54 and 55 are different. All samples, including those from strata A1 and A2 (not shown), had zinc concentrations below the ERL value of 150 ug/g. Spatial patterns in the concentrations of other trace metals generally followed those observed with zinc and most elements equalled or exceeded their respective ERL values in a few samples; none exceeded the ERM values.

Detectable concentrations of total DDTs were observed primarily in stations in the upper reaches of the river. Station F2 (Dundee Canal) directly adjacent to the city of Savannah and station A1-2 in the upper river (not shown) had the highest concentration; otherwise, most stations had undetectable DDT concentrations (Figures 56-57). Total PCB concentrations also were highest in station F2, followed by station C3 in Back River (Figure 58), and diminished considerably downstream (Figure 59). Total PCB concentrations in all samples, including those from strata A1 and A2 were well below the ERL value for total PCBs (22.7 ng/g). Total PAH concentrations were low (<4022 ng/g, the ERL value) in nearly all samples (Figures 60-61). The highest PAH concentrations occurred in samples from stations H1 and B2-4 in Stevens Terminal and the upper river, respectively (Figure 60). PAH concentrations were considerably lower in the lower reaches of the river than in the upper reaches.

Toxicant concentrations in St. Simons Sound. Chemical analyses were performed with all twenty samples collected in St. Simons Sound. Samples from most stations in the Turtle River (strata A, D, and E) and the lower Sound (stratum G) were sandy, while samples from the Brunswick Harbor (strata B and C) and upper Back River/Terry Creek (stratum F, station G1) were dominated by fine-grained materials (Figure 62). TOC content was relatively low (<2%) in most of the sandy stations and somewhat higher (>3%) in samples from stations with high percent silt and clay (Figure 63). Terry Creek and Brunswick Harbor stations had the highest TOC concentrations.

Compared to the other trace metals, the concentrations of chromium, lead and mercury, were elevated relative to ERL/ERM values. Brunswick Harbor samples (B and C strata) had the highest chromium concentrations (exceeding the ERL value of 81 ug/g) followed by those from Terry Creek (F stratum). All samples from the Turtle River, lower sound, and lower Back River had very low chromium concentrations (Figure 64). Lead concentrations were elevated above the ERL value (46.7 ug/g) only in stations from inner Brunswick Harbor (B stratum) and Terry Creek (F stratum); all other samples had very low lead concentrations (Figure 65). The distribution of elevated levels of mercury was further restricted to one sta-

tion in Purvis Creek (P-1), two stations in Terry Creek, and one Back River station (Figure 66). The ERL value for mercury (0.15 ug/g) was exceeded in these four samples.

The concentrations of both total DDTs and total PCBs were very high in the sample from station P-1 in Purvis Creek and relatively low in all other samples (Figures 67, 68). In the sample from station P-1, the total DDT concentration (15 ng/g) exceeded the ERL value (1.58 ng/g) by a considerable amount but was lower than the ERM value (46.1 ng/g). Most (12 ng/g) of the DDT was in the form of o,p'-DDD. In other stations (B1, B2, C1, and G1) with detectable amounts of DDT, this pesticide was primarily in the form of p,p'-DDD, p,p'-DDT, and p,p'-DDE. In the Purvis Creek sample the sum of 20 PCB congeners was 886.8 ng/g, equivalent to a total PCB concentration of 1774 ng/g. This concentration is well above the ERM value of 180 ng/g. The PCB at this station was primarily in the form of 9- and 10-chlorine congeners (Appendix B). Total PCB concentrations in samples from stations A3 and B1 also exceeded the ERM value for total PCBs.

Samples from three stations (B1, B2, and F2) exceeded the ERL value for total PAHs (Figure 69); all other stations had relatively low PAH concentrations. The PAHs in these three samples were primarily high molecular weight (4- and 5-ring) compounds, notably pyrene, fluoranthene, and benzo fluoranthene (Appendix B).

Incidence of Toxicity among Regions. All tests were performed according to widely-accepted protocols. Tests of both negative (non-toxic) controls and positive (pollutant) controls were within acceptable limits.

In the amphipod tests all measures of dissolved oxygen, temperature, salinity, and pH showed the tests were run within acceptable limits for *Ampelisca abdita*. Un-ionized ammonia concentrations were below levels known to cause toxicity. In the 1993 tests of samples from Charleston Harbor/Winyah Bay/Leadenwah Creek performed by NBS, the tests of SDS in water showed 96-h LC50s of 13.76 mg SDS/L (95% confidence limits of 13.24-14.29 mg/L) and 14.73 mg SDS/L (95% confidence limits of 13.64-15.90 mg/L). During year 2 the LC50 values calculated by SAIC ranged from 5.27 mg SDS/L to 11.22 mg SDS/L (no confidence limits calculated). These data suggested that the amphipods from New England were slightly more sensitive than those from San Francisco Bay. Both results are acceptable.

Survival ranged from 85-89% in San Francisco Bay reference sediments in the amphipod tests in the 1993 tests. In the 1994 tests of St. Simons Sound/Savannah River samples performed by SAIC, control (central Long Island Sound) survival ranged from 76-98% in eleven experiments. Samples in which control survival dropped below acceptable levels (85%) were re-tested with control survivals increasing to 92% and 98% in the re-tests. The 1994 tests of the positive controls showed LC50s ranging from 5.27 to 11.22 mg SDS/L, suggesting somewhat higher sensitivity among east coast amphipods compared to those from the west coast.

In the 1993 urchin fertilization tests, porewater dissolved oxygen ranged from 78 to 104% saturation and all sulfide concentrations were below detection (0.01 mg/L). Values for pH ranged from 7.04 to 8.31. The concentrations of ammonia ranged from 1.2 to 249.0 ug/ L and averaged 54.3 ug/ L, well below the toxicity threshold (LOEC) of 800 ug/ L established by NBS. Percent fertilization success in the Redfish Bay, Texas control was 97.4% in 100%

porewater. In the North Inlet reference sample fertilization success was 97.6% in 100% porewater. Similarly, in the 1994 tests dissolved oxygen concentrations ranged from 86 to 128% saturation and sulfide concentrations were not detectable. Values for pH ranged from 6.97 to 8.67. Un-ionized ammonia concentrations ranged from 4.1 to 1263 ug/l, exceeding the toxicity threshold in only one sample (B2-3 in the upper Savannah River).

In the Microtox™ tests performed by NMFS, results from reference toxicant (phenol) tests showed EC50 values ranging from 19.98 to 32.83%, averaging 24.78%. Similarly, tests of DMSO blanks showed no toxicity for each of the tests. Tests of the North Inlet reference sediment showed EC50s of 1.46, 3.02, 0.84, 0.82, and 0.77 mg/mL, indicating a relatively consistent response among test batches.

Summarized results of the toxicity tests are listed in Table 2 for Charleston Harbor, Winyah Bay, and Leadenwah Creek. The numerical results are accompanied by indications of statistical significance: “ns” for non-toxic ($p>0.05$); a single asterisk for statistical significance ($p<0.05$ only); and a double asterisk for highly significant ($p<0.05$ and $>20\%$ difference from control). In the Microtox™ tests, three asterisks were shown when results were significant in Mann-Whitney, Dunnett’s, and distribution-free analyses. Also, a “toxicity tally” is shown which is the summation of the levels of significance observed in each toxicity test, i.e. the number of asterisks assigned to each result. The mean ERM quotients, indicators of the overall degree of chemical contamination for the samples, are listed for those samples analyzed for chemical concentrations. This index was based upon the average of 25 chemical concentrations divided by their 25 respective ERM values (from Long et al., 1995).

None of the samples from Charleston Harbor and vicinity caused a significant decrease in amphipod survival relative to the controls (Table 2). In contrast both the Microtox™ bioluminescence and sea urchin fertilization tests indicated toxicity in numerous samples. In some samples, such as those from stations B5-1 and B6-2 in Winyah Bay, these two bioassays showed good consensus regarding the toxicity of the samples (toxicity tallies of 9 and 8, respectively). In most of the samples that were toxic in the Microtox™ tests, they were also toxic in the urchin fertilization tests, but these two tests did not show good agreement in many other samples. Only two samples from Leadenwah Creek were toxic in any of the tests and the test results were only slightly different from the controls. There was a much wider range in response among the Winyah Bay samples: one sample was non-toxic in all tests and two others showed very high toxicity. The sample from station B4-1 was unusually toxic and also had a relatively high mean ERM quotient (0.147).

Two samples from the Ashley River had the highest mean ERM quotients (>0.2), indicating the highest concentrations of mixtures of substances; however, neither of these samples was very toxic. However, most of the samples from the Ashley River were toxic in at least one of the bioassays, especially those performed with sea urchins. In the Cooper River, all except the sample from station H1-1 were slightly to moderately toxic, as indicated by toxicity tallies of 0 to 4, and most had low to moderate chemical concentrations (mean ERM quotients of 0.01 to 0.18). The six samples from the lower Wando River had similarly low to moderate toxicity and chemical concentrations. Among the eleven samples tested for the Charleston Harbor Project, five had moderate to high toxicity (toxicity tallies of 6-8), and all had low to moderate chemical concentrations (mean ERM quotients of 0.01 to 0.17) similar to those collected elsewhere in Charleston Harbor.

Responses in the cytochrome P-450 RGS bioassays performed on selected samples from Charleston Harbor and Winyah Bay ranged from 1.8 to 86.3 ug/g benzo(a)pyrene equivalents (Table 3). Induction was considerably higher in the samples from the Ashley River (D strata) than in those from the Cooper River (H strata). Induction was highest in two adjacent samples (D3-3 and D4-1) from the Ashley River and in a sample (G2-1) from Shipyard Creek; intermediate in several samples from upper Winyah Bay and the Ashley and Cooper rivers; and lowest in several samples from the lower reaches of Winyah Bay.

In the meiobenthic copepod bioassays of reproduction, copepodite stage production and naupliar stage production were significantly reduced in one sample collected at station CHP 4 (Table 3). However, total copepodite+naupliar production was not significantly reduced. Clutch size was significantly reduced in one sample from Leadenwah Creek and seven samples from Charleston Harbor; responses ranged from 8.8 to 100.7.

Toxicity and chemistry data for the Savannah River are summarized in Table 4. The sample from station A1-1 was extremely toxic in the amphipod test, but non-toxic in all other tests and had very low chemical concentrations. Three other samples from strata A1 and A2 in the upper-most stretch of the study area were toxic in the amphipod tests, two were toxic in the Microtox™ tests, and only one was toxic in the sea urchin development test of 100% porewater. In the B strata adjacent to downtown Savannah, results of the toxicity tests showed a wide range in response; toxicity tallies ranged from 0 to 12. Toxicity tallies were 10-12 in seven of the samples. However, the mean ERM quotients indicated slight to moderate chemical concentrations (ranging from 0.01 to 0.14). One sample each from the Back River and lower Savannah River strata was toxic in amphipod tests and none from the south channel was toxic in that test. One sample from the lower Savannah River and three samples from the three Savannah harbor strata were toxic in Microtox™, urchin fertilization, and urchin development tests (toxicity tallies of 10-11). All seven samples from the Savannah harbors had similar chemical concentrations (mean ERM quotients of 0.09 - 0.14).

In the St. Simons Sound study area, two samples (both from Terry Creek) were toxic in the amphipod tests (Table 5). Amphipod survival was 0.0% in the sample from station F2-1, a very unusual result for this relatively resistant test. Also, the chemical concentrations were relatively high in this sample (mean ERM quotient of 0.22). Elsewhere in this estuary, two samples from the Brunswick Harbor/East River area showed relatively high toxicity (toxicity tallies of 10). The sample from station C2-1 was toxic in Microtox™, urchin fertilization, and urchin development tests. Another sample from this region (station B2-1) had relatively high chemical concentrations (mean ERM quotient of 0.31). The sample from the Purvis Creek stratum was the most contaminated (mean ERM quotient of 0.47), but it was not toxic in any of the tests. Overall, the highest chemical concentrations observed in the entire study area were encountered in samples from stations B2-1 and P1 in the St. Simons Sound estuary.

The incidence of test results that were statistically significant ($p<0.05$) is summarized in Table 6. Data from the copepod reproduction and urchin embryo development tests are not included since they were not performed with all samples. Toxicity was most prevalent among the Winyah Bay samples in which 6 of 9 and 8 of 9 samples were toxic in the tests performed with Microtox™ and urchin fertilization in 100% porewater, respectively. Among all 45 tests performed with Winyah Bay samples, a total of 21 (46.7%) results showed significant differences from controls. Many of the samples from Charleston Harbor were significantly toxic,

especially in the Microtox™ and urchin fertilization tests, the two most sensitive assays. In Charleston Harbor, a total of 97 (30%) test results of 325 bioassays performed showed significant differences from controls. Much smaller proportions of samples from the Savannah River and St. Simons Sound were toxic (21% and 12%, respectively). Ten of the samples from the Savannah River were toxic in the least sensitive test, the amphipod survival bioassay. The relatively high incidence of toxicity in the urchin fertilization tests performed with 100% Savannah River porewater diminished markedly with dilution of the samples - none of the samples were toxic in tests of 25% porewater. Finally, the samples from Leadenwah Creek had the lowest incidence of toxicity; none of the samples was toxic in the amphipod survival or urchin fertilization tests performed with either 100% or 50% porewater. Unexpectedly, however, two samples were toxic in tests of 25% porewater.

Spatial distribution of toxicity in Charleston Harbor. None of the 65 samples collected in Charleston Harbor and North Inlet from either the NOAA stations or the Charleston Harbor Project stations were toxic ($p>0.05$) in the amphipod tests. Therefore, no distributional map is provided.

Microtox™ test results were analyzed with three statistical analyses, representing increasing levels of conservativeness: Mann-Whitney, Dunnett's, and distribution-free. Samples in which average results were not significantly different from the North Inlet reference samples in the least conservative analysis (Mann-Whitney) are shown as open circles in Figure 70. Those determined to be significantly different from the reference samples in the three statistical analyses are shown with different symbols. In Charleston Harbor none of the samples was significant in the distribution-free analysis. However, test results for the samples from stations CHP 10 in the Wando River, CHP -3 and E2-1 in Charleston Harbor, H1-1 and H7-1 in the lower Cooper River, and CHP 7 and D3-1 in the Ashley River were significant in both the Mann-Whitney and Dunnett's analyses. In addition, eight of the samples from the Cooper River showed significant results in only the Mann-Whitney analyses. Only one of the samples from the lower Ashley River and lower harbor were toxic in this test.

The sea urchin fertilization tests were performed in three porewater concentrations (100%, 50%, and 25%) and test results for each dilution were compared with those from the controls to determine significant differences (Figure 71). Samples with the highest toxicity were those that cause significant results in all three porewater concentrations. Toxicity in the 100% porewater tests was pervasive throughout Charleston Harbor; at least some samples collected in all three rivers were toxic to urchin fertilization. Many of the most toxic samples came from the lower Ashley River and the region of the lower harbor that receives the Ashley River. Most samples from Shipyard Creek (G stations) and the lower Cooper River in the vicinity of Shipyard Creek caused toxicity in 50% or 25% porewater tests. The most seaward stations, C1 and C2, were not toxic in these tests.

Cytochrome P-450 RGS assays were performed on selected Charleston Harbor and Winyah Bay samples. Data are shown for the 20 Charleston Harbor samples as benzo(a)pyrene equivalents (ug/g) of fold-inducing substances in solvent extracts of the sediments. In Charleston Harbor, most of the samples tested were from the Ashley River where petroleum-like sheens were often observed in the samples. Station B5-2 from lower Winyah Bay (not shown) produced a baseline level of response (2 ug B(a)P equivalents/g), comparable to responses at

other clean sites. Sediments from two Ashley River stations (D3 and D4) and one Shipyard Creek station (G2) produced the highest levels of induction (70-86 ug/g) in this assay. Sediments with levels of B[a]PEq above ug/g exhibited degraded benthic communities in a survey of sediment quality in San Diego Bay (Fairey et al., 1996). Stations in the upper Cooper River and lower Winyah Bay provided the lowest responses in these tests.

Twelve samples were tested for adult survival and reproductive success among meiobenthic copepods (Figure 73). None of the survival rates were significantly lower than North Inlet reference samples. Only three reproductive end-points showed significant results relative to reference samples: naupliar production, copepodite production, and clutch size. Clutch size was significantly depressed in sediments from seven stations, including all samples from the lower Ashley River and all except one of the samples from the lower harbor/Charleston Harbor. Sediment from station CHP 4 near the mouth of the Ashley River caused significant responses in all three end-points.

In summary there were many samples from Charleston Harbor that were toxic in one or more of the bioassays; however, none were toxic in the least sensitive bioassay performed - the amphipod survival test. Samples indicated as toxic in one or more tests were scattered throughout the study area. No single region was remarkably toxic in all tests and there was relatively little concordance among the tests. These data suggest, therefore, that toxic samples were scattered throughout the area, not concentrated in any particular area, and, as judged by the amphipod tests, toxicity was not severe. The Microtox™ tests indicated many samples from the Cooper River were toxic and only two samples were toxic from the Ashley River, whereas the urchin fertilization and copepod reproduction tests indicated that most Ashley River samples were toxic. However, both the Microtox™ and urchin fertilization tests showed toxicity in some of the same stations; notably, CHP 7, CHP 10, D3-1, G1, H5-4, H8-1, and H6-1. The station most toxic to copepod reproduction, CHP 4, was also toxic in the urchin fertilization test, but not in the Microtox™ test. Several of the most seaward stations in the lower harbor were consistently non-toxic in the tests performed, suggesting that toxic conditions did not extend beyond the harbor entrance.

Spatial distribution of toxicity in Leadenwah Creek. None of the 9 samples collected in Leadenwah Creek were toxic in the amphipod tests. Therefore, no distributional map is provided. Cytochrome P-450 assays were not performed on Leadenwah Creek samples. Copepod reproduction tests were performed on one sample from Leadenwah Creek, station A1-8, and significantly reduced clutch size was observed in that sample.

Results of the Microtox™ tests are summarized in Figure 74. The sample from one station, A1-1, in the upper reaches of the creek showed significant results in both the Mann-Whitney and Dunnett's analyses. In the urchin fertilization tests, none of the samples of 100% porewater or 50% porewater were toxic. However, two samples both from upper reaches of the creek showed slight toxicity in the tests of 25% porewater only (Figure 75).

In summary, the samples from Leadenwah Creek, a system that has historically received considerable pesticide runoff from agriculture, were not highly toxic. Only one sample showed toxicity in the Microtox™ tests, none were toxic in the amphipod survival tests, and a very slight decrease in urchin fertilization was observed in the porewaters of two samples.

Spatial distribution of toxicity in Winyah Bay. None of the nine samples collected in Winyah Bay were toxic in the amphipod survival tests. Copepod reproduction tests were performed on one sample (station B 3-1) from the lower Sampit River and no significant effects were observed.

In the Microtox™ tests, six of the nine samples were significantly different from controls in at least the Mann-Whitney analysis (Figure 76). Samples from stations B1-3 and B1-2 in the Georgetown harbor area, B4-1 in the lower Sampit River, and B5-1 in the upper Winyah Bay were the most toxic. A similar pattern was observed with the results of the urchin fertilization tests (Figure 77). The sample from station B3-1 was not toxic and the samples from stations B1-2, B4-1 and B5-1 were the most toxic. The remaining samples were toxic only in the tests of 100% porewater.

In the cytochrome P-450 RGS assays, fold induction was low in the sample from station B3-1 which was not toxic in the Microtox™ and urchin fertilization tests (Figure 78). However, fold induction also was very low in the samples from stations B4-1 and B5-1 which were toxic in the other bioassays. Assay results were highest in stations B1-3 and B7-1.

In summary, eight of the nine samples from Winyah Bay showed toxicity in at least one of the bioassays and toxicity was most severe in the Georgetown harbor area. Toxicity generally decreased seaward down the bay, however, the most seaward station was toxic in two tests and showed the highest fold induction rate in the cytochrome P-450 assay. Therefore, the seaward extent of toxicity was not determined.

Spatial distribution of toxicity in St. Simons Sound. In amphipod survival tests two of the 20 samples from St. Simons Sound were significantly different from controls (Figure 79). Both samples (from stations F1-1 and F2-1) were collected in Terry Creek, a tributary to the Back River northeast of the city of Brunswick. The remaining 18 samples tested were not toxic in this test. Similarly, the Microtox™ tests indicated that samples from stations F1-1 and F2-1 were highly toxic (Figure 80). In addition, samples from station C2-1, E2-2, and G1-1 were highly toxic in this test. As in the amphipod tests, samples from the Turtle River and St. Simons Sound were not toxic in the Microtox™ tests.

The urchin embryological development tests also indicated that samples from stations F1-1 and F2-1 were toxic (Figure 81). However, in this test, four of the samples (B1-1, C1-1, C2-1, and C3-1) from Brunswick harbor were the most toxic, with significant effects at all three porewater concentrations. Some samples from the Turtle River were toxic in tests of 100% or 50% porewater, but toxicity diminished downstream and samples from lower St. Simons Sound were not toxic in this test.

In contrast to the results of the other tests, the urchin fertilization test did not indicate that samples from stations F1-1 and F2-1 were toxic (Figure 82). However, as indicated in the urchin embryological development tests, most samples from the Brunswick harbor were toxic. Also, two samples from the upper reaches of the Turtle River were toxic. None of the samples from the lower Turtle River, lower Back River, or lower St. Simons Sound were toxic in this test.

In summary, toxicity was most severe in the two samples collected in Terry Creek; all except the urchin fertilization tests showed relatively severe toxicity there; and average amphipod survival was 0.0% in one of the samples. Samples from the Brunswick harbor area were toxic in several bioassays, notably the urchin tests. Samples from some stations in the Turtle River were toxic in only the urchin tests. The samples from the lower reaches of the Turtle River and St. Simons Sound were consistently non-toxic, indicating, as in Charleston Harbor, that toxic conditions did not extend beyond the entrance to the estuary.

Spatial distribution of toxicity in upper Savannah River. In amphipod survival tests only five samples were significantly different from LIS controls; three samples from the uppermost reach (stratum A) of the river upstream of the city of Savannah and one sample from Back River (station C-1) below the city of Savannah (Figure 83). Mean amphipod survival in the sample from station A1-1 was 4.0% of the control, an unusual result for this test. All other samples were non-toxic in this test.

In the Microtox™ tests, considerably more samples were different from controls than in the amphipod tests, including many samples collected near the city of Savannah (Figure 84). Of the 43 samples tested, 23 were toxic as determined with Mann-Whitney analysis. Toxicity in this test was most severe among samples taken in strata B4, B5, B7, B9, and H and least severe among the samples from strata B1 and B8. Two of samples from the upper most reaches of the river - strata A1 and A2 - were toxic.

Similar to the Microtox™ tests, the urchin embryological development bioassays showed a high incidence of toxicity in this area (Figure 85). In these tests 31 of 43 samples were significantly different from controls in at least 100% porewater. Samples with the most severe toxicity were collected from strata B2, B3, B5, B9, G and H. Many of the samples from the uppermost reaches of the river (strata A1, A2 and B1) were among the least toxic.

In the urchin fertilization tests, 18 of 43 samples were toxic in at least 100% porewater (Figure 86). Samples showing highest toxicity were clustered around the middle of the region within strata B3, B9, and F. Many of the samples from this central cluster also were toxic in the embryological development bioassays. Most of the samples from the upper reaches of the river and from the downstream strata were non-toxic.

In summary, stations in which toxicity was observed were scattered throughout the region and each bioassay showed somewhat different patterns in toxicity. However, three tests - the Microtox™ tests and both urchin tests - indicated that samples from the central portion of the region were toxic. Specifically, all except two samples collected within strata B3, B4, B9 and F were toxic in these three tests. With some major exceptions, most samples collected upstream of the city of Savannah were non-toxic in these three tests. Also, there was generally less toxicity downstream of the city and in Back River. Data from the amphipod survival tests, the least sensitive bioassay performed, showed a different pattern in toxicity; the only toxic stations were those from the upper reaches of the river and from Back River.

Spatial distribution of toxicity in lower Savannah River. Seventeen samples were collected below Fort Johnson. In amphipod survival tests only one sample (from station D3,1-4) was toxic (Figure 87). This sample was collected for a location downstream of the city of Savannah where most pollutant sources would be expected.

In the Microtox™ tests, the sample from station D3,1-4 was not toxic; however, several other samples collected downstream were toxic (Figure 88). Toxicity was most severe in samples from station D3,3-2, station E3,5, and station E3,9. Samples from strata E1, E2, and D2 were among the least toxic. Toxicity did not disappear at the most seaward stations near the mouth of the river, suggesting that the seaward extent of toxicity was not encountered.

Samples from three stations - D1,3-1 and E3,5 and D3,3-2 - which were toxic in the Microtox™ tests also were highly toxic in the urchin embryological development bioassays (Figure 89). Many of the stations within strata D2, E1 and E2 that were non-toxic in the Microtox™ tests also were non-toxic in this test. As in the amphipod tests, only one sample was toxic in the urchin fertilization tests (Figure 90). In this case, however, the toxic sample was collected from station D1, 3-1 near Elbe Island. Station D1,3-1 was toxic in both urchin tests and in the Microtox™ tests.

In summary, toxicity was scattered among the stations and no clear spatial pattern in toxicity was evident. Two samples - one from the upper reach of this region and another collected near the mouth of the river - showed toxicity in three of the four tests. Overall, based upon the results of all tests, the samples from the lower Savannah River/south channel stations were considerably less toxic than those from the upper Savannah River.

Concordance Among Toxicity Tests. Spearman-rank correlations were calculated to estimate the degree of concordance among the different toxicity tests. Concordance was determined for each region (Charleston Harbor and vicinity, Savannah River, St. Simons Sound) and for the entire study area. Positive correlation coefficients (Table 7) indicated the results of the tests showed similar spatial trends in toxicity, whereas negative coefficients indicated different patterns.

In Charleston Harbor, the only significant positive correlation was between results of the Microtox™ and urchin fertilization tests ($\text{Rho} = +0.389$, $p < 0.001$). All of the remaining correlations were non-significant. In the Savannah River samples, the strongest positive correlation was, as expected, between urchin fertilization and urchin development. Also, the results of the Microtox™ tests were correlated with both urchin tests. In contrast, amphipod test results showed negative correlations with both of the urchin tests. Similarly, in St. Simons Sound results of the Microtox™ tests were positively correlated with urchin fertilization, whereas amphipod survival was negatively correlated with urchin fertilization. As in the Savannah River, urchin fertilization and development were strongly correlated with each other in the St. Simons Sound samples.

When the results of these tests from all regions were combined, the patterns observed in Savannah River and St. Simons were also apparent. Urchin fertilization and development were strongly correlated with each other; both urchin tests were positively correlated with Microtox™ results; and both urchin tests were negatively correlated with amphipod survival. These results suggest that the results of both urchin tests and the Microtox™ tests followed overlapping and similar, but not duplicative, patterns whereas a different spatial pattern in toxicity was indicated in the amphipod survival tests.

Spatial Extent of Toxicity. The spatial extent of toxicity was estimated for each estuary based upon toxicity test data from the three bioassays (urchin fertilization, amphipod survival, Microtox™) that were performed throughout the entire area (Table 8). The critical value used in these calculations was <80% of controls; that is samples in which test results were less than the critical value assigned to the “toxic” category. The sizes of the strata weighted to the number of samples within the stratum that were “toxic” were summed to provide an overall estimate for each estuary. Also, the estimates for each estuary were summed for the entire study area.

The total study area encompassed approximately 88 km², including 41 km² in Charleston Harbor, the largest estuary studied (Table 8). In the amphipod survival tests, “toxic” samples, as defined by the critical value, were observed only in samples from the Savannah River and St. Simons Sound. The spatial extent of toxicity in amphipod survival tests in these two estuaries was approximately 0.2 and 0.1 km², equivalent to 0.3% of the total study area. In the urchin fertilization tests, the spatial extent of toxicity was approximately 18.7 km² (21.3% of the total area), most of which (12.5 km²) occurred in Charleston Harbor. In Winyah Bay, toxic samples in the urchin fertilization tests represented about 3.1 km² (42.2% of this small estuary).

The estimated spatial extent of toxicity was highest in the Microtox™ tests (48% of the total area, Table 8). In these tests, toxicity was most pervasive in Winyah Bay and Savannah River, 70% and 57% of these areas, respectively. Approximately 43% and 46% of Charleston Harbor and St. Simons Sound, respectively, were toxic in these tests. Only one sample, representing 20% of the area, was toxic from Leadenwah Creek.

Correlations between Toxicity and Chemical Concentrations. Simple, Spearman-rank correlations between measures of toxicity and the concentrations of chemical substances in the samples were determined to identify and quantify concordance. Correlations were determined for the 66 samples from Charleston Harbor/Winyah Bay/Leadenwah Creek, 61 samples from Savannah River, and 20 samples from St. Simons Sound in which chemical concentrations were determined (Appendices B1-B3). Correlations do not identify or imply cause-effect relationships; rather, they simply identify the degree, if any, of concordance between independent variables (chemistry) and dependent variables (toxicity) in the same samples. Correlations were determined for individual chemicals and classes of chemicals. In addition, correlations were determined with several cumulative chemical indices, which were calculated by dividing concentrations by their respective ERM values (Long et al., 1995) and summing the quotients.

Correlations were expressed as coefficients (rho, corrected for ties) with either a positive or negative sign. A negative sign would suggest that, for example, survival decreased as a chemical concentration increased. Correlations were identified as non-significant (ns, p>0.05), and significant (p<0.05*, p<0.001**, or p<0.0001***). However, correlations performed with a sufficiently large number of variables can be significant by random chance alone. In these analyses correlations identified as ** (p<0.001) and *** (p<0.0001) would remain significant if the number of variables were taken into account, whereas those identified as * (p<0.05) would not remain significant.

In Charleston Harbor and vicinity (including North Inlet, Winyah Bay, and Leadenwah Creek), 66 samples were tested for toxicity and analyzed for chemical substances (Table 9). None of the samples tested for amphipod survival showed significant effects. Therefore, with no significant gradient in response, none of the correlations with chemical substances were significant. All of the coefficients listed in Table 8 had positive signs except for those with un-ionized ammonia and results of the P-450 RGS assays.

In contrast to the amphipod tests, the microbial bioluminescence (MicrotoxTM) tests showed significant correlations with many different substances in Chareston Harbor and vicinity (Table 9). The large number of significant correlations indicated that many substances were correlated with each other. Among the trace metals, all except silver, cadmium, and mercury showed significant negative correlations with bioluminescence. The concentrations of copper showed the strongest correlations among the trace metals. However, the ratios between total SEM concentrations and AVS concentrations were positively correlated with bioluminescence, suggesting light production increased with increasing metals concentrations normalized to respective AVS concentrations. Bioluminescence significantly decreased with increasing percent fines and percent total organic carbon, indicating fine-grained sediments had the highest toxicity, probably due to the presence of the highest chemical concentrations. With negative correlation coefficients of 0.26 to 0.35, all classes of organic compounds were similarly correlated with MicrotoxTM test results. Again, these data suggest that organic compounds covaried with each other in mixtures.

MicrotoxTM test results were significantly correlated with all chemical concentration/ERM ratios in Charleston Harbor (Table 9), including those for all 25 substances for which an ERM was prepared (Long et al., 1995b). These data, again, suggest that bioluminescence decreased as the concentrations of mixtures of substances normalized to their respective ERM values increased. Furthermore, the correlations among all classes of organics (total PAH, total PCBs, total DDTs, total pesticides) were significant. For example, the concentrations of total PAH and total PCB were highly correlated ($\rho = 0.661$, $p < 0.0001$). Similarly, total PCBs and total pesticides ($\rho = 0.492$, $p < 0.0001$) and total PCBs and total DDTs ($\rho = 0.485$, $p < 0.0001$) were highly correlated with each other. Consistent with the correlations for individual trace metals, the correlation between bioluminescence and the sum of the 9 trace metals/ERM quotients was strongest.

Correlations between urchin fertilization and chemical concentrations in the Charleston Harbor samples were significant only for silver and the sums of total PAHs, total PCB congeners, and sums of non-DDT pesticides (Table 9). The correlation with silver was strongest but would not remain so if the number of variables were taken into account.

Cytochrome P-450 RGS bioassays were performed on solvent extracts of 29 of the Charleston Harbor and Winyah Bay samples. These tests have been shown in laboratory trials to respond to a number of different mixed-function oxidase inducing chemicals, notably dioxins, and to a lesser extent PAHs and some PCB congeners. Among the 29 samples chosen from Charleston, there was a wide range in response (2 to 86 ug/g) in the test results. As expected, none of the correlations were significant with any trace metals. However, test results correlated very strongly with the concentrations of a number of organic substances that would be expected to induce responses (Table 9). Spearman-rank correlations were highest for the high-molecular weight PAHs and PCBs, whether or not they were normalized to the ERM

concentrations. These data suggest that, as expected, the P-450 results co-varied with complex mixtures of many different organic substances in the sediments. The 4- and 6-ring PAHs, which are the primary compounds associated with the Ah-receptor, would be particularly important in triggering these bioassay responses.

A series of copepod reproduction bioassays was performed with 19 of the Charleston Harbor samples. Total copepodite/naupliar production and total clutch size were not significantly correlated with any of the measured chemicals in the samples. A few trace metals indicated weak correlation coefficients, but none were significant (Spearman-rank, $p < 0.05$, $n = 19$), especially if the numbers of variables were accounted for.

In the Savannah River, matching toxicity and chemistry data were developed for 61 samples (Table 10). As observed in Charleston Harbor and vicinity, none of the substances measured showed a significant negative correlation with amphipod survival, largely due to the relatively low sensitivity of this bioassay. Chemical concentrations in the station in which amphipod survival was 0.0% were not very high. Only the concentrations of cadmium, total SEM and the SEM:AVS ratios were significantly correlated with Microtox™ test results.

In sharp contrast to the amphipod and Microtox™ test results, both of the urchin tests performed on Savannah River samples showed strong negative correlations with many different substances (Table 10). These correlations were particularly strong for ammonia, zinc, total SEM, total metals/ERM quotients, total PAHs, and total high molecular weight PAHs. Among the highest correlations was that with sum of the 24 chemicals/ERMs quotients. The highly significant correlations with sums of chemical concentrations normalized to respective ERM values suggested that, as observed in Charleston Harbor, bioassay results co-varied with mixtures of numerous substances. Furthermore, the correlations between all classes of organic compounds (PAHs, PCBs, DDTs, pesticides) were highly significant ($\rho = 0.40$ to 0.69 , $p < 0.001$ or $p < 0.0001$).

In St. Simons Sound, matching toxicity and chemistry data were available for 20 samples (Table 11). Results of Microtox™ tests were highly correlated with aluminum, iron, and nickel none of which are highly toxic substances. Test results also were correlated with the sum of 5 SEM concentrations, but not with the SEM:AVS ratios. Relatively strong associations were apparent between microbial bioluminescence and both the concentrations of PAHs and the sums of the 24 concentrations/ERMs quotients. Results of the urchin fertilization tests were highly correlated with many trace metals concentrations, but not with the concentrations of PCBs, DDTs, and pesticides. Urchin embryological development was correlated with many of these same substances, but the coefficients and significance of the correlations were weaker than seen with the fertilization tests. In both bioassays the concentrations of un-ionized ammonia were associated with test results.

Toxicity and chemistry data from all 147 samples were compiled to determine if any correlations with classes of substances observed in individual regions remained significant over the entire study area (Table 12). Correlations were determined only for summed chemical classes. Many correlations remained significant and some strengthened in this combined data set. The correlations between ammonia and both urchin tests were highly significant. Also, there was a very strong association between the cumulative ERM quotients and toxicity in the

Microtox™ and both urchin tests. As noted above in each individual region, it was apparent that mixtures of substances co-varying with each other were strongly associated with toxicity in these three bioassays.

Comparisons of Trace Metals Concentrations to Reference. Trace metals concentrations in sediments collected from reference areas in the southeastern USA have been observed to covary with average grain size in the sediments, which, in turn, co-varies with aluminum content (Schropp et al., 1990; Schropp and Windom, 1988; Windom et al., 1989). By comparing trace metals concentrations to aluminum concentrations in clean samples, these investigators determined the metals-to-aluminum ratios to be expected in clean reference areas and the 95% confidence limits of these ratios. Exceedances of the upper 95% confidence limits are assumed, by this technique, to represent polluted conditions in which there was an excess of a trace metal.

Trace metals analyses in this survey were performed with nitric acid digestions, which would be expected to extract less of the clay-bound aluminum in the sediments than in hydrofluoric acid digestions. The metals-aluminum ratio tool is based upon total (hydrofluoric acid) digestions (Schropp et al., 1990). Therefore, the following metals-aluminum plots may exaggerate the concentrations of metals relative to the concentrations of aluminum (perhaps by a factor of roughly two).

Figures 91-96 illustrate the relationships between aluminum concentrations in the sediments from this study and the concentrations of arsenic, cadmium, copper, chromium, lead, and zinc in the samples. The lower and upper 95% confidence intervals for reference areas from Schropp et al. (1990) are included on the figures for each metal. Also shown are the Spearman-rank correlation coefficients for the data from this study. The concentrations of all trace metals measured except silver were highly correlated ($p < 0.0001$) with the concentrations of aluminum; the coefficients ranged from 0.449 for cadmium to 0.962 for iron. Along with iron, the concentrations of nickel, chromium, tin, and zinc were very highly correlated with aluminum (all correlation coefficients > 0.9).

The concentrations of arsenic, although highly correlated with aluminum, were within the 95% confidence intervals expected for reference sediments (Figure 91). In contrast, the concentrations of cadmium, copper, chromium, lead, and zinc in some samples exceeded the expected concentrations. In approximately 30 samples the concentrations of cadmium exceeded the concentrations expected from the aluminum content (Figure 92). Similarly, many samples with relatively high aluminum concentrations had excess levels of chromium and copper (Figures 93, 94). Among these six trace metals the concentrations of lead and zinc were most elevated most frequently (Figures 95, 96).

These data suggest that, based upon the metal-to-aluminum ratios, some samples from the study area had excess trace metals concentrations, ostensibly due to anthropogenic sources. As exemplified with data from the lower Miami River, Schropp et al. (1990) interpret excess metals concentrations as accumulations of these elements from anthropogenic sources on fine-grained particles. These observations, if they correctly portray metals-aluminum ratios (see cautionary note above), would suggest that some portions of the trace metals concentrations in the South Carolina/Georgia estuaries were contributed by human sources.

Comparisons of toxicant concentrations with sediment guidelines. The concentrations of all 27 substances for which numerical guidelines (Long et al., 1995b) exist were compared to the guideline values to identify which toxicants may have been elevated (Table 13). Although silver showed a strong correlation with urchin fertilization in Charleston Harbor (Table 9) and St. Simons Sound (Table 11), none of the silver concentrations exceeded the ERL concentration of 1.0 ppm. Arsenic exceeded the ERL value of 8.2 ppm in 64 samples but not the ERM value of 70 ppm. The concentrations of cadmium, chromium, copper, lead, and zinc exceeded their respective ERL values in 3-9 samples. The concentrations of mercury and nickel were elevated in 12 and 33 samples, respectively, relative to the ERL values, but Long et al. (1995b) reported relatively low confidence in these values. The mercury concentration in one sample from the lower Cooper River (station H1-4) exceeded the ERM value of 0.71 ppm.

The concentrations of all individual PAHs were elevated relative to respective ERL values in 4 to 44 samples (Table 13). In addition, the concentrations of anthracene, pyrene, and the sum of low molecular weight PAHs exceeded their respective ERM values in one or two samples (station D2-1 in the lower Ashley River, station B2-1 in the Brunswick Harbor, and station F2-1 in Terry Creek). Chlorinated hydrocarbon (PCBs, DDTs) were elevated in numerous samples relative to ERL values, but did not exceed the ERM values in any samples.

Based upon these data, it appears that many samples were not highly contaminated and a few were moderately contaminated, and none were highly contaminated. Samples in which all chemical concentrations were lower than the respective ERL values rarely would be expected to cause toxicity (i.e., <13% of samples in amphipod survival tests, Long et al., in prep.). Samples in which toxicant concentrations exceeded 1 to 10 ERL values, but were lower than all the ERM values, may be toxic in some cases, but not frequently (10-30% of samples in amphipod survival tests; Long et al., in prep.). Samples in which only one or two chemicals exceeded the ERM values would be expected to cause toxicity in only 30-40% of samples in amphipod survival tests (Long et al., in prep).

Therefore, although some trace metals were elevated in concentrations relative to both the aluminum content of the sediments and the ERL values, only one of these concentrations was above an ERM value. Also, only one or two samples had very high concentrations of PAHs and none had high concentrations of chlorinated substances. None of the chemical concentrations exceeded any of the five proposed national sediment quality criteria (U. S. EPA, 1994). As a consequence, a high degree of toxicity would not be expected in these samples, and, indeed, only a few showed toxicity in the amphipod survival tests. However, the chemical concentrations exceeded numerous ERL values and could be expected to cause toxicity in highly sensitive bioassays, such as the urchin fertilization and Microtox™ tests, and, indeed, these tests frequently showed toxicity in many samples.

Relationships between toxicity and concentrations of selected toxicants. The relationships between measures of toxicity and the concentrations of some substances were examined further with scatterplots when the correlations were significant and respective guideline values were exceeded. The scatterplots were prepared to provide visual evidence that some substances ostensibly indicated, as correlated with toxicity, a reasonable pattern of association and were elevated in concentration in the most toxic samples. Most of the scatterplots were prepared for the Microtox™ and sea urchin test results, since only two samples showed high

toxicity (one in the Savannah River and one in St. Simons Sound) in the amphipod tests, and, consequently, most chemicals showed weak correlations with amphipod survival.

The concentrations of total ammonia and un-ionized ammonia were determined for the water overlying the sediments in the amphipod exposure chambers. The two samples in which amphipod survival was lowest (0.0% and 4.0% survival) had relatively low concentrations of un-ionized ammonia - the most toxic form of ammonia (Figure 97). The Spearman-rank correlation was significant, but indicated a positive association, whereas a negative association would be expected if ammonia were a major contributor to toxicity. Five of the samples tested had un-ionized ammonia concentrations that exceeded the No Observable Effects Concentration for *Ampelisca abdita* (approximately 0.236 mg/L (from Kohn et al., 1994); none exceeded the LC50 concentration 0.83 mg/L.

In the sea urchin fertilization tests, ammonia concentrations were measured in the porewaters used in the toxicity tests. The Spearman-rank correlation between fertilization success and un-ionized ammonia was highly significant and the scatterplot shows a relatively consistent pattern of decreasing fertilization success with increasing ammonia concentrations (Figure 98). However, many samples with very low levels of ammonia were highly toxic, and only one sample exceeded the Lowest Observed Effects Concentration (LOEC) of 800 ug/L for this bioassay test; indicating other factors must have caused or contributed to toxicity. Therefore, it appears that no more than one of the 147 samples had concentrations of un-ionized ammonia sufficiently high to contribute substantially to toxicity in this test.

In sharp contrast, the sea urchin development bioassay is known to be highly sensitive to the presence of ammonia in the porewater; the LOEC concentration is 90 ug/L. Results of the embryo development tests were highly variable among samples with un-ionized ammonia concentrations less than the LOEC. However, as ammonia concentrations increased above the LOEC, percent normal development diminished rapidly. Many of the samples had un-ionized ammonia concentrations greater than 90 ug/L and all of those samples were highly toxic to embryo development (Figure 99).

To account for the possible additive effects of numerous chemicals occurring in mixtures, mean ERM quotients were calculated as the average of 25 chemical concentrations divided by their respective ERM values. As indicated in Table 9, the correlations between the results of the Microtox™ and two sea urchin tests and the mean ERM quotients were significant for the combined study area data set ($n = 147$). In the Microtox™ tests a relatively consistent pattern of decreasing light production with increasing mean ERM quotients was apparent (Figure 100). Several of the samples with highest mean ERM quotients (>0.2) were highly toxic; however, many more samples with considerably lower concentrations of these mixtures also were highly toxic and the two samples with the highest concentrations were not toxic. In addition, the correlations between microbial bioluminescence and mean ERM quotients were relatively weak, although significant, in the Charleston Harbor and St. Simons Sound samples and non-significant in the Savannah River samples (Tables 9, 10 and 11). Therefore, this relationship between microbial bioluminescence and the concentrations of complex chemical mixtures showed relatively high variability and inconsistency.

Similarly, the relationship between the mean ERM quotients and sea urchin fertilization was relatively inconsistent (Figure 101), despite the observations of significant correlations in

samples from the Savannah River and St. Simons Sound (Tables 10 and 11). Although, a large number of samples with high fertilization success and low chemical concentrations clustered together, there was no strong and consistent pattern of decreasing fertilization with increasing chemical concentrations. The two samples with highest chemical concentrations were among the least toxic in this test.

The strongest associations between the mean ERM quotients and toxicity were observed in the sea urchin embryo development tests (Figure 102) and the cytochrome P-450 RGS assays (Figure 103). There was a large cluster of samples with low chemical concentrations and high percent normal embryo development and another large cluster of samples with somewhat higher chemical concentrations and very low percent normal development. The correlations were significant for the samples from the Savannah River and all samples combined, but not for the St. Simons Sound samples, where one outlier (station P1) with the highest concentrations was not toxic. Results of the cytochrome P-450 RGS assays showed a strong pattern of increasing induction with increasing concentrations of total PAHs (Figure 104). This is a pattern that was expected and observed in previous studies (Fairey et al., 1996), especially because high molecular weight PAHs are strong inducers of the assay response.

The correlation between the concentrations of copper and results of the Microtox™ bioassays were highly significant in the Charleston Harbor area (Table 6). The scatterplot indicated a consistent pattern of decreasing light production with increasing copper concentrations (Figure 105). All five samples with copper concentrations greater than the ERL value were highly toxic and the sample with the highest single concentration (>75 ppm, station CHP 10 in Charleston Harbor) was among the most toxic. Microtox™ results were highly variable and scattered among samples with copper concentrations less than the ERL value.

Another group of chemicals that showed a strong correlation with toxicity in Charleston Harbor was the pesticides (Table 9) which were significantly correlated with urchin fertilization. However, the scatterplot (Figure 106) shows that this relationship was very inconsistent and variable and two samples (those from stations B1-3 in Winyah Bay and H4-3 in the lower Cooper River) with highest concentrations were not toxic in the urchin fertilization tests. No guideline values are available for mixtures of total pesticides.

In the samples from the Savannah River, mixtures of trace metals, PAHs, and PCBs showed significant correlations with toxicity (Table 10). The sums of the 8 metals-to-ERMs quotients were highly correlated with urchin embryo development and many samples that had relatively high metals concentrations were highly toxic (usually 0.0% normal development (Figure 107). However, there were about 11 samples with relatively high metals concentrations that were not toxic. The concentrations of the five SEM also were highly correlated (Table 10), but when normalized to the concentrations of AVS, the correlations disappeared.

In contrast, the relationship between urchin embryo development and the concentrations of total HPAH showed a very strong correlation (Table 10) and a strong associative pattern (Figure 108). All samples with concentrations greater than the ERL value were highly toxic and many samples with low concentrations had greater than 90% normal development. There were numerous samples that were highly toxic as well that had low HPAH concentrations, but toxicity in those samples could have been associated with the presence of other substances, notably un-unionized ammonia.

Finally, for the Savannah River samples, the relationship between the concentrations of total PCBs and urchin embryo development is illustrated in Figure 109. Although the correlation was highly significant and there was a reasonable strong pattern of decreasing normal development with increasing PCB concentrations, unlike the concentrations of total HPAH, the concentrations of total PCBs exceeded the ERL value in only two samples (one of which was non-toxic). Therefore, on balance this association was not nearly as strong as observed with the total HPAHs.

In the 20 samples from the St. Simons Sound, the sample from Purvis Creek had relatively high concentrations of PCBs and mercury and low PAH concentrations, while the samples from Terry Creek had the opposite: low PCB and mercury concentrations and relatively high PAH concentrations. The upper Terry Creek sample was extremely toxic and the Purvis Creek sample was not toxic. The other St. Simons Sound samples had much lower chemical concentrations. The Microtox™ and two sea urchin tests showed significant correlations with many different substances (Table 11), including mixtures of substances normalized to their respective ERM values.

Many of the different trace metals showed strong correlations with measures of toxicity in St. Simons Sound, including the sum of the five SEM (Figure 110) and the cumulative metals/ERM quotients (Figure 111). Both the five SEM extracted with the acid-volatile sulfides (AVS) and the eight total metals normalized to dry weight ERMs showed a strong pattern of increasing toxicity with increasing concentrations. The sample from station F1-1 in Terry Creek had the highest concentrations of five SEM and it was highly toxic in the Microtox™ test. However, when the concentrations of five SEM were divided by the AVS concentrations, the correlation (Table 11) changed considerably (from rho = -0.836 to rho = +0.774), indicating no significant correspondence with toxicity in the Microtox™ tests. Based upon equilibrium-partitioning theory, these data suggest that although trace metals concentrations may have been elevated in some samples and they co-varied with measures of toxicity, trace metals extracted from AVS explained little of the toxicity.

Another association observed among the samples from St. Simons Sound was the correlation between measures of toxicity and the concentrations of PAHs (Table 11). In both urchin tests and in the Microtox™ test, there were significant correlations with the concentrations of most individual PAHs, total LPAHs, total HPAHs, and total PAHs. However, as illustrated in Figure 112, this correspondence was inconsistent and variable. Many samples with low concentrations of total PAHs were non-toxic in the urchin fertilization tests, but only one of the samples with total PAH concentrations greater than the ERL value was toxic and two others were non-toxic in this test. In contrast, although the correlation between normal urchin development and total PAHs was lower (rho = -0.497) than in the fertilization test (rho = -0.697), the correspondence was more, as expected (Figure 113). Fertilization test results were variable among samples with low total PAH concentrations, many of which were non-toxic, and as total PAH concentrations increased above the ERL value, all samples were highly toxic. None of these samples exceeded the ERM value of 44,792 ppb total PAH.

In summary, there was considerable evidence suggesting that toxicity observed in these samples was associated with elevated concentrations of mixtures of many different substances, most notably some trace metals, most PAHs, and un-ionized ammonia. Measures of toxicity often were highly correlated with indices of chemical mixtures (ERM quotients) and numer-

ous substances co-varied with each other and with concentrations of fine-grained particles. Also, it appears that the composition of the chemical mixtures varied among the estuaries. The concentrations of some trace metals (notably copper) exceeded background levels, exceeded effects-based guideline values, and showed strong associations with some measures of toxicity. Similarly, the concentrations of some individual and classes of PAHs were elevated above effects-based guidelines, correlated with toxicity, and showed strong associative patterns with toxicity. Ammonia probably contributed to toxicity in some of samples tested for sea urchin development, but played only a minor or no role in contributing to toxicity in the other bioassays. However, although the concentrations of some substances were elevated above background levels and effects-based guideline values and many substances showed significant correlations with toxicity; there were none that could be considered unequivocally as chemical(s) of highest concern.

Discussion and Conclusions

Generally, the concentrations of most of the potentially toxic substances differed very little among the different regions of the study area. Average concentrations of most substances were similar among all the different estuaries. Generally, many different chemicals co-occurred with each other in mixtures of differing composition. The concentrations of these mixtures, as normalized to their respective ERM values, were similar and relatively low in most samples. Several samples (from stations B1-1, B2-1, F2-1 and P1), all from tributaries of St. Simons Sound, were notably more contaminated than all others; only one of which (F2-1) was highly toxic in the bioassays.

Trace metals concentrations exceeded background levels in many samples, exceeded effects-based guideline values in some samples, were strongly correlated with some measures of toxicity, and showed a reasonable associative pattern of relatively high concentrations in the most toxic samples. Many of the trace metals co-varied with each other, occurring at relatively high concentrations in many of the same samples. However, numerous samples in all regions had SEM/AVS ratios of less than 1.0, suggesting that trace metals were not highly bioavailable in most samples. Also, only one sample had a trace metal concentration (mercury) greater than an ERM value. These data agreed very well with those from the entire Carolinian province, in which concentrations of numerous substances exceeded the ERL values, but they rarely exceeded the ERM values (Hyland et al., 1996).

Average concentrations of major organic compounds were often highest in St. Simons Sound, however, one sample from Purvis Creek and one sample from Terry Creek had relatively high concentrations of total PCBs and total PAHs, respectively, and, therefore, had the effect of elevating the average concentrations for that area. Also, average concentrations of some PAHs were somewhat higher in samples from Charleston Harbor than elsewhere. Otherwise, these substances occurred in relatively similar concentrations among the different estuaries.

The spatial patterns in chemical concentrations were unique to each estuary. In Charleston Harbor the samples from the upper Ashley and Cooper rivers and Shipyard Creek had some of the highest chemical concentrations and those from the harbor entrance and lower Wando River generally had the lowest concentrations. In Winyah Bay, samples collected in

Georgetown harbor had the highest concentrations of trace metals and organics, but none of these concentrations were particularly high relative to effects-based guideline values. In the Savannah River, the concentrations of most chemicals were highest in samples collected directly adjacent to downtown Savannah (especially in the industrial harbors), intermediate in samples collected upstream of downtown Savannah and in the Back River, and generally decreased toward the mouth of the river. In St. Simons Sound, the concentrations of PCBs and DDTs were very high in one sample from Purvis Creek, concentrations of PAHs and some trace metals were very high in two Terry Creek samples, concentrations of mixtures of substances were intermediate in samples from the East River/Brunswick Harbor and Back River, and concentrations of all chemicals were lowest in samples collected near the mouth of the estuary. Except for the DDTs, the concentrations of most chemicals in Leadenwah Creek were highest near the mouth of this tidal stream.

Results of the cytochrome P-450 RGS assays (indicative of the presence of mixtures of mixed-function oxidase-inducers) performed in Charleston Harbor, showed the highest response in a sample collected in the lower Ashley River, followed by several other samples collected nearby. The lowest responses in these tests occurred in samples from the upper Cooper River.

Toxicity was most prevalent among the Winyah Bay samples, where significant toxicity occurred in a total of 21 (47%) of all the tests combined. The incidence of overall toxicity was lower in the tests performed with samples from Charleston Harbor and Savannah River (31% and 34%, respectively), and lowest in the samples from St. Simons Sound (23%) and Leadenwah Creek (7%). The tests of amphipod survival were least sensitive, indicating significant results in 8 of 163 samples (5%) tested from all areas. In the urchin fertilization tests of 100% porewater and the microbial bioluminescence tests, 39% and 36%, respectively, of the samples were toxic. The incidence of significant toxicity was highest (62%, 50 of 81 samples) in the urchin development tests of 100% porewater.

Some of the toxicity tests showed strong concordance and agreement on the most and least toxic samples, while others demonstrated little concordance among results. The test endpoints measured with the sea urchins agreed relatively well and both showed good agreement with the Microtox™ tests. However, none showed good concordance with the results of the amphipod tests. Therefore, spatial patterns in toxicity, based upon all of the results, were difficult to tease out of the data. The amphipod tests, the least sensitive tests performed, indicated very high toxicity in one sample from Terry Creek, a tributary to St. Simons Sound, and a sample from the upper Savannah River.

There were many samples from Charleston Harbor that were toxic in one or more of the bioassays; however, none were toxic in the least sensitive bioassay performed - the amphipod survival test. Samples indicated as toxic in one or more tests were scattered throughout the study area. No single region was remarkably toxic in all tests and there was relatively little concordance among the tests. These data suggest, therefore, that toxic samples were scattered throughout the area, not concentrated in any particular area, and, as judged by the amphipod tests, toxicity was not severe. The Microtox™ tests indicated many samples from the Cooper River were toxic and only two samples were toxic from the Ashley River, whereas the urchin fertilization and copepod reproduction tests indicated that most of the Ashley River samples were toxic. Several of the most seaward stations in the lower harbor were consistently non-toxic in the tests, suggesting that toxic conditions did not extend beyond the harbor entrance.

The samples from Leadenwah Creek, a system that has historically received considerable pesticide runoff from agriculture, were not highly toxic. Only one sample showed toxicity in the Microtox™ tests, none were toxic in the amphipod survival tests, and a very slight decrease in urchin fertilization was observed in the porewaters of two samples.

Eight of the nine samples from Winyah Bay showed toxicity in at least one of the bioassays and toxicity was most severe in the Georgetown harbor area. Toxicity generally decreased seaward down the bay. However, the most seaward station was toxic in two tests and showed the highest fold induction rate in the cytochrome P-450 assay. Therefore, the seaward extent of toxicity was not determined.

Toxicity in St. Simons Sound was most severe in the two samples collected in Terry Creek; intermediate in samples from the Brunswick harbor area and the Turtle River (where toxicity occurred in only the urchin tests); and least toxic in the samples from the lower reaches of the Turtle River and St. Simons Sound. These data indicated that, as in Charleston Harbor, toxic conditions did not extend beyond the entrance to the estuary.

The Savannah River stations in which toxicity was observed were scattered throughout the region and each bioassay showed somewhat different patterns in toxicity. However, three tests, the Microtox™ tests and both urchin tests, indicated that samples from the central portion of the region adjacent to downtown Savannah were toxic. Also, there was generally less toxicity downstream of the city. Data from the amphipod survival tests, the least sensitive bioassay performed, showed a different pattern in toxicity; the only toxic stations were those from the upper reaches of the river and from Back River.

The total study area encompassed approximately 88 km². By weighting the results of the toxicity tests to the sizes of the sampling strata, we were able to estimate the spatial extent of toxicity. The spatial extent of toxicity differed among the estuaries and the different tests performed. As estimated by the amphipod survival, urchin fertilization, and Microtox™ tests the spatial extent of toxicity throughout the entire survey area was 0.3%, 21.3%, and 47.7%, respectively. In a survey conducted in numerous estuaries throughout the entire Carolinian province, including those surveyed in the present study, Hyland et al. (1996) reported that only one of 84 samples, representing 2% of the area of the province, was toxic in *Ampelisca abdita* bioassays. They estimated that 19% of the area was toxic in solid-phase Microtox™ tests, whereas we estimated that 47.7% of the area within the five estuaries was toxic in our Microtox™ tests of solvent extracts.

Relationships between measures of toxicity and the concentrations of individual toxicants and chemical mixtures were determined in several statistical analyses. Based upon the metal-to-aluminum ratios, some samples from the study area had excess trace metals concentrations, ostensibly due to anthropogenic sources. The concentrations of nickel, chromium, tin, and zinc were highly correlated with aluminum content and the concentrations of cadmium, copper, chromium, lead, and zinc exceeded background levels. These observations would suggest that some portions of the trace metals concentrations in the South Carolina/Georgia estuaries were contributed by human sources.

However, although chemical concentrations in some samples exceeded ERL values, no more than two samples exceeded an ERM value by any amount. Also, all except two samples had mean ERM quotients of 0.2 or less, indicating that the concentrations of chemical mixtures were relatively low (Long , E. R. in press). Based upon these data, it appears that many samples were not highly contaminated, some were moderately contaminated, and none were highly contaminated. Therefore, although some trace metals were elevated in concentrations relative to both the aluminum content of the sediments and the ERL values, only one of these concentrations (mercury in one sample) was above an ERM value. Also, the concentrations of different PAHs were elevated in only one or two samples and none of the concentrations of chlorinated substances were elevated. None of the chemical concentrations exceeded any of the five proposed national sediment quality criteria (U. S. EPA, 1994). As a consequence, a high degree of toxicity in the least sensitive bioassay (amphipod survival) would not be expected in these samples, and, indeed, only a few samples showed toxicity in the amphipod survival tests. However, the chemical concentrations exceeded numerous ERL values and could be expected to cause toxicity in highly sensitive bioassays, such as the urchin fertilization and Microtox™ tests, and, indeed, these tests showed toxicity in many samples.

Largely because the majority of the samples indicated similarly non-toxic results in the amphipod tests, correlations between chemical concentrations and amphipod survival were not significant. A similar result was observed in a survey of the Carolinian province, in which amphipod survival was either not significantly or only weakly correlated with most substances (Hyland et al., 1996).

There was considerable evidence suggesting that toxicity observed in the other tests was associated with elevated concentrations of mixtures of many different substances, most notably, some trace metals, PAHs, and un-ionized ammonia. Measures of toxicity often were highly correlated area-wide with summed indices of chemical mixtures (total PAHs, total SEM, total metals/ERM quotients, mean ERM quotients, etc.). Also, numerous substances co-varied with each other and with concentrations of fine-grained particles. The concentrations of some trace metals (notably copper) exceeded background levels, exceeded effects-based guideline values, and showed strong associations with some measures of toxicity. The concentrations of some individual PAHs and classes of PAHs, similarly were elevated above effects-based guidelines, correlated with toxicity, and showed strong associative patterns with toxicity. Ammonia probably contributed to toxicity in some of samples tested for sea urchin development, but, played only a minor or no role in contributing to toxicity in the Microtox™, urchin fertilization and amphipod tests. Also, as expected, it appeared that the composition of the chemical mixtures varied among the different estuaries. For example, in Charleston Harbor microbial bioluminescence was correlated with many trace metal concentrations and urchin fertilization was correlated only with PAHs and other organics, while in the Savannah River microbial bioluminescence was not correlated with many chemicals and urchin fertilization was strongly correlated with many substances. In conclusion, although the concentrations of some substances were elevated above background levels and effects-based guideline values and many substances showed significant correlations with toxicity; there were none that could be considered unequivocally as chemicals of highest concern.

Figures	43
Tables	133
Appendices	160

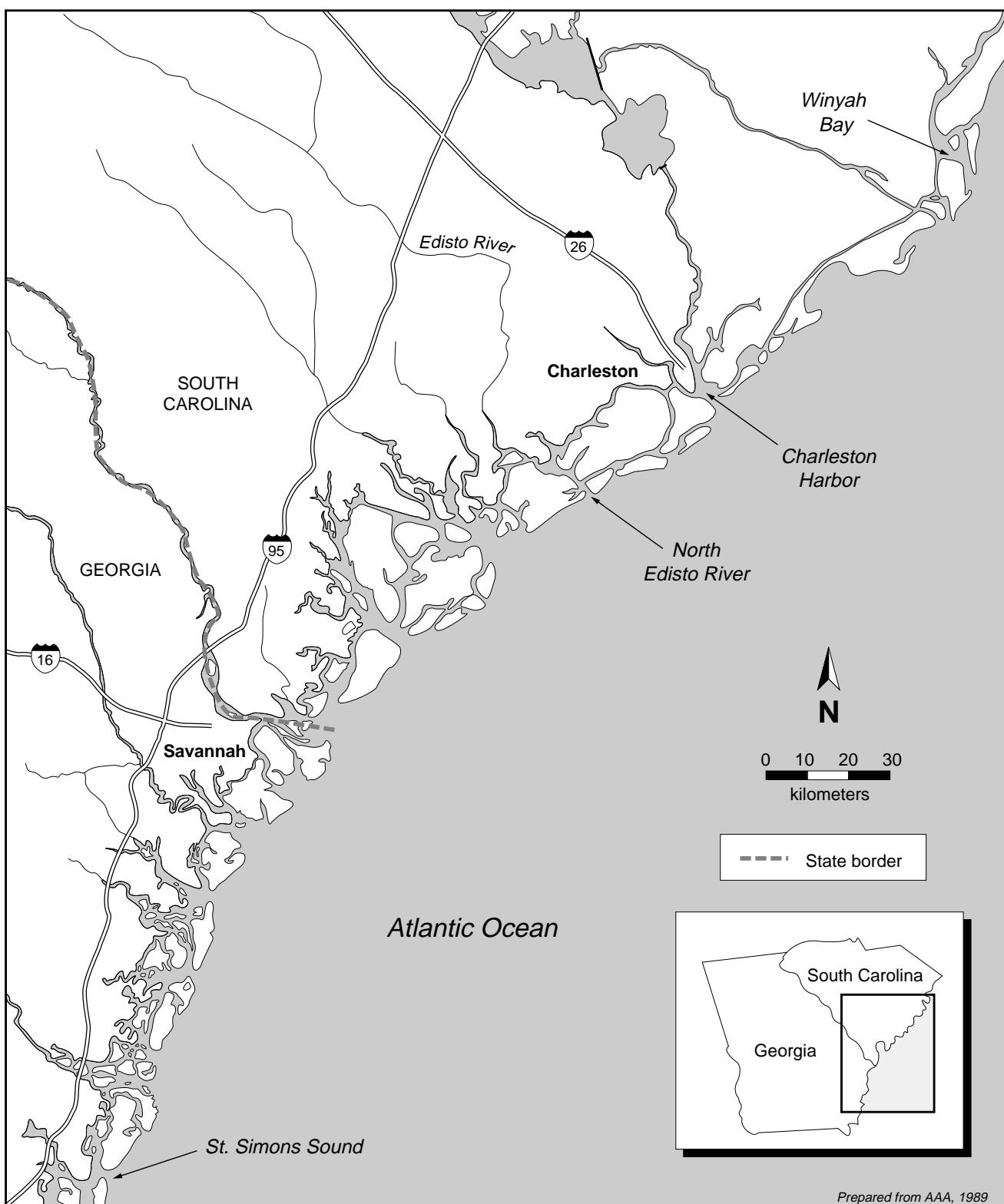


Figure 1. South Carolina and Georgia coastline.

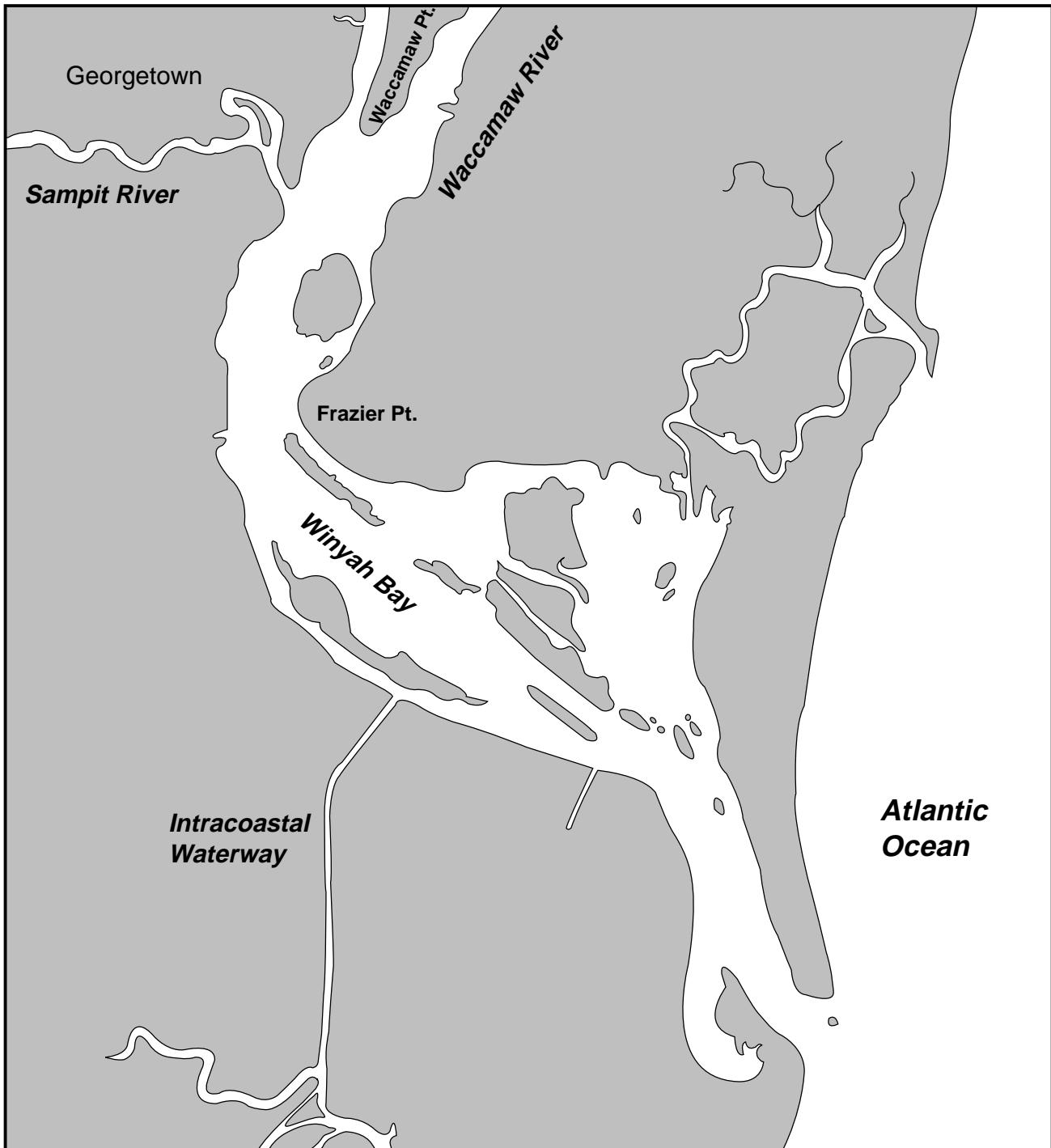


Figure 2. Winyah Bay, SC, study area.

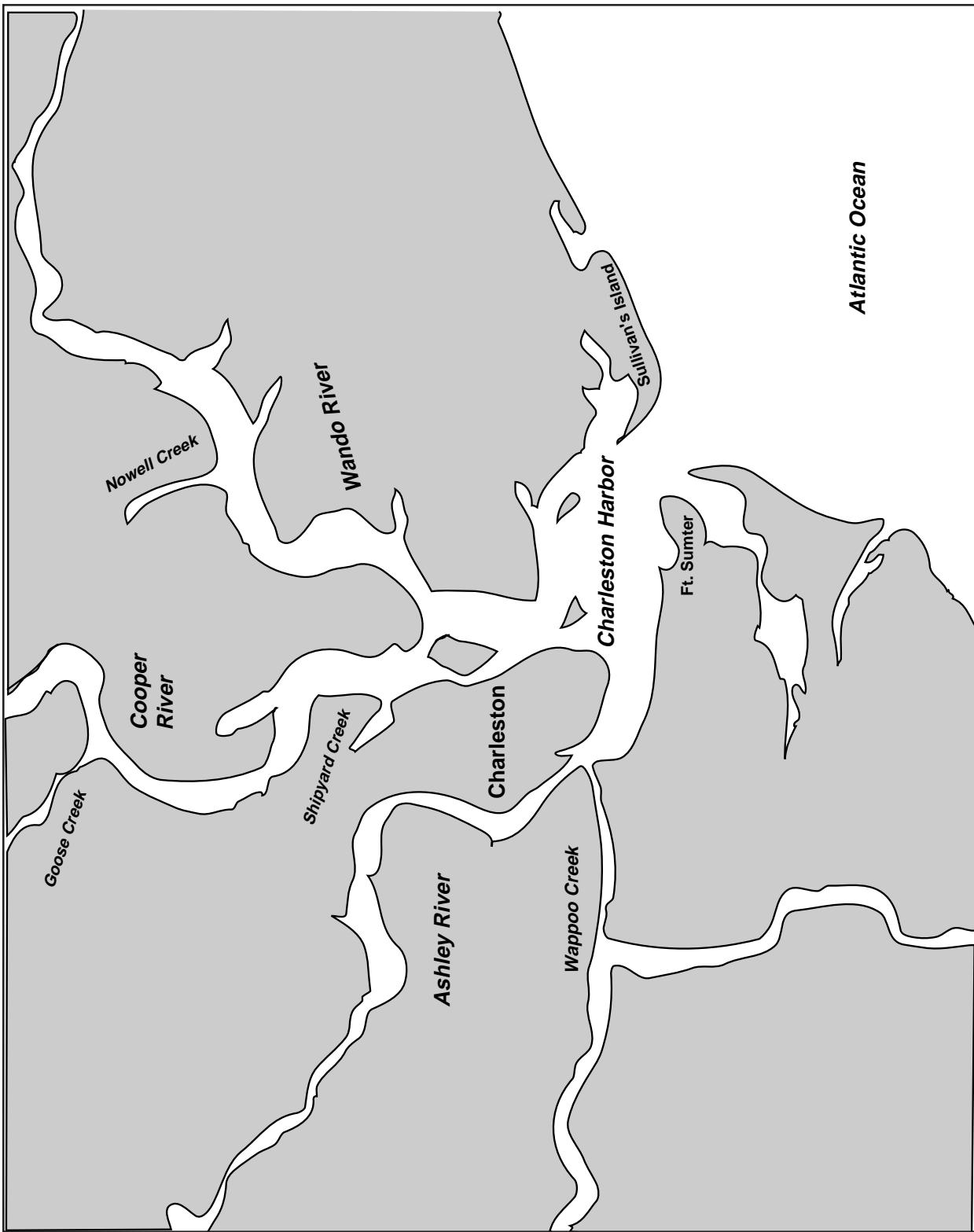


Figure 3. Charleston Harbor, SC, study area.

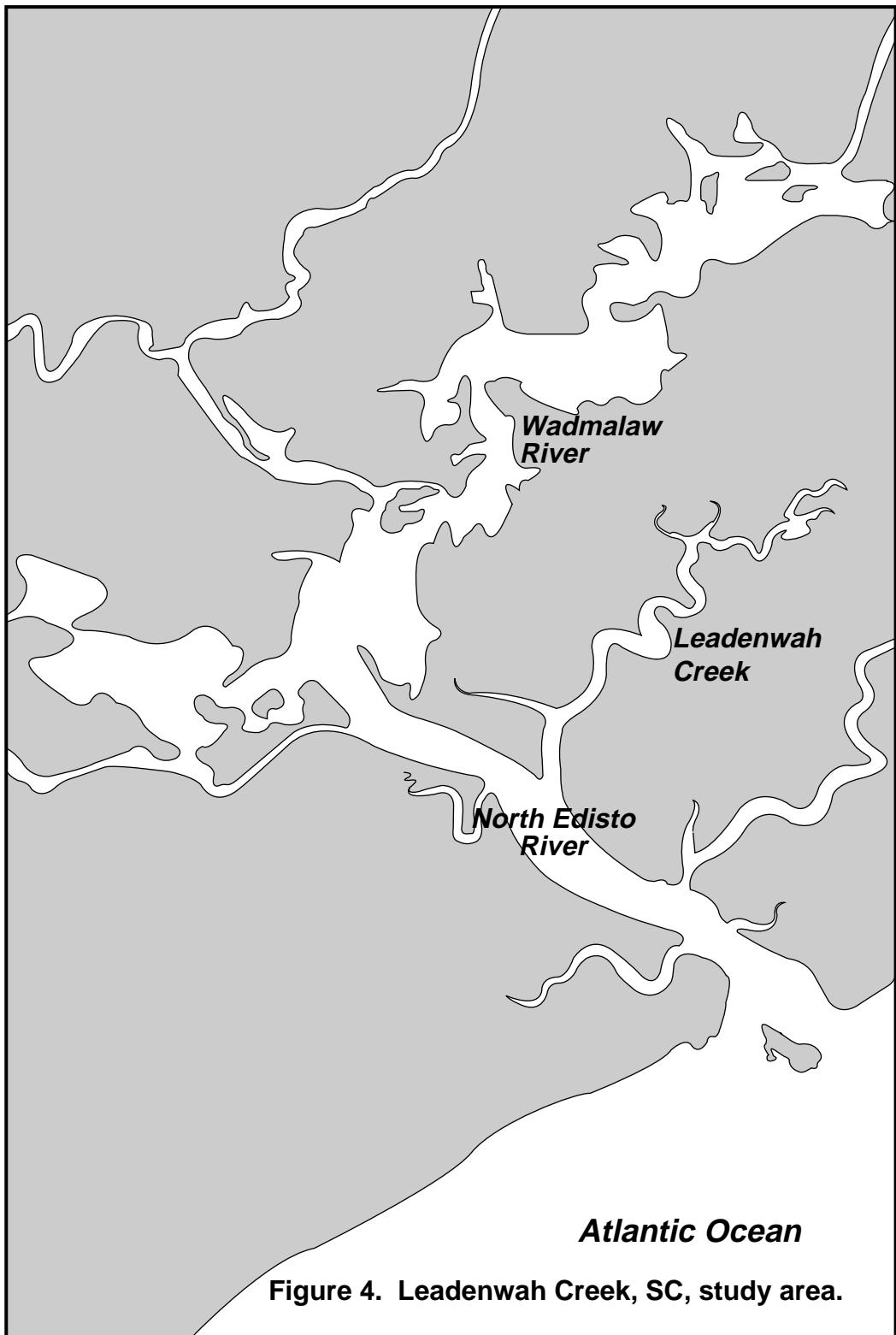


Figure 4. Leadenwah Creek, SC, study area.

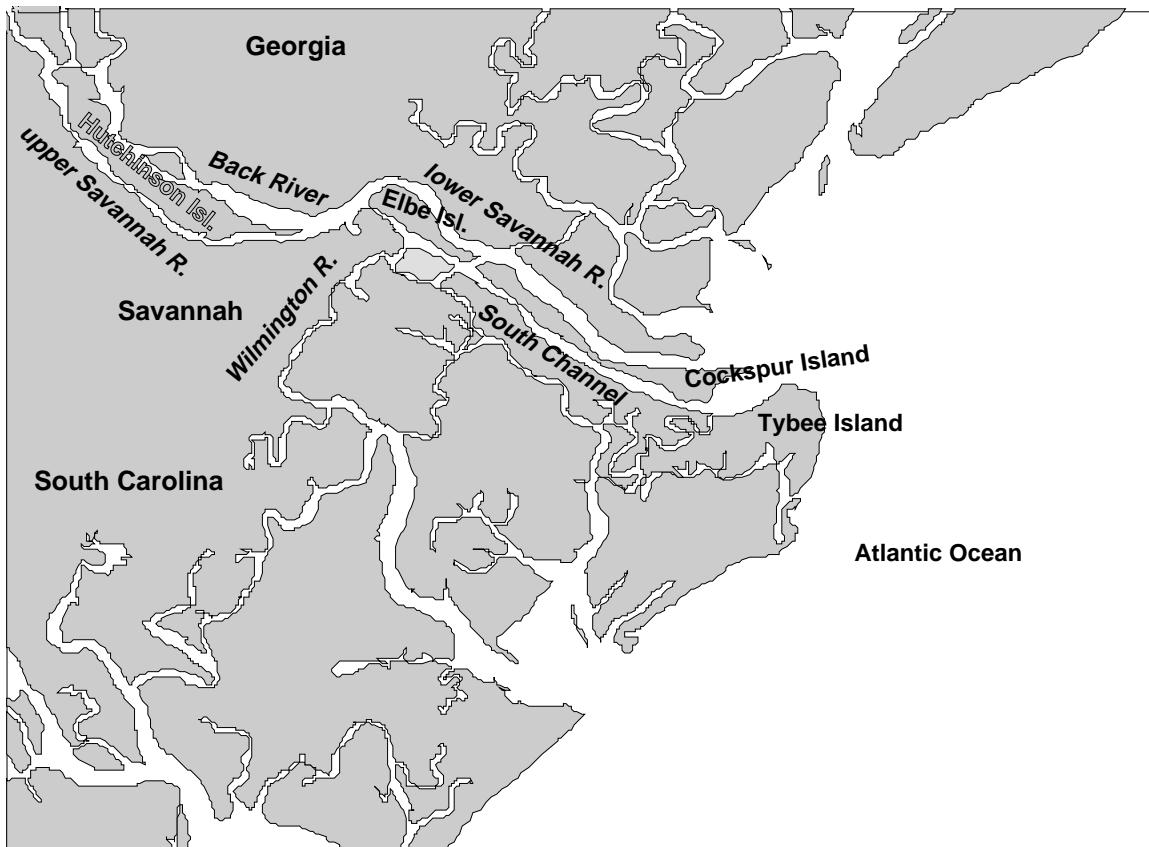


Figure 5. Savannah River (SC,GA) study area.

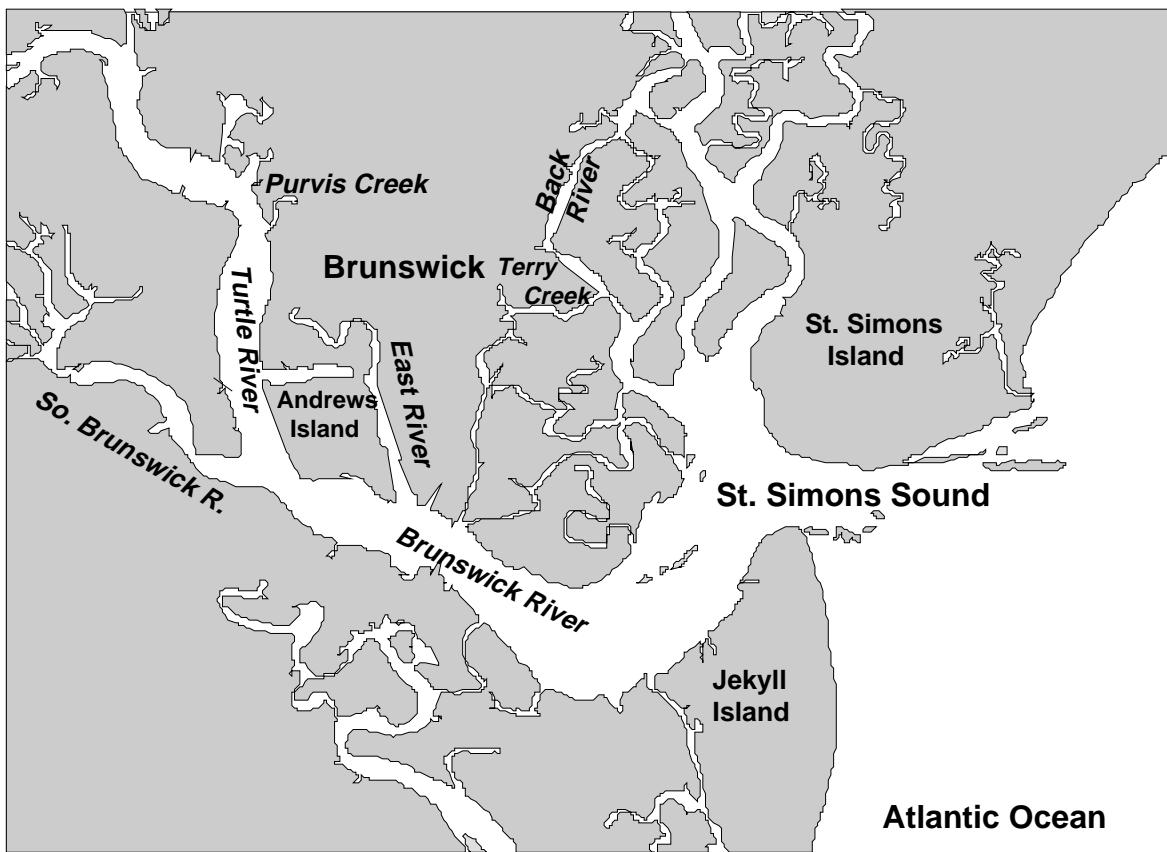


Figure 6. St. Simons Sound, GA, study area.



Figure 7. Locations of sampling stations in Charleston Harbor.

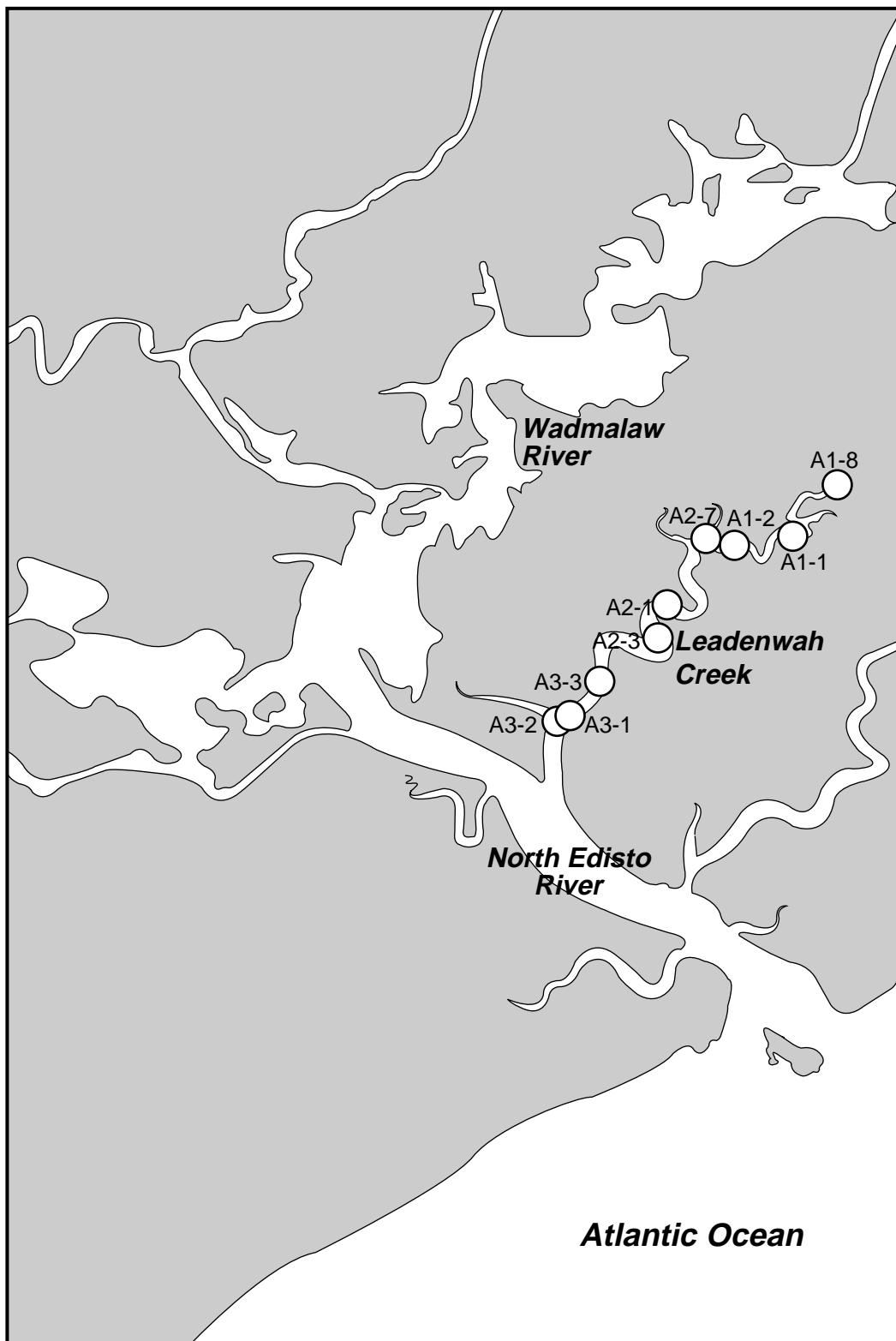


Figure 8. Locations of sampling stations in Leadenwah Creek.

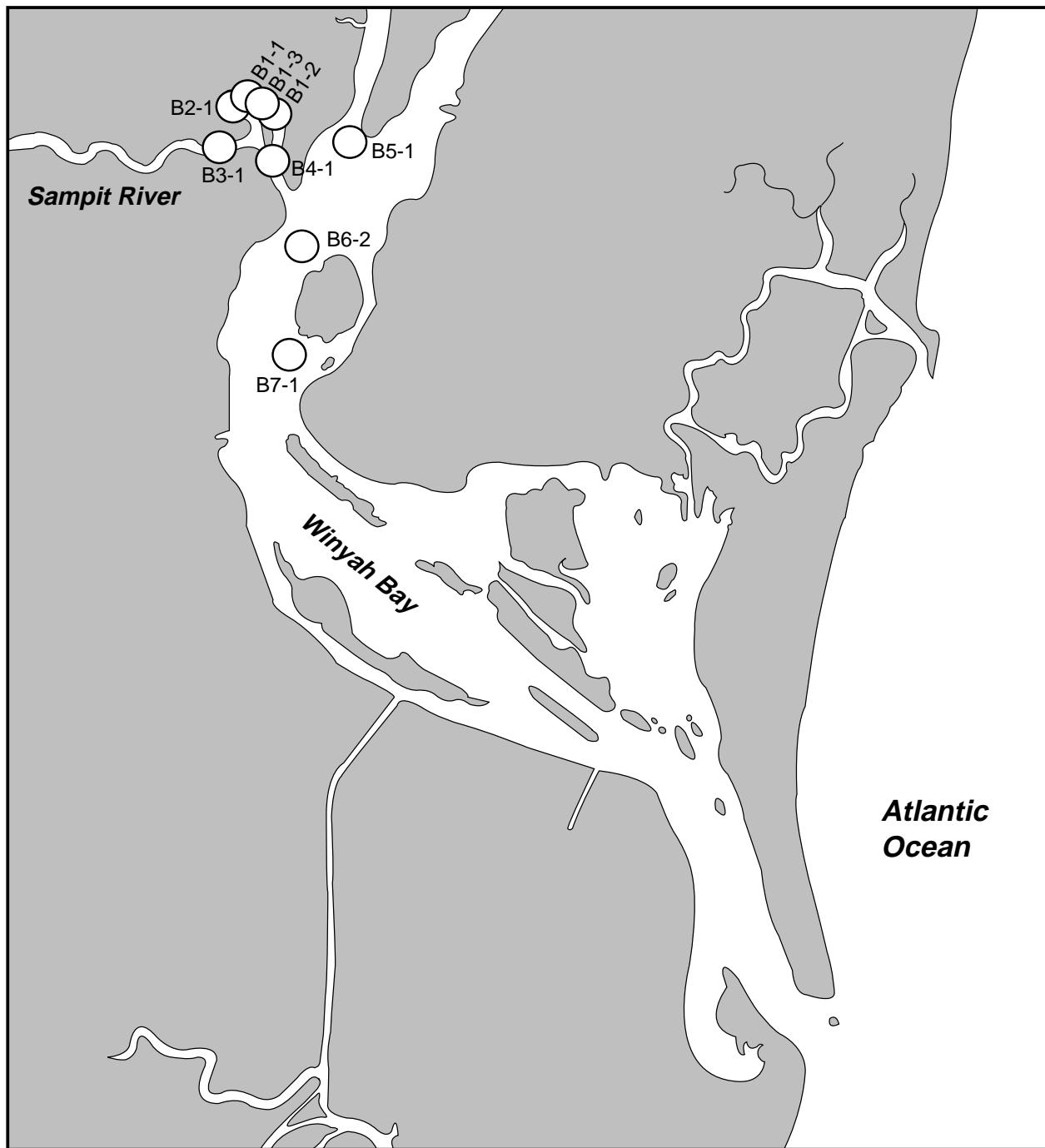


Figure 9. Locations of sampling stations in Winyah Bay.

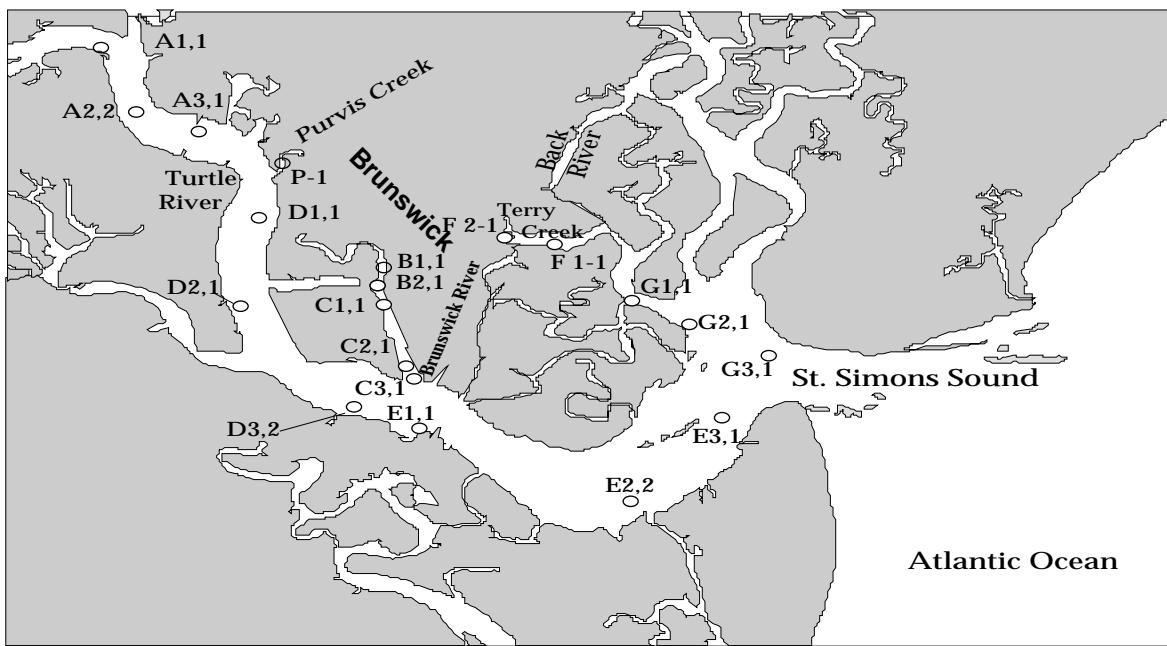


Figure 10. Locations of sampling stations in St. Simons Sound.

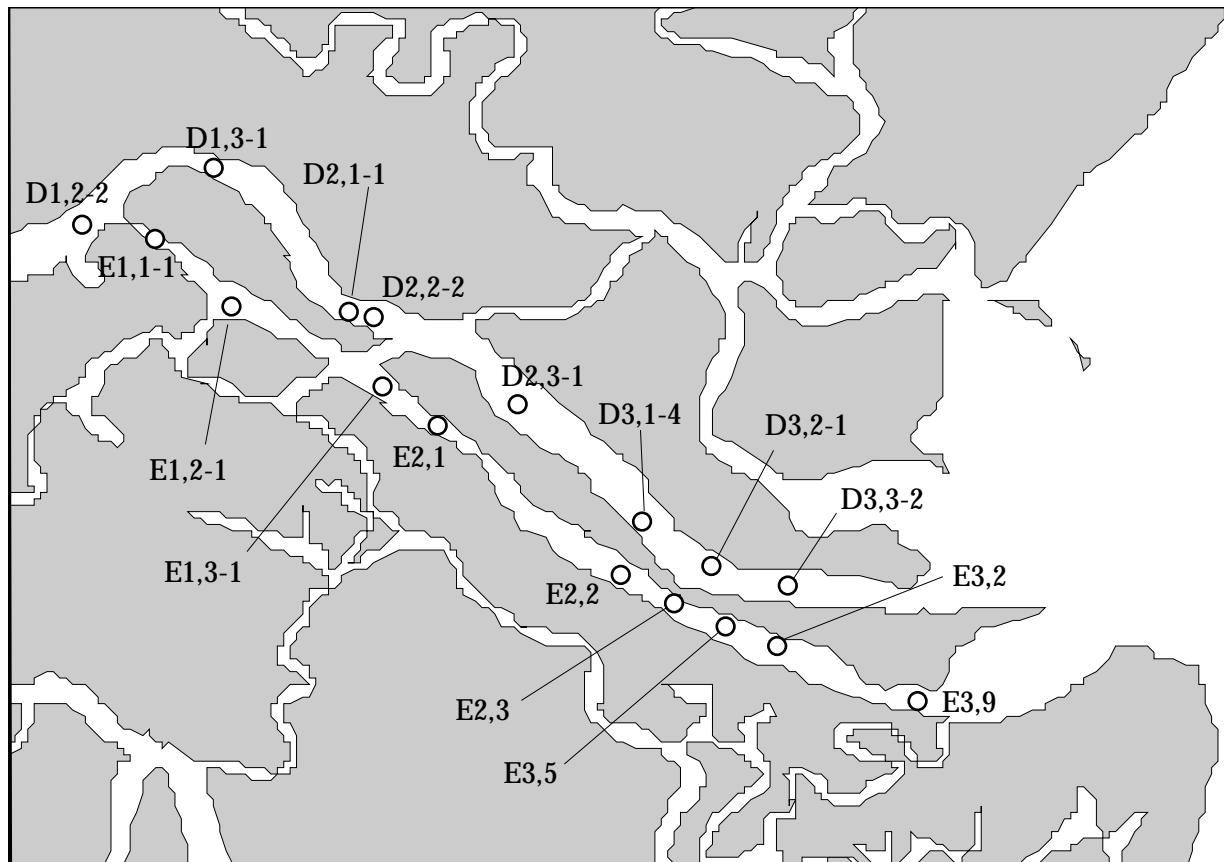


Figure 11. Locations of sampling stations in the lower Savannah River.

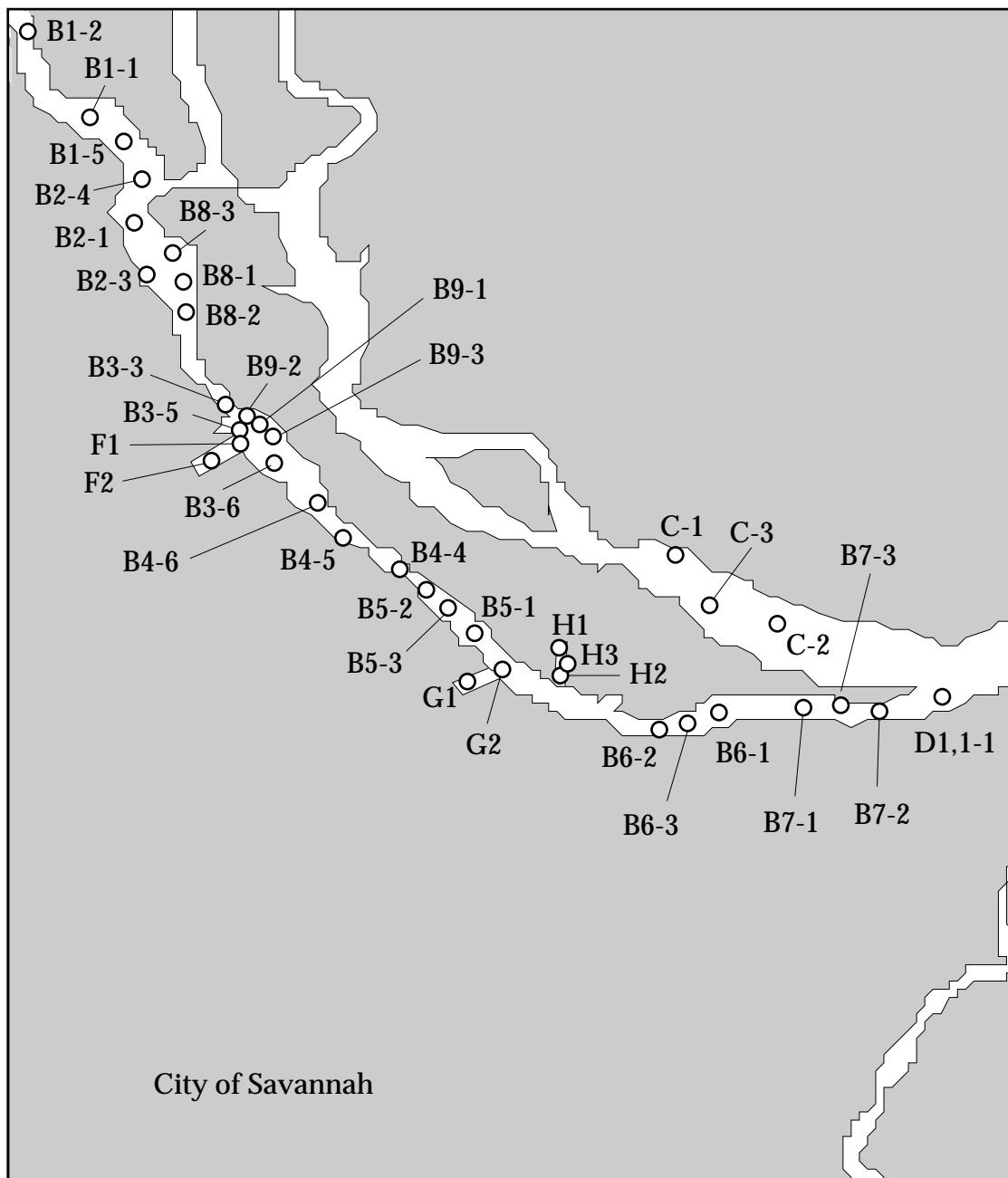


Figure 12. Locations of sampling stations in the mid-Savannah River.

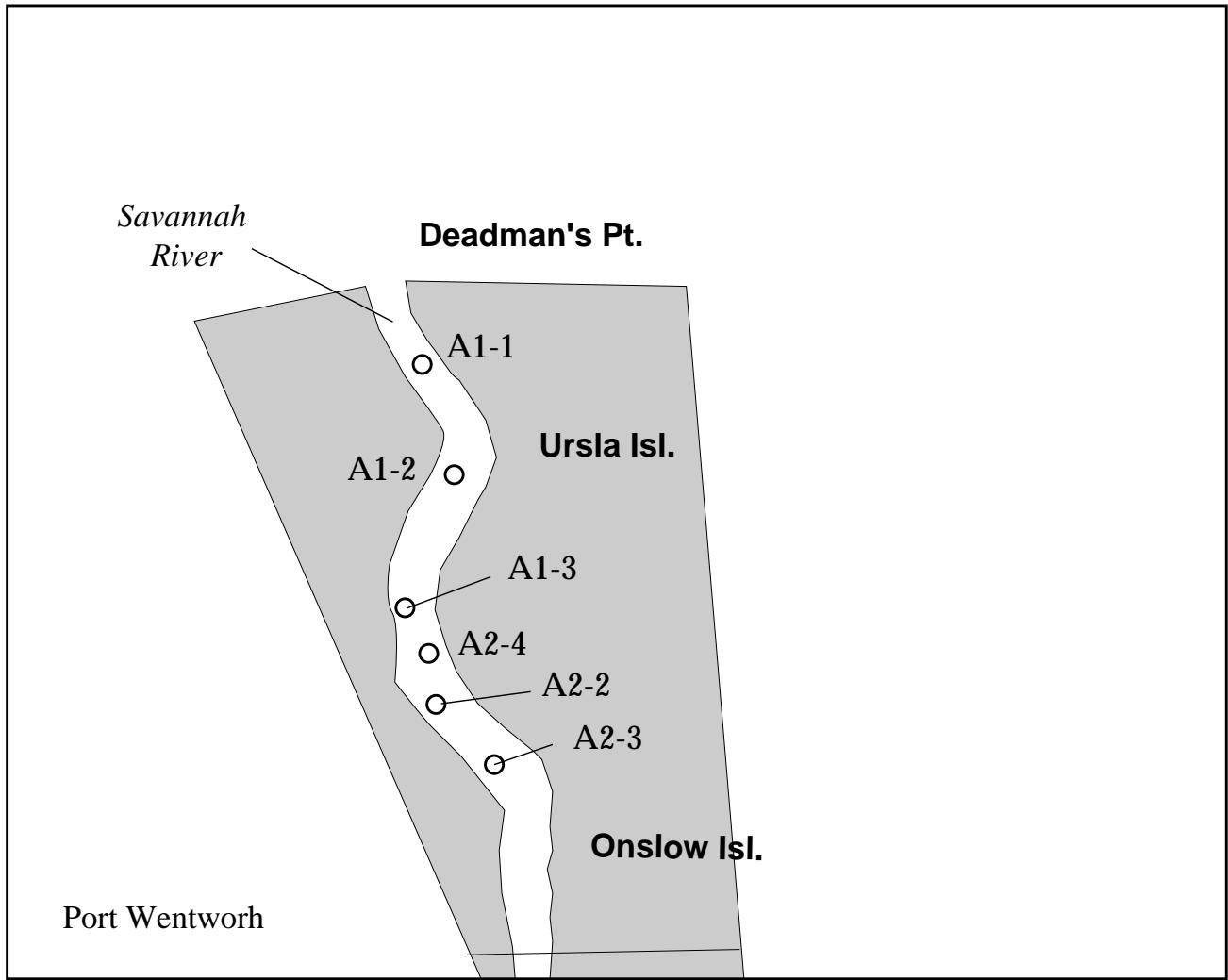


Figure 13. Locations of sampling stations in the upper Savannah River.

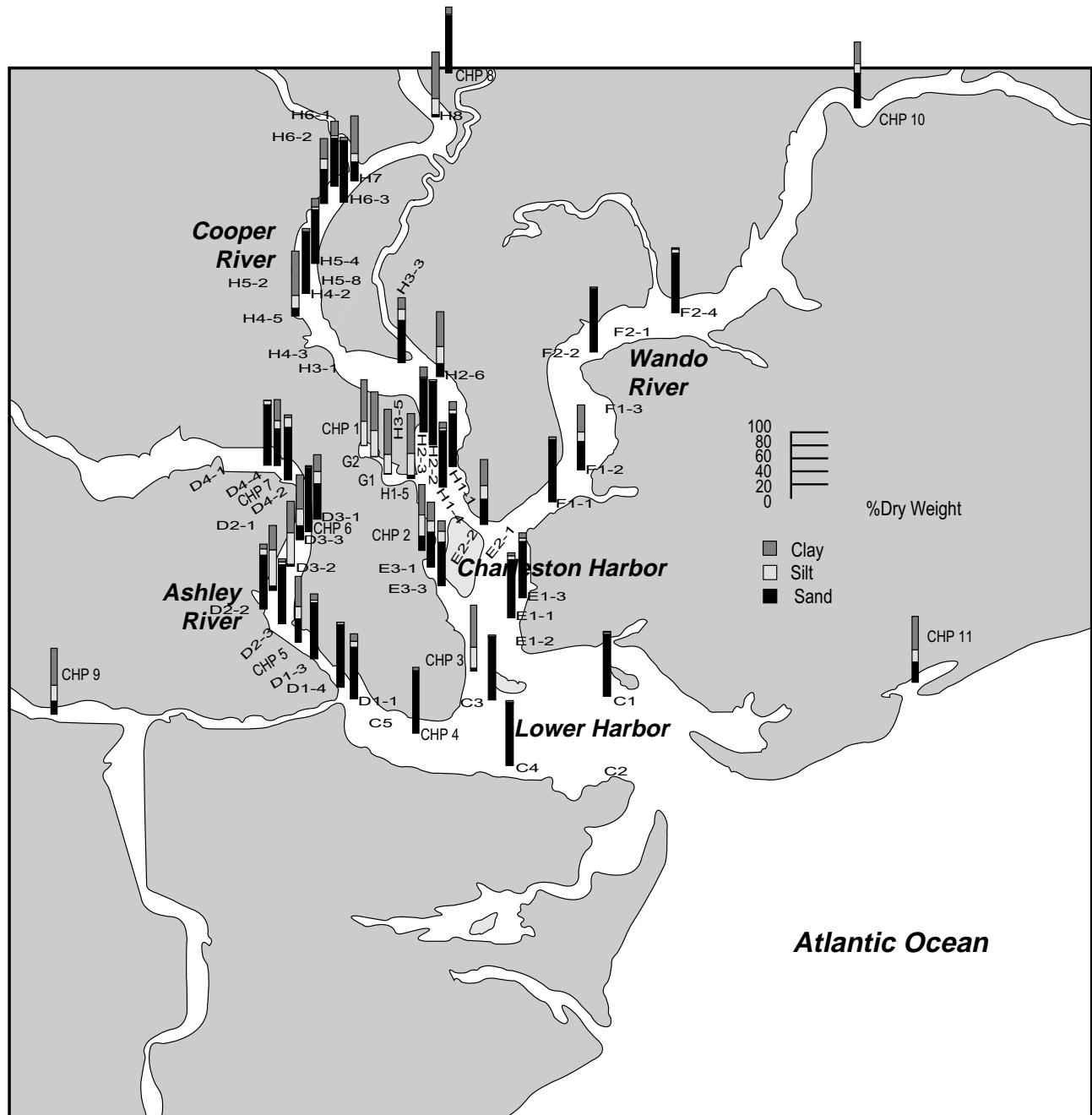


Figure 14. Percent sand, silt, and clay in sediment samples from Charleston Harbor.

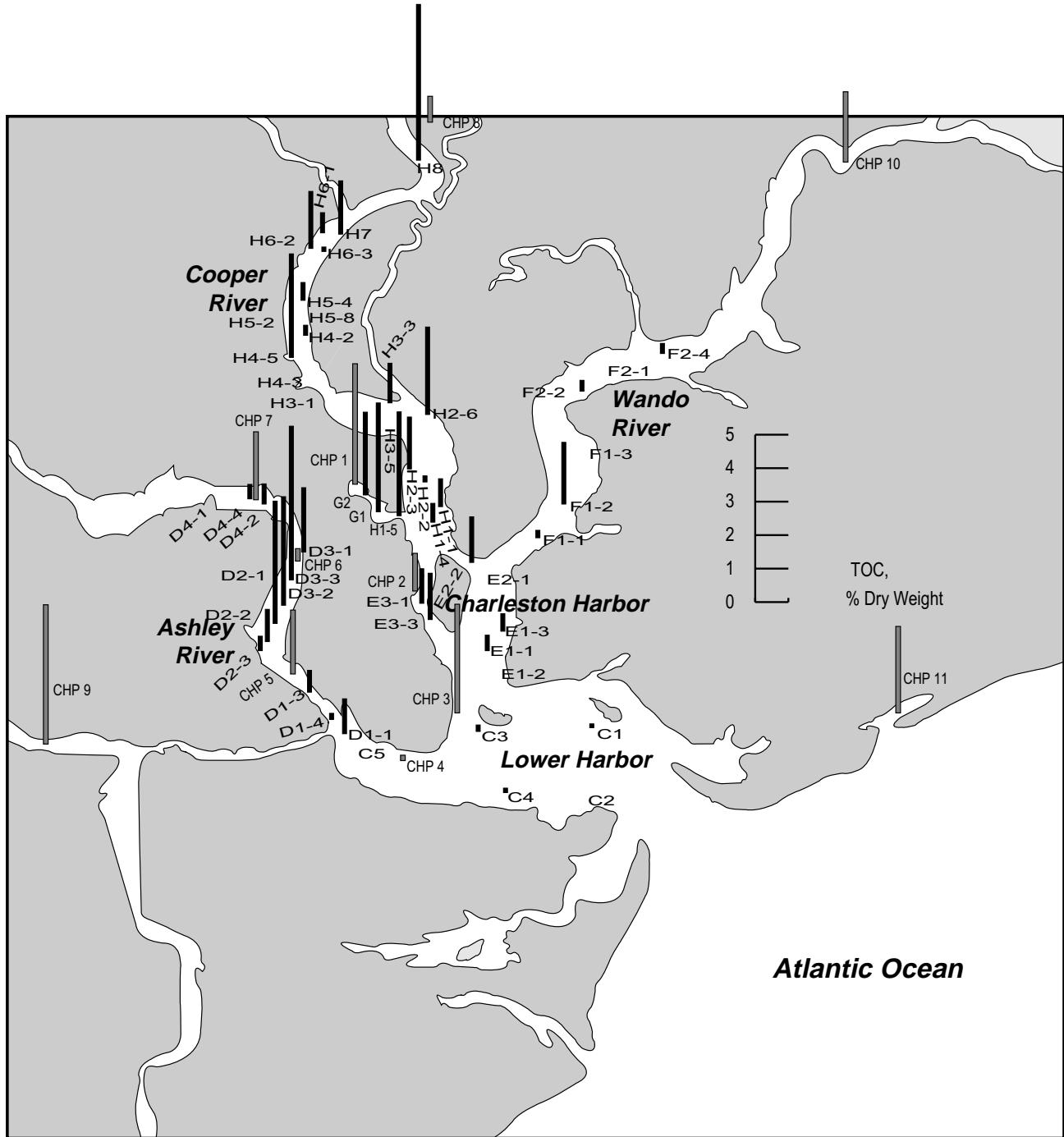


Figure 15. Concentrations of total organic carbon in sediments from Charleston Harbor.

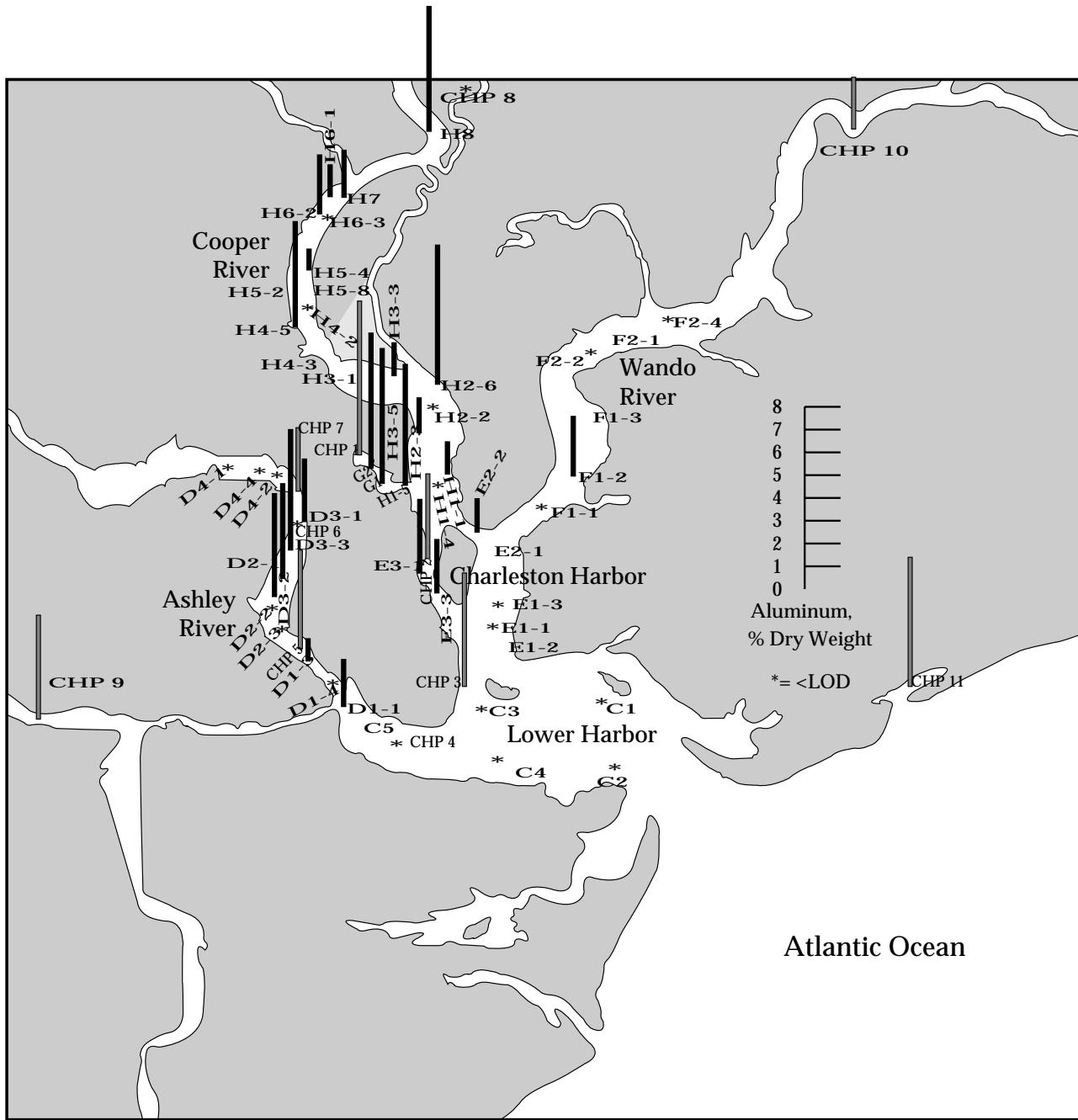


Figure 16. Concentrations of aluminum in sediments from Charleston Harbor.

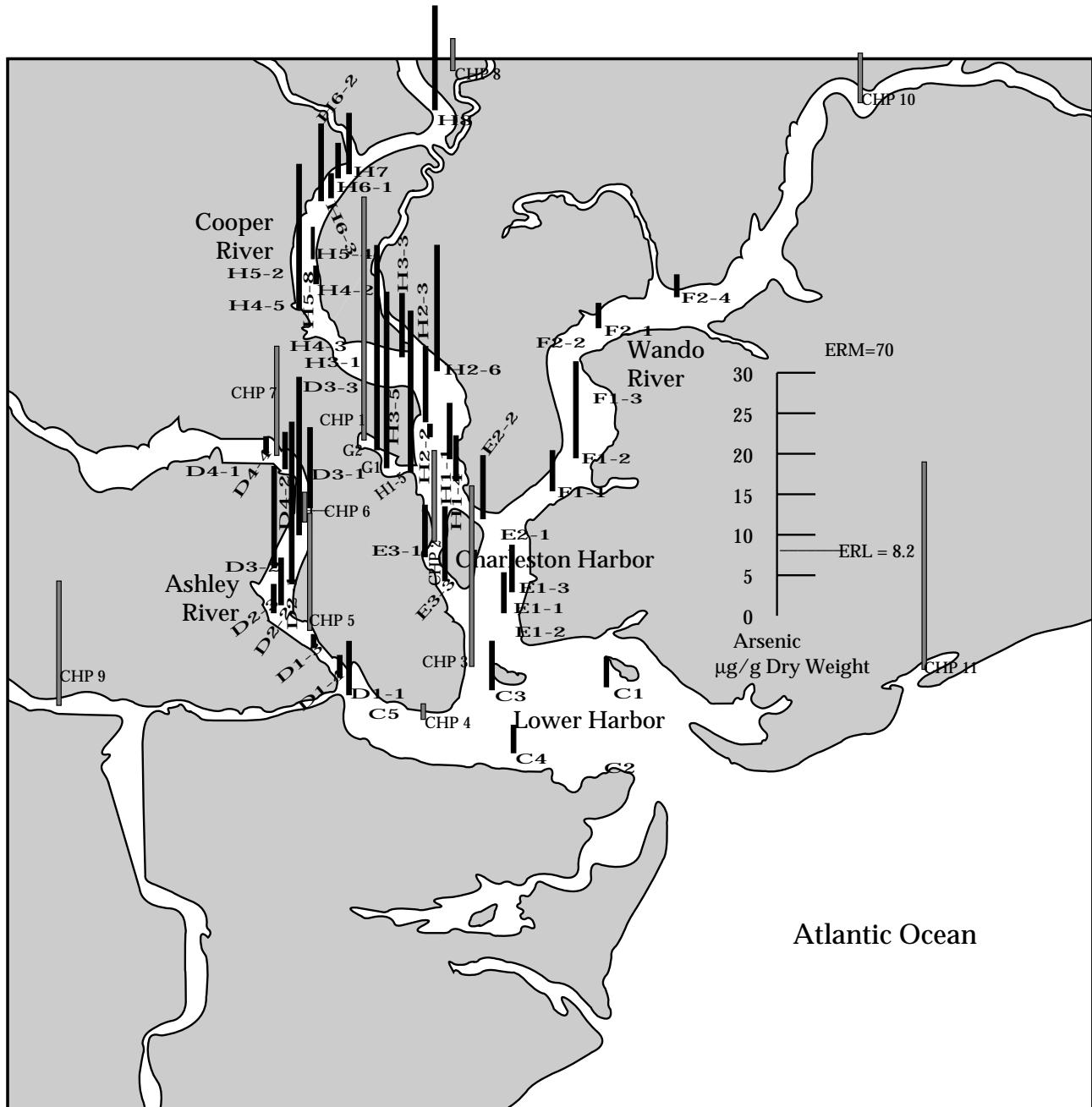


Figure 17. Concentrations of arsenic in sediments from Charleston Harbor.

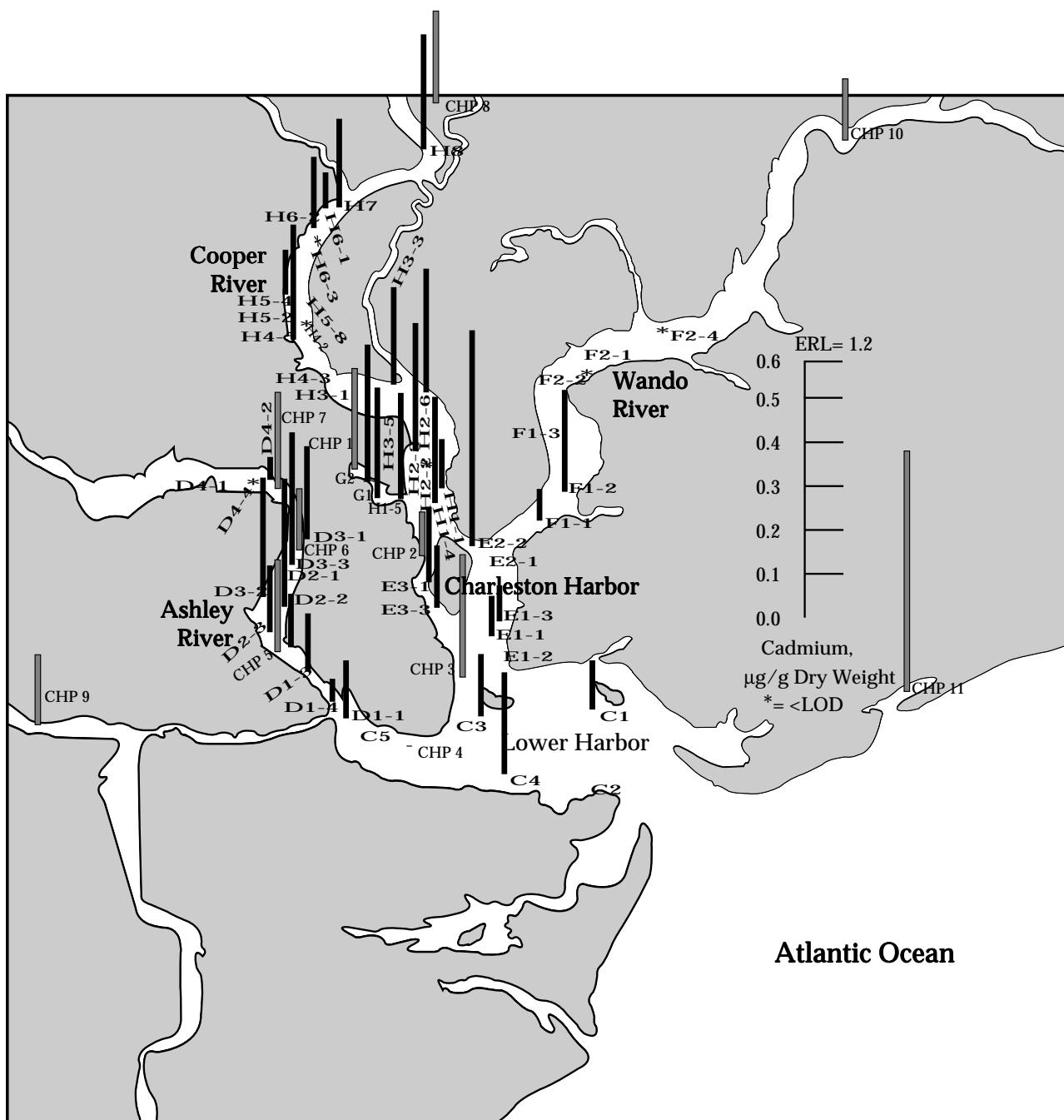


Figure 18. Concentrations of cadmium in sediments from Charleston Harbor.



Figure 19. Concentrations of copper in sediments from Charleston Harbor.

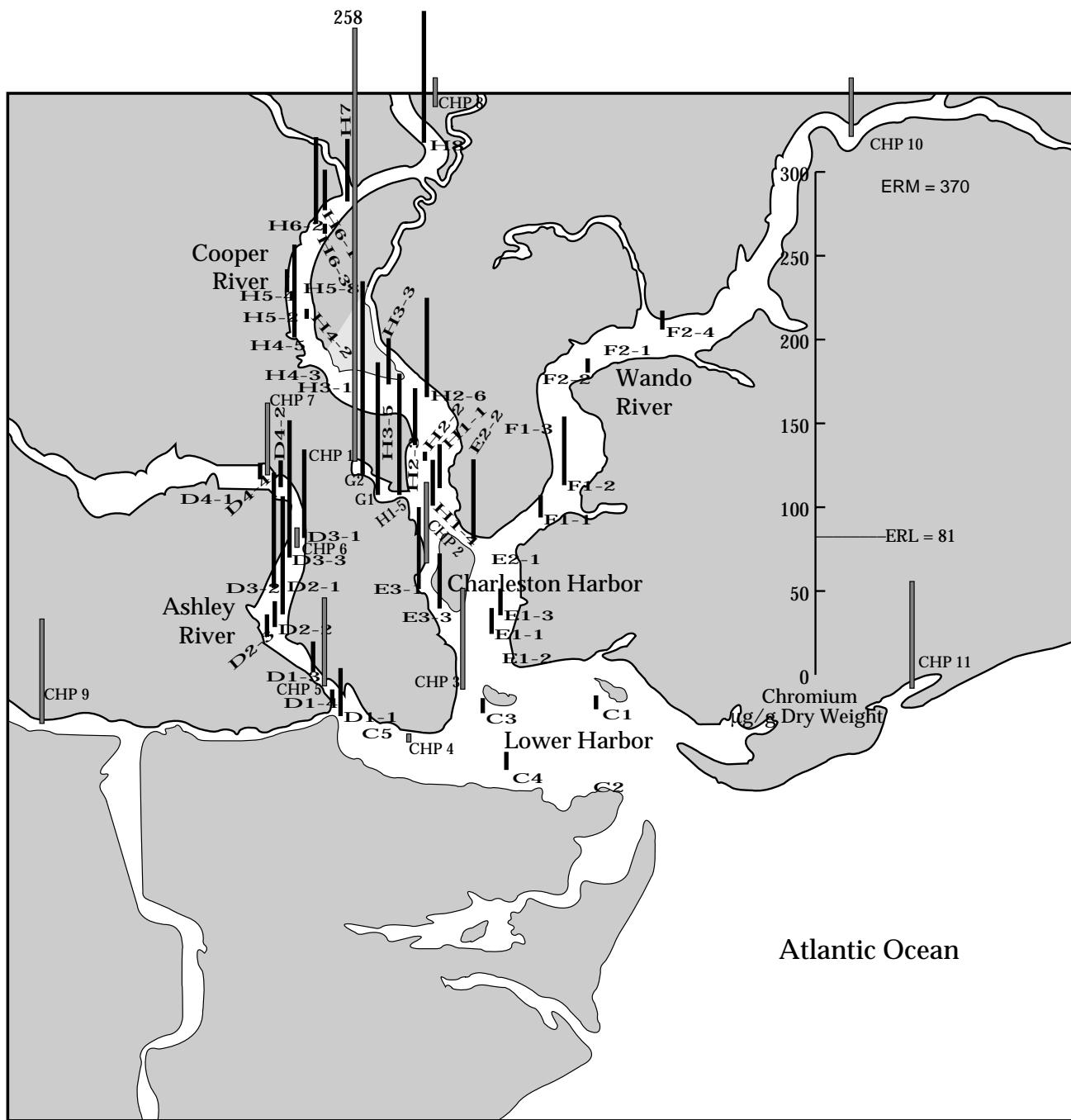


Figure 20. Concentrations of chromium in sediments from Charleston Harbor.

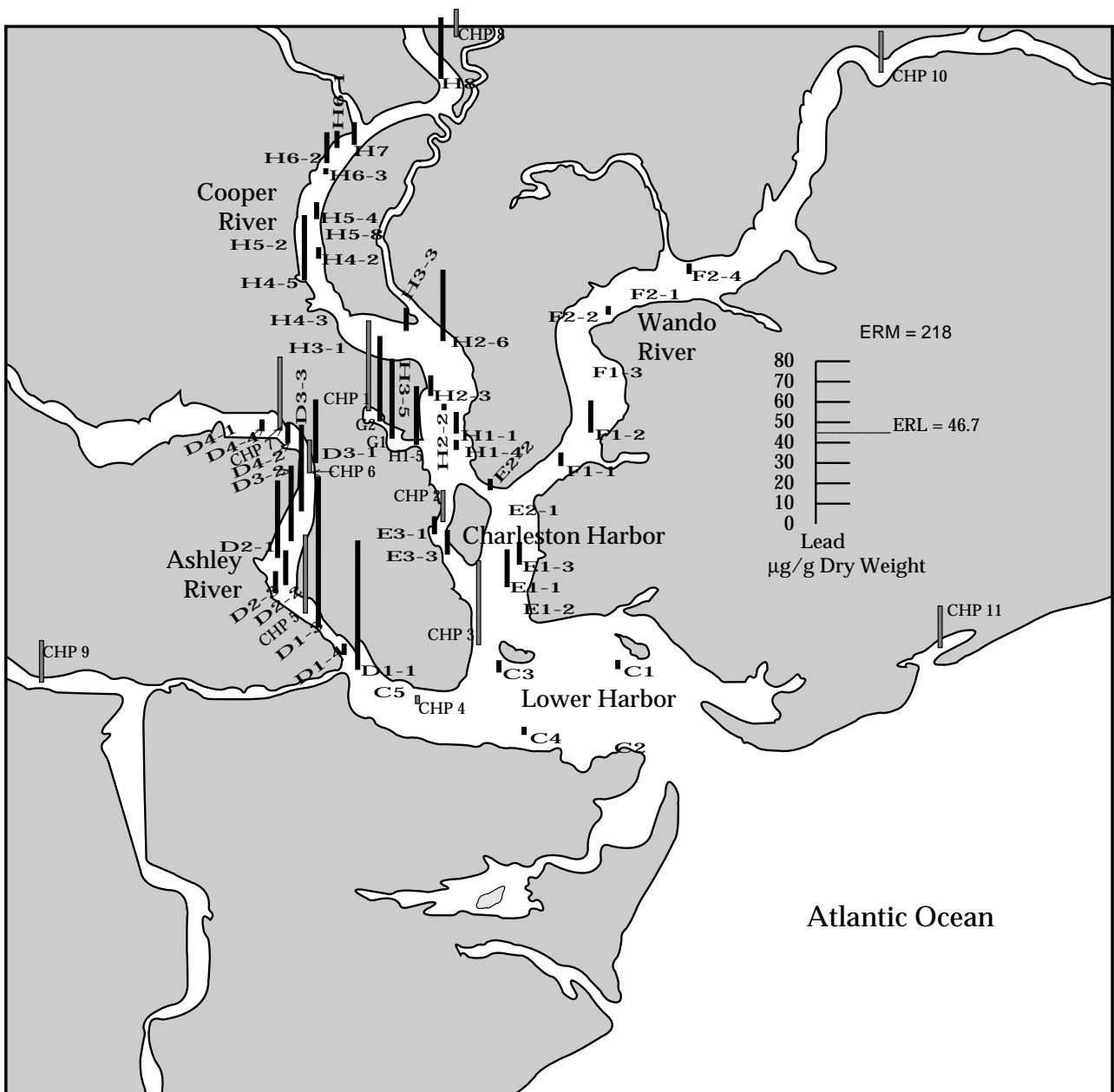


Figure 21. Concentrations of lead in sediments from Charleston Harbor.

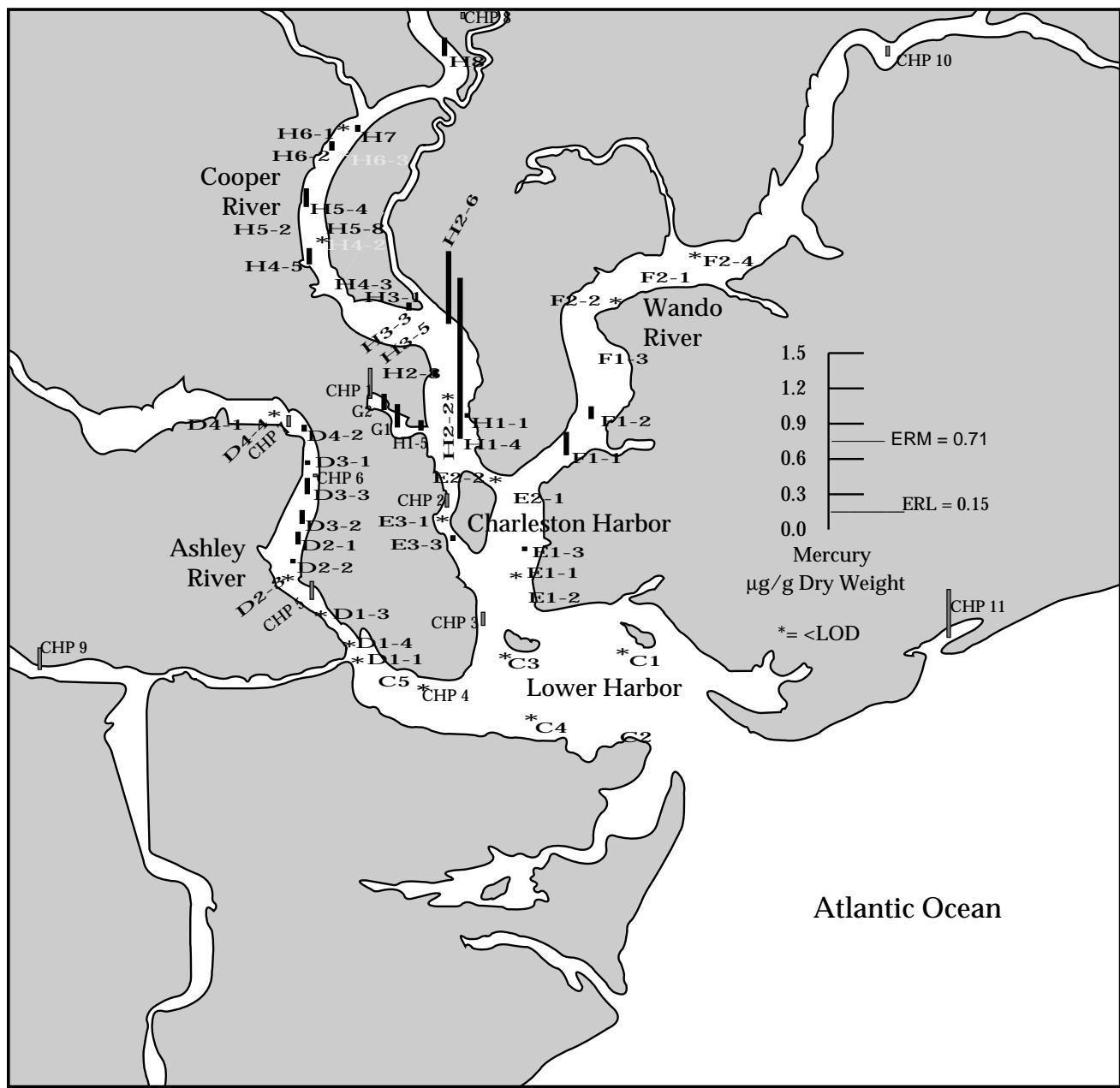


Figure 22. Concentrations of mercury in sediments from Charleston Harbor.

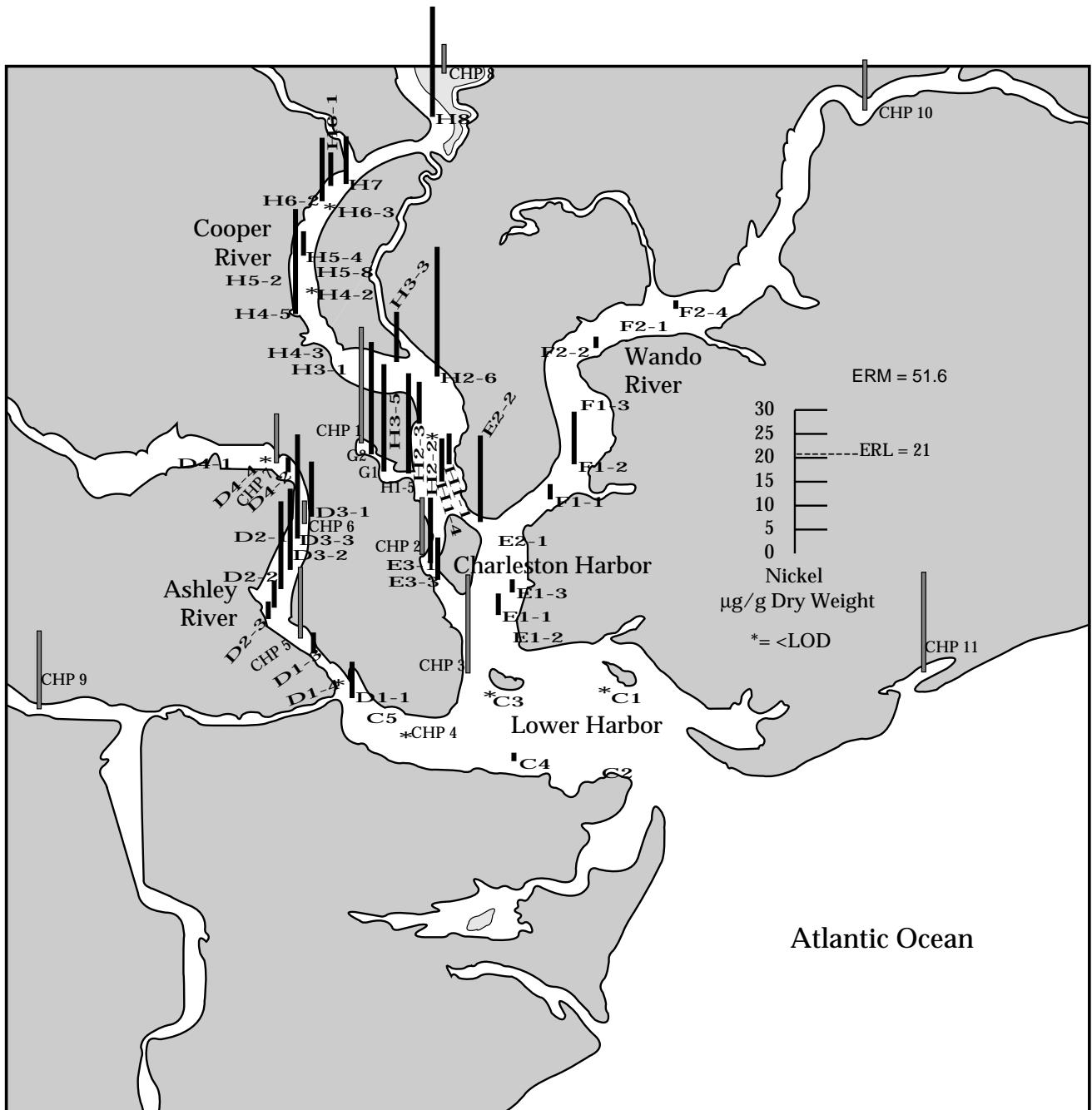


Figure 23. Concentrations of nickel in sediments from Charleston Harbor.

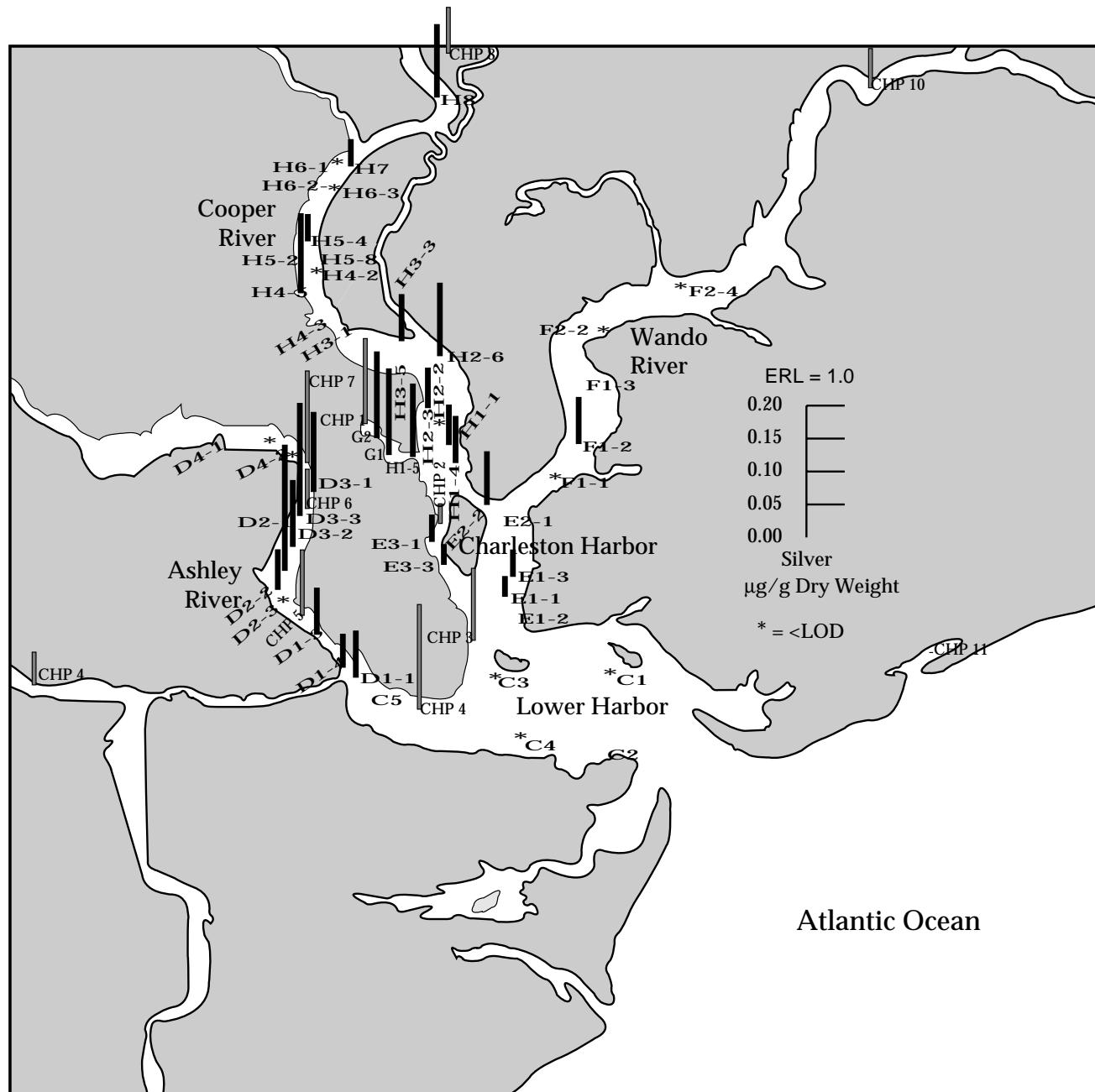


Figure 24. Concentrations of silver in sediments from Charleston Harbor.

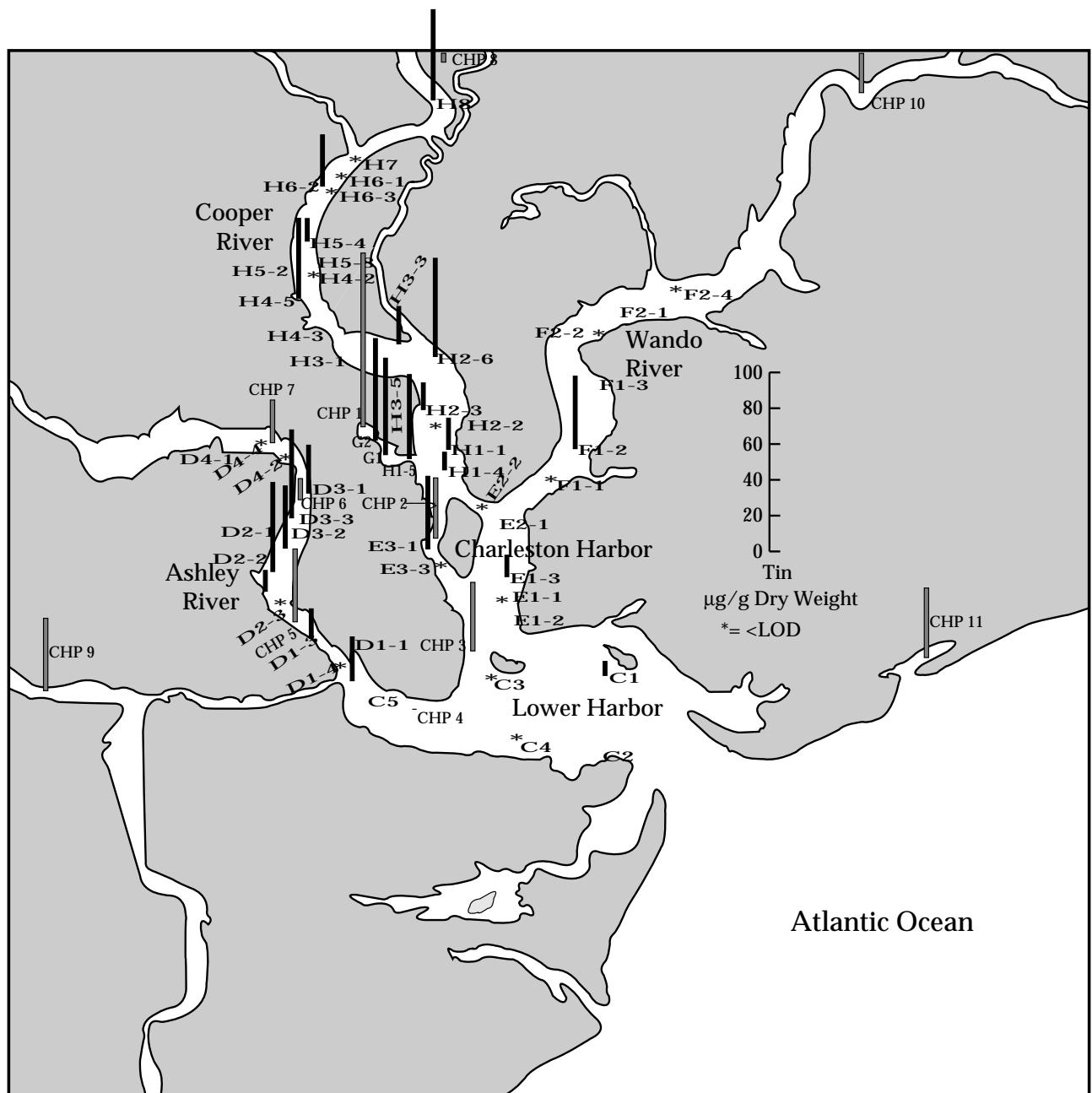


Figure 25. Concentrations of tin in sediments from Charleston Harbor.

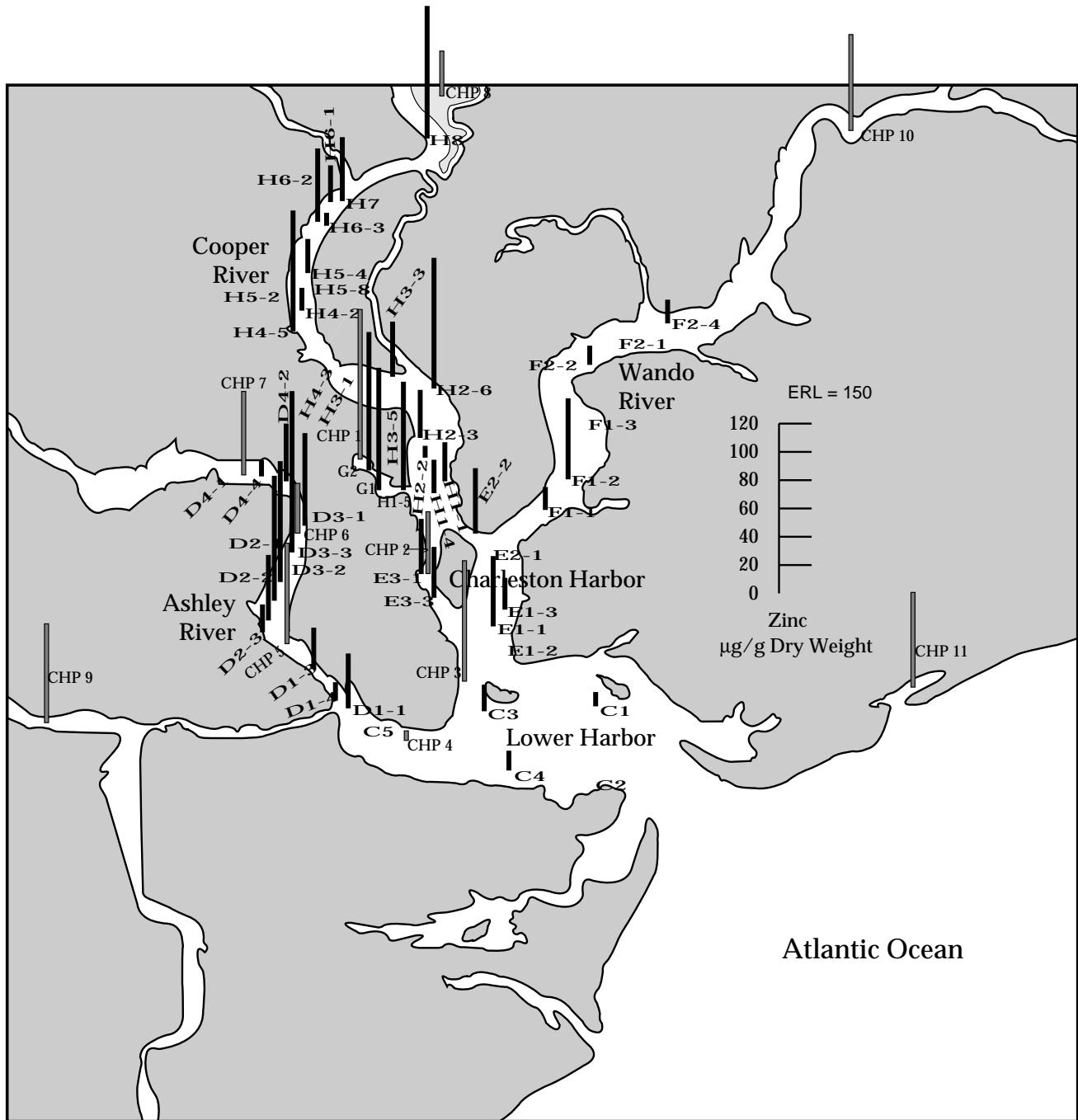


Figure 26. Concentrations of zinc in sediments from Charleston Harbor.

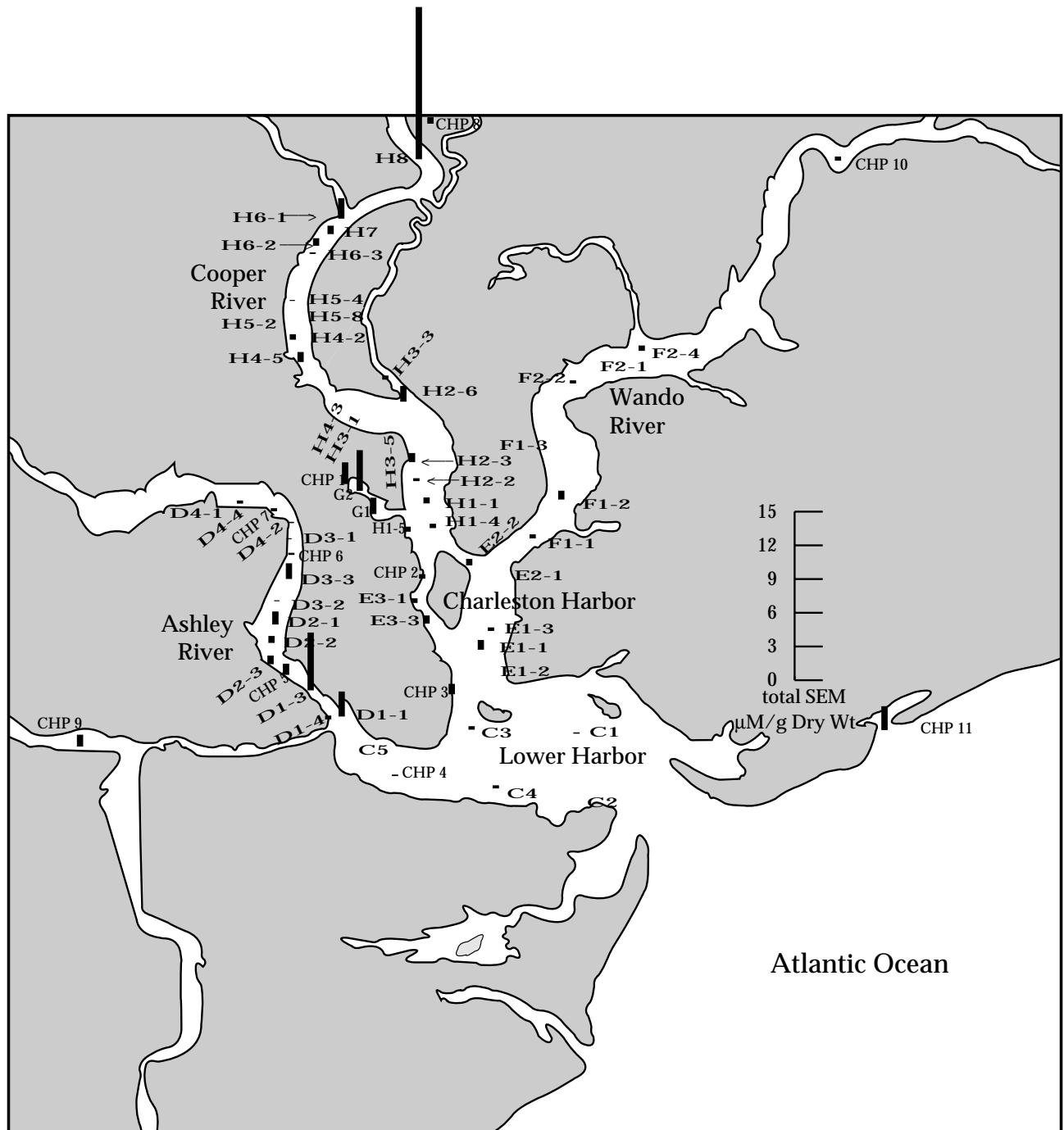


Figure 27. Concentrations of total simultaneously-extracted divalent metals in sediments from Charleston Harbor.

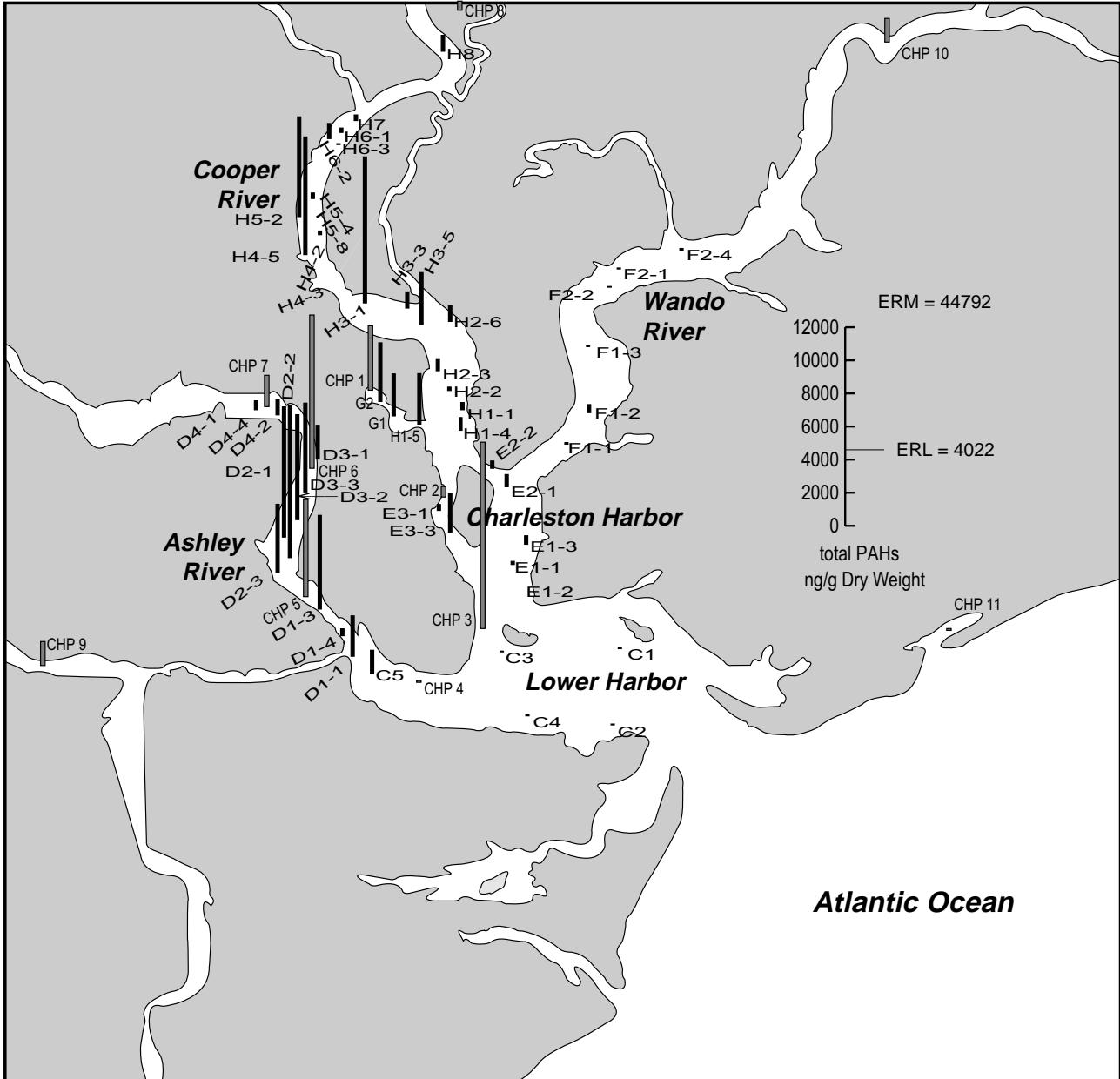


Figure 28. Concentrations of total PAHs in sediments from Charleston Harbor.

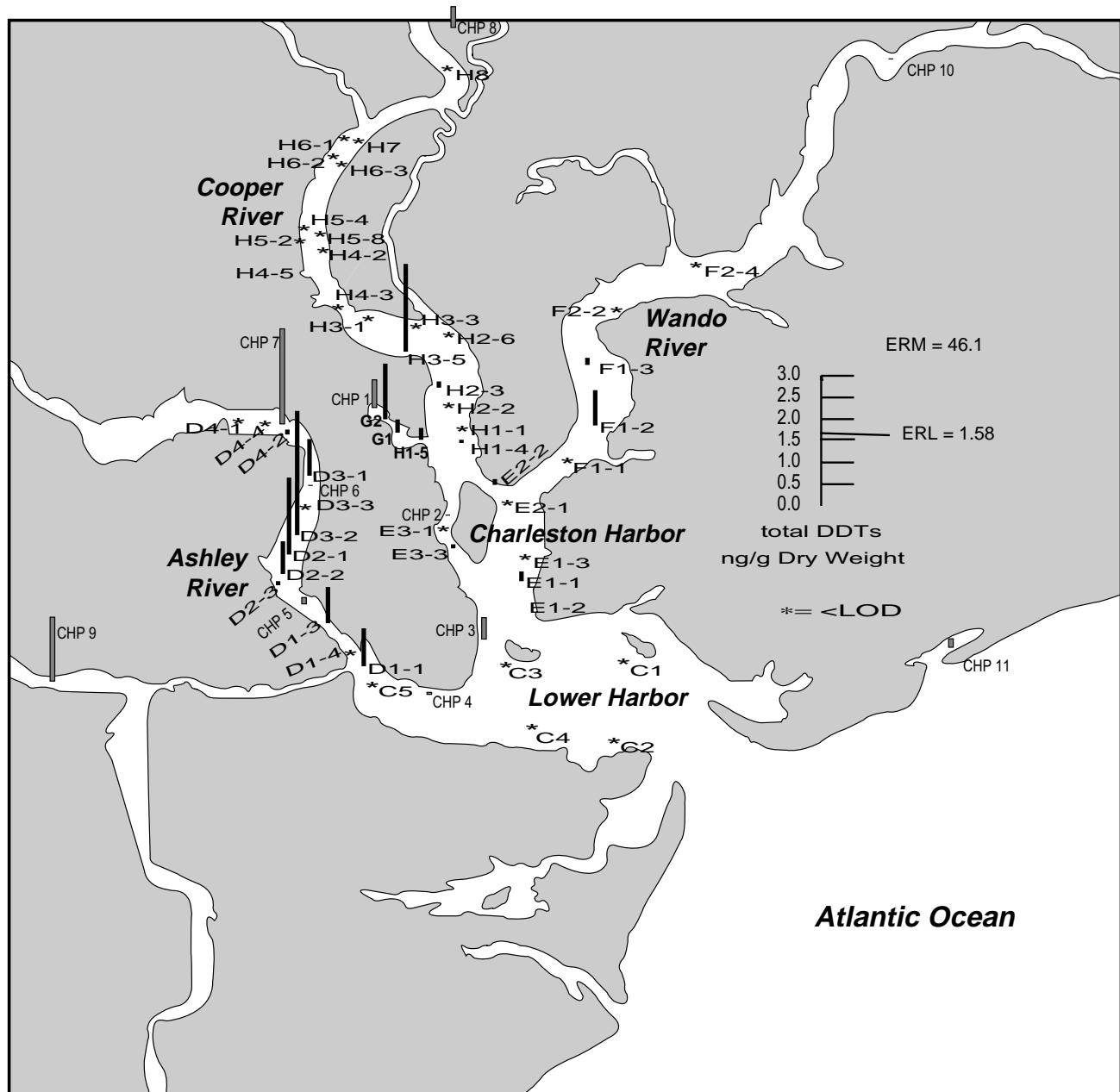


Figure 29. Concentrations of total DDTs in sediments from Charleston Harbor.

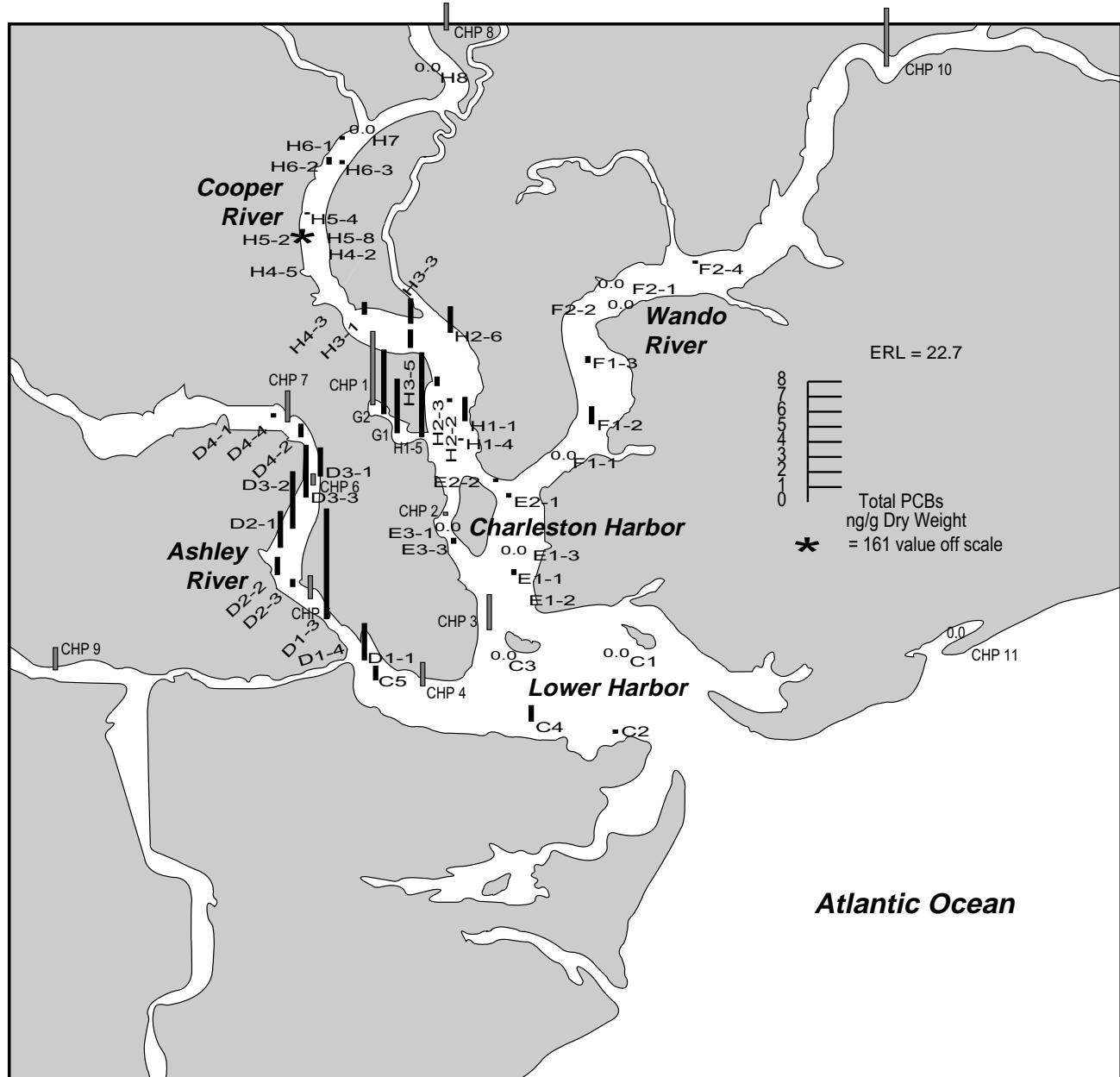


Figure 30. Concentrations of total PCBs in sediments from Charleston Harbor.

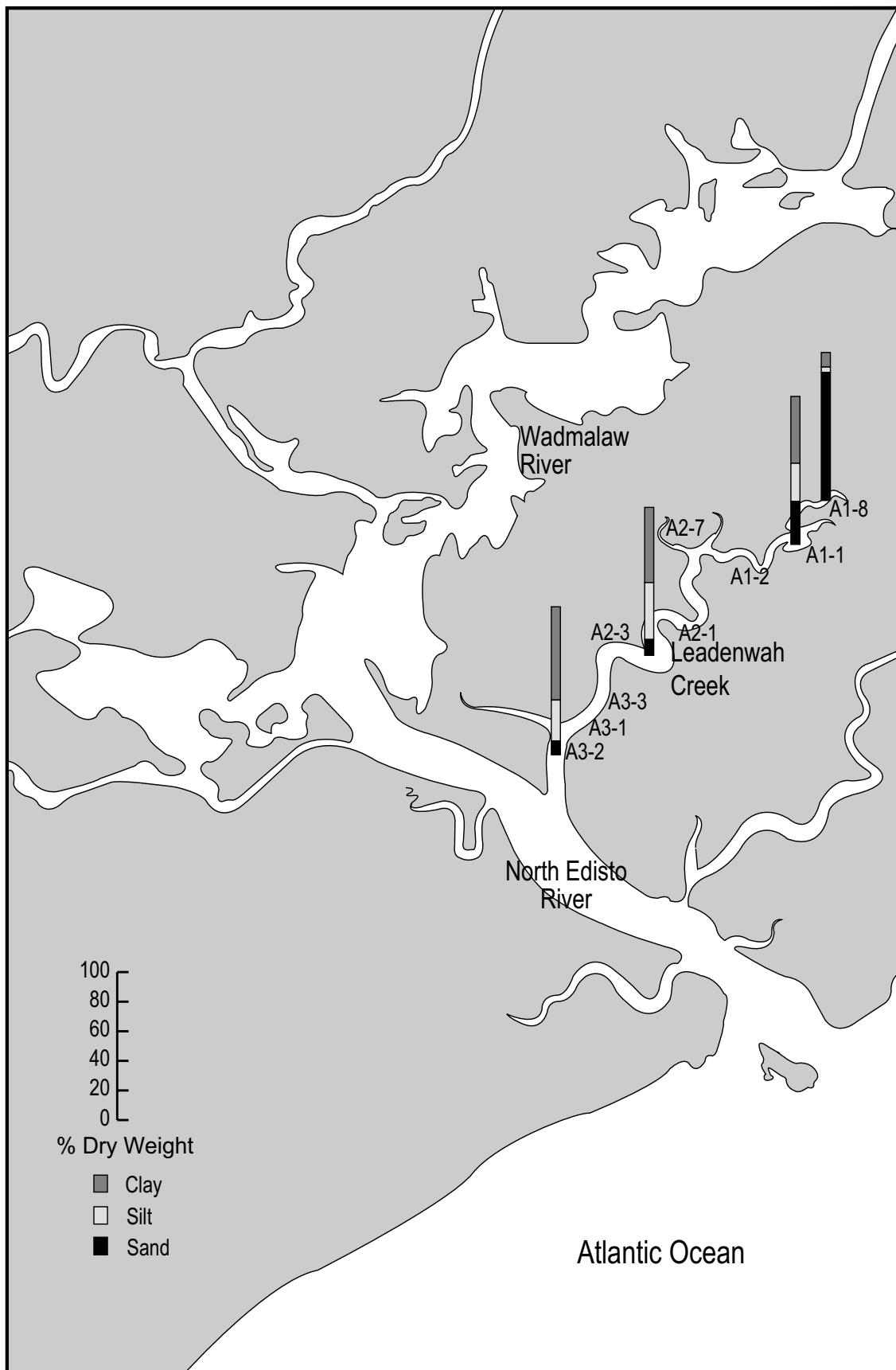


Figure 31. Percent sand, silt, and clay in sediments from Leadenwah Creek.

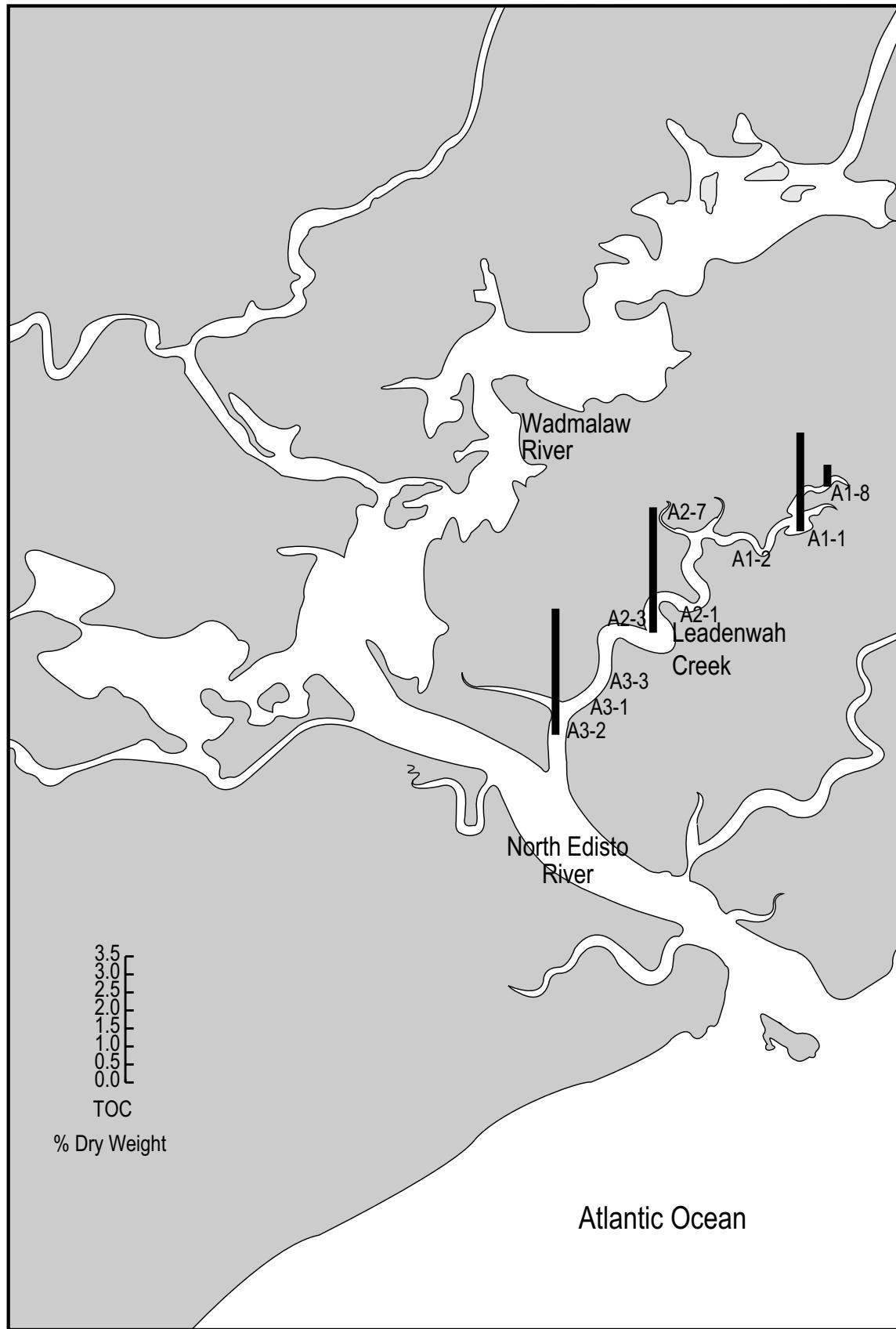


Figure 32. Concentrations of total organic carbon in sediments from Leadenwah Creek.

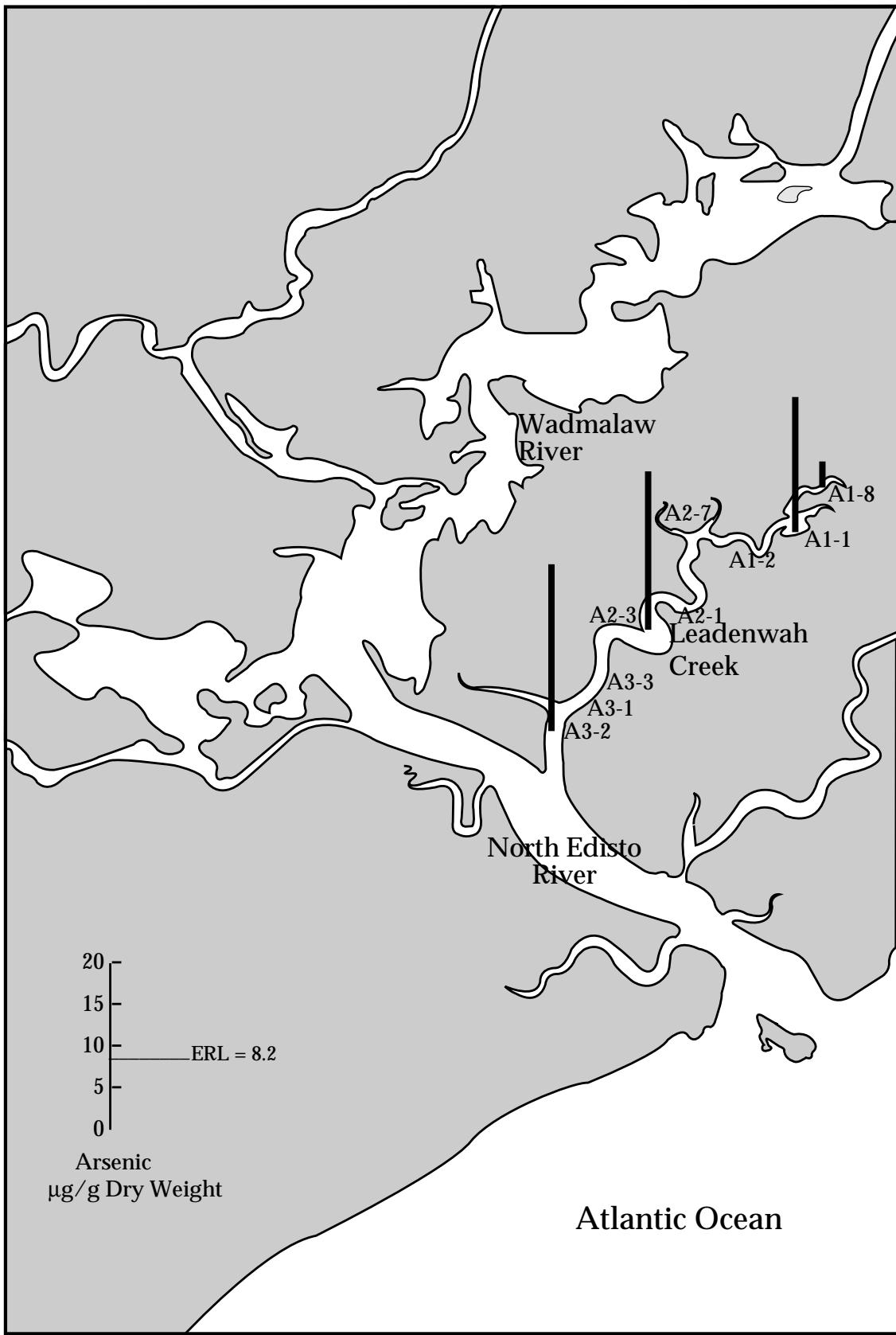


Figure 33. Concentrations of arsenic in sediments from Leadenwah Creek.

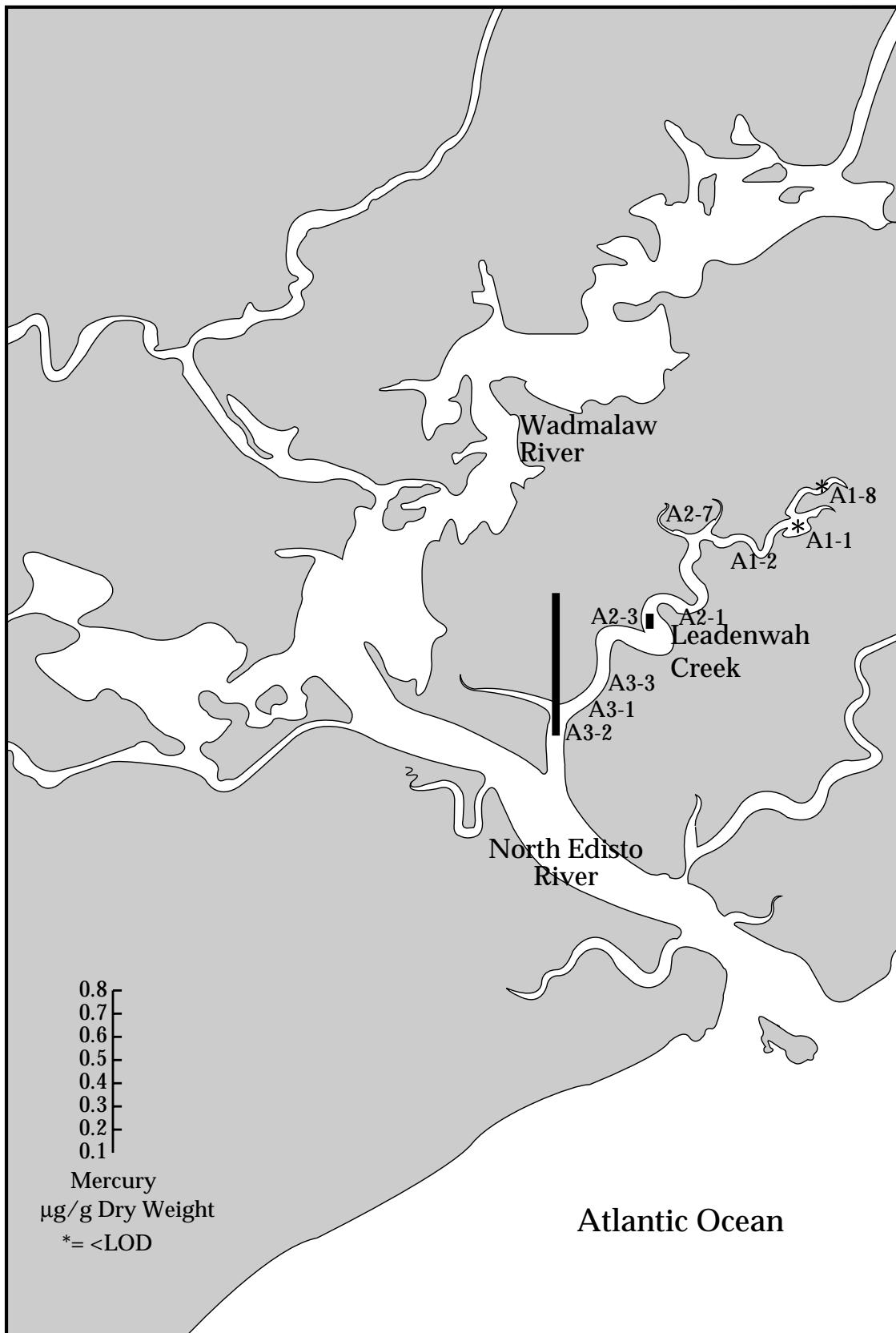


Figure 34. Concentrations of mercury in sediments from Leadenwah Creek.

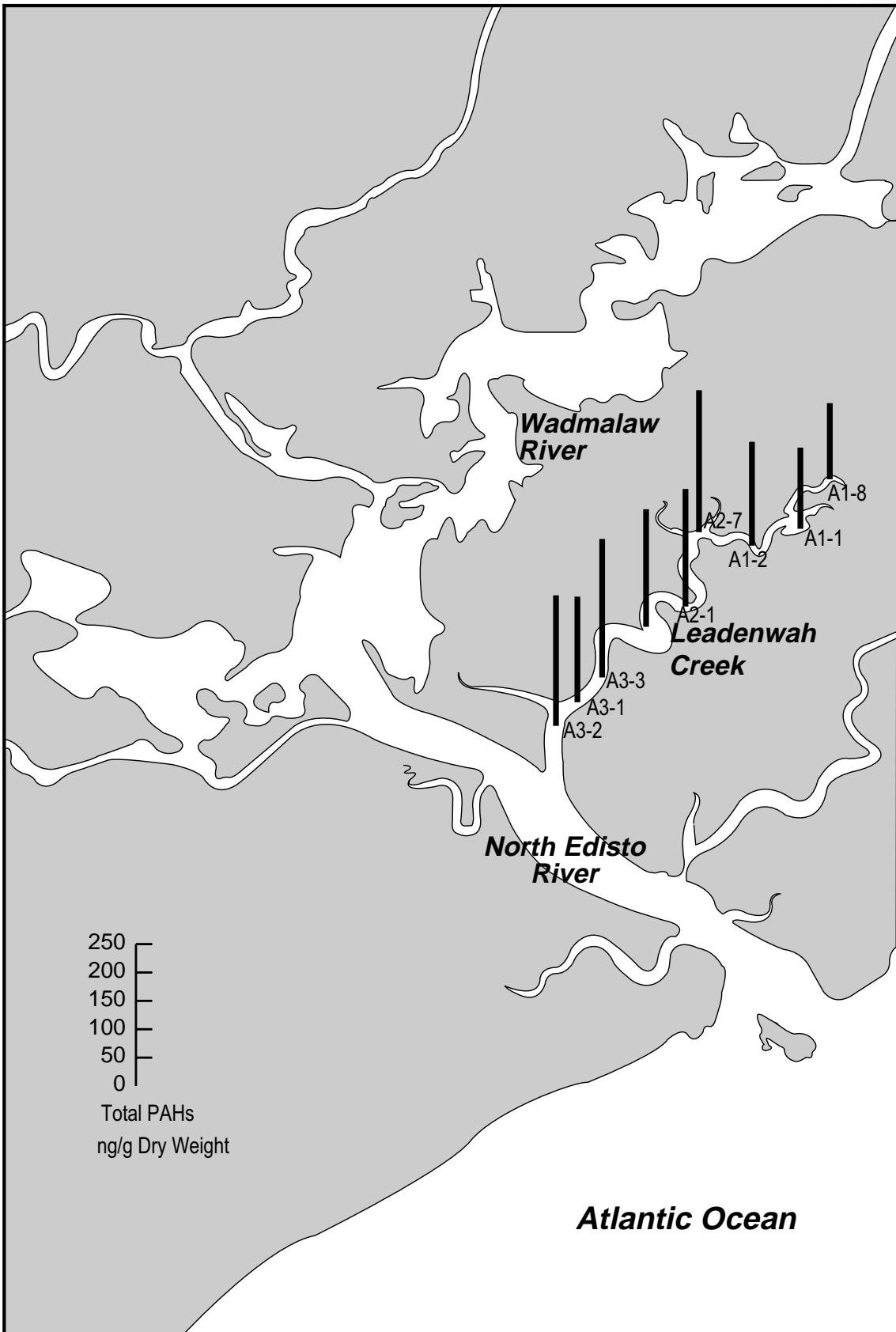


Figure 35. Concentrations of total PAHs in sediments from Leadenwah Creek.

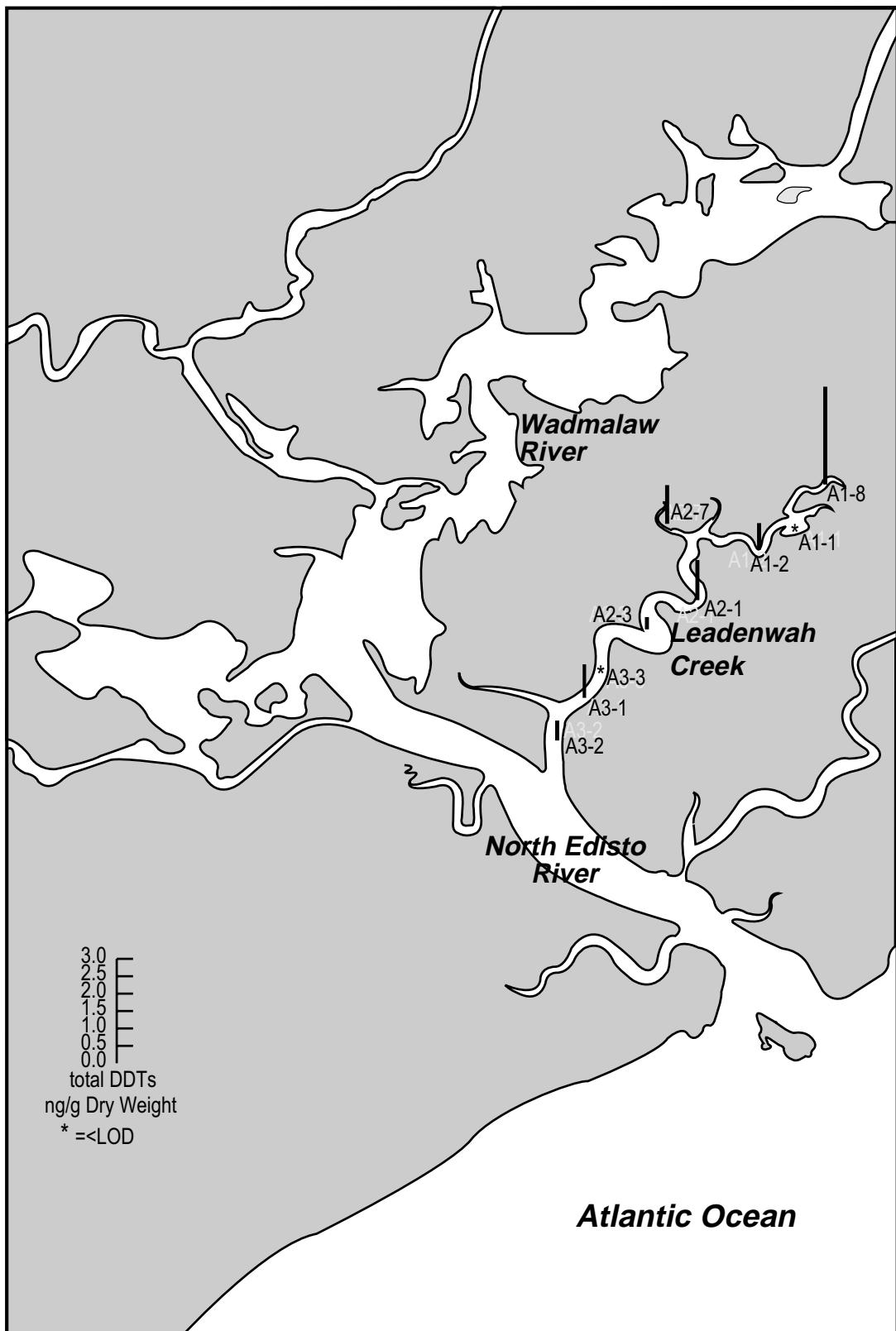


Figure 36. Concentrations of total DDTs in sediments from Leadenwah Creek.

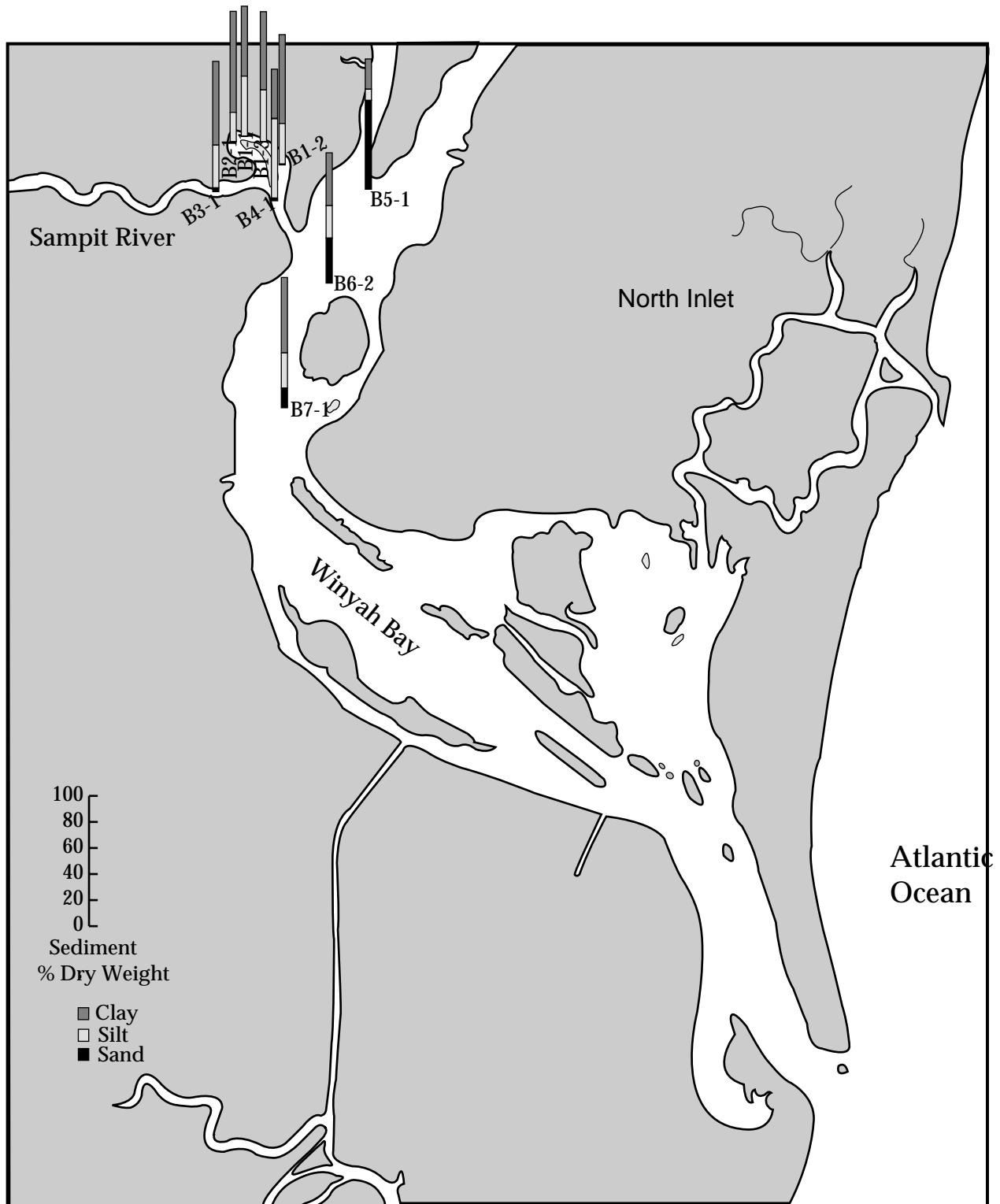


Figure 37. Concentrations of sand, silt and clay in sediments from Winyah Bay.

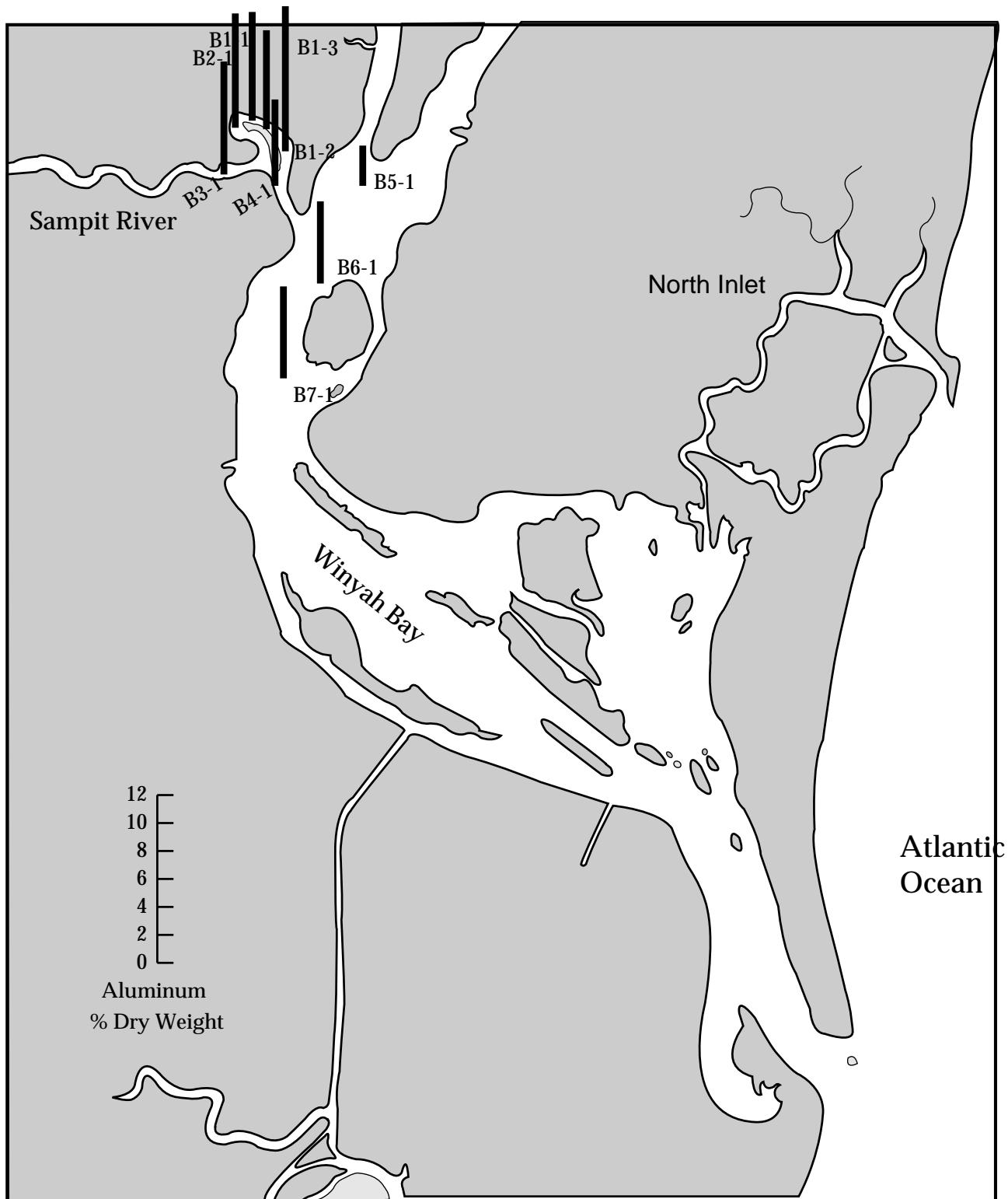


Figure 38. Concentrations of aluminum in sediments from Winyah Bay.

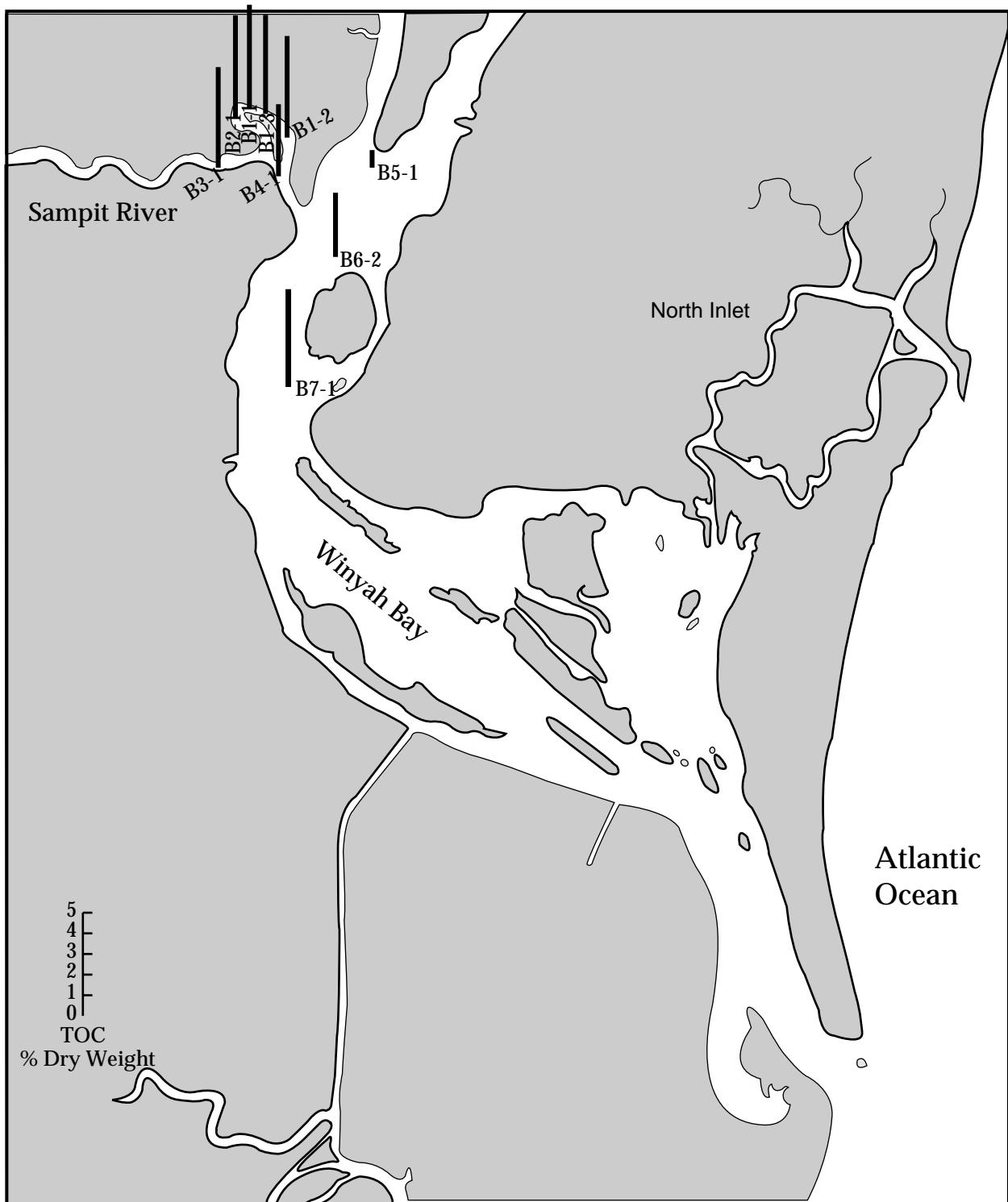


Figure 39. Concentrations of total organic carbon in sediments from Winyah Bay.

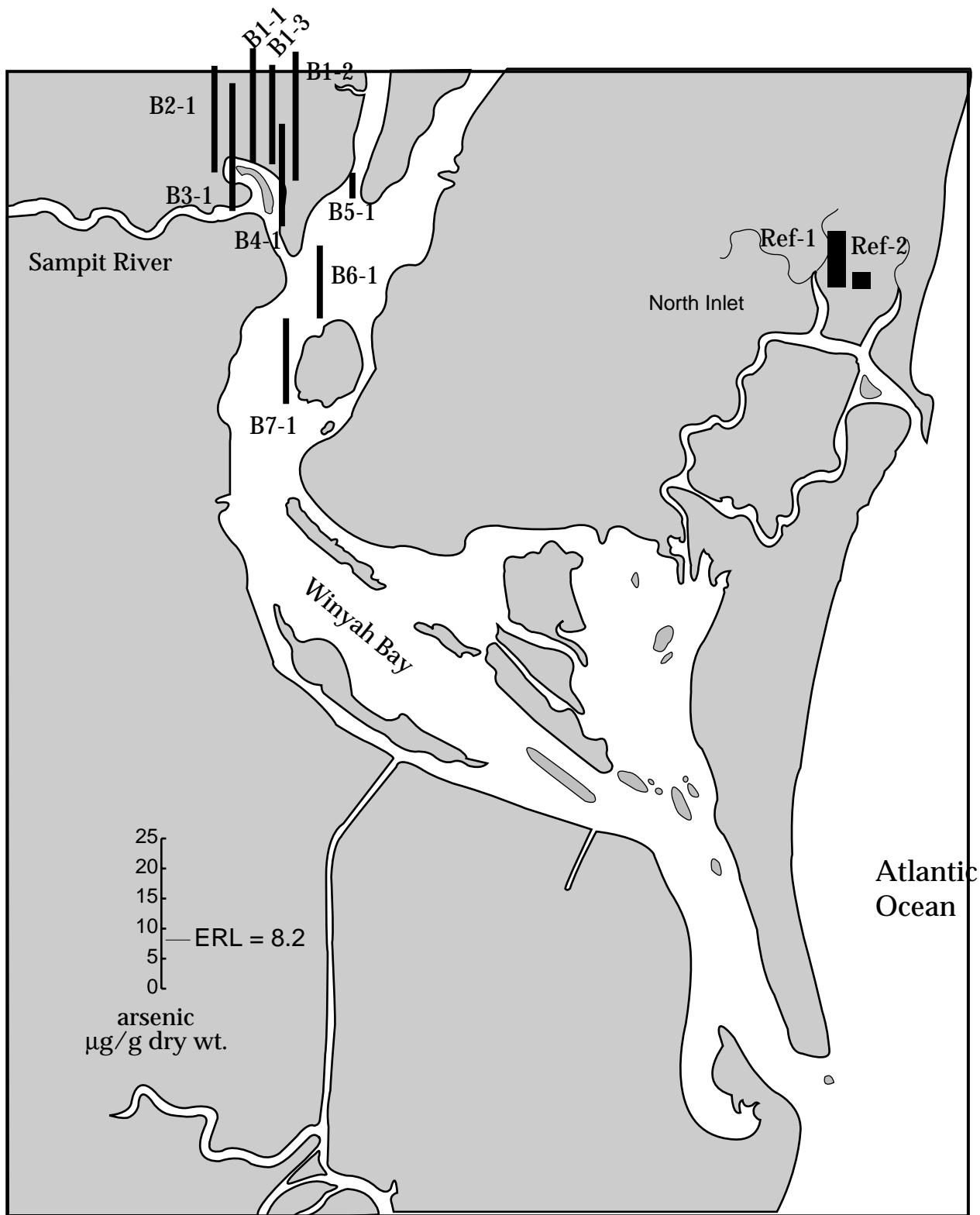


Figure 40. Concentrations of arsenic in sediments from Winyah Bay.

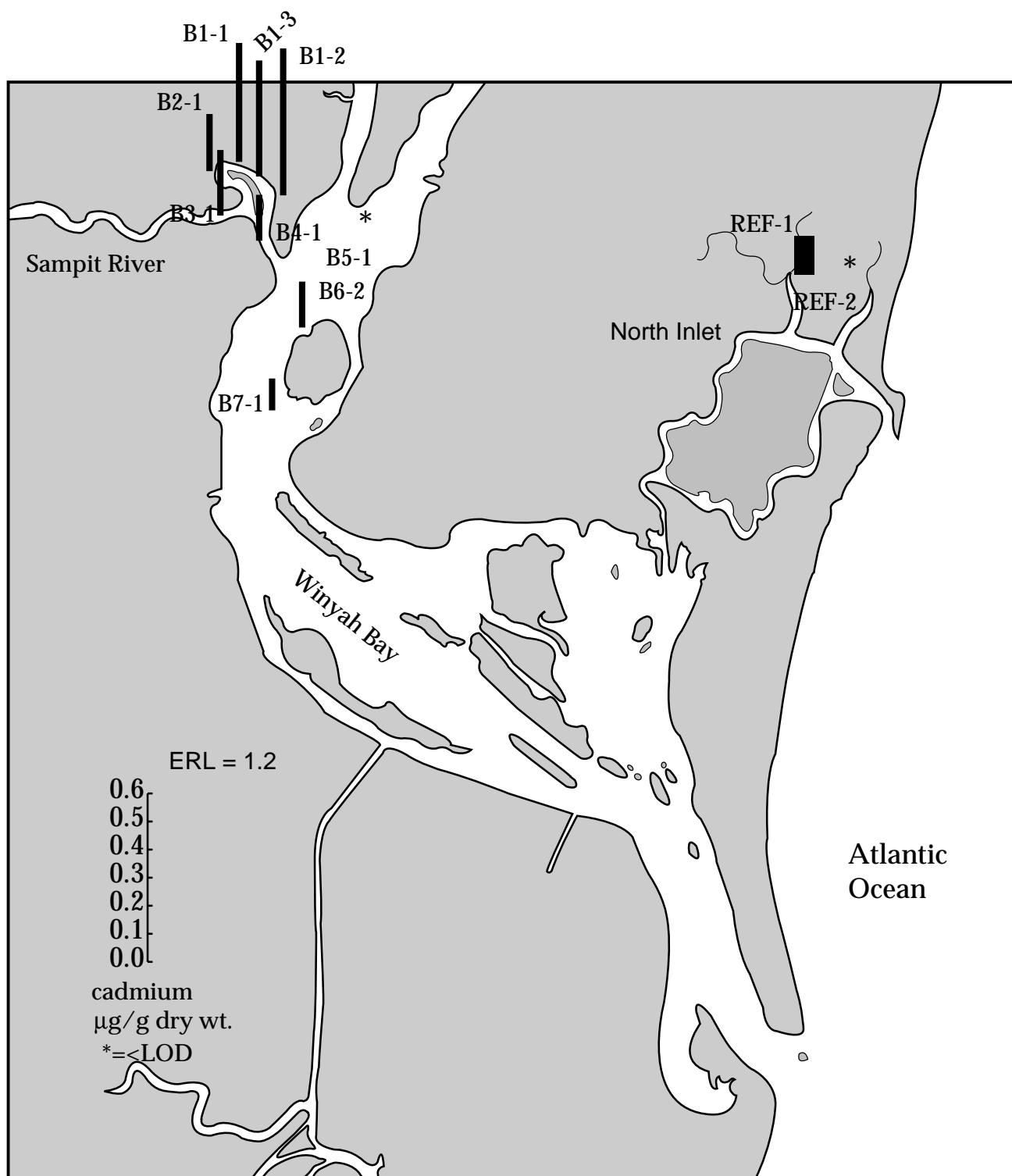


Figure 41. Concentrations of cadmium in sediments from Winyah Bay.

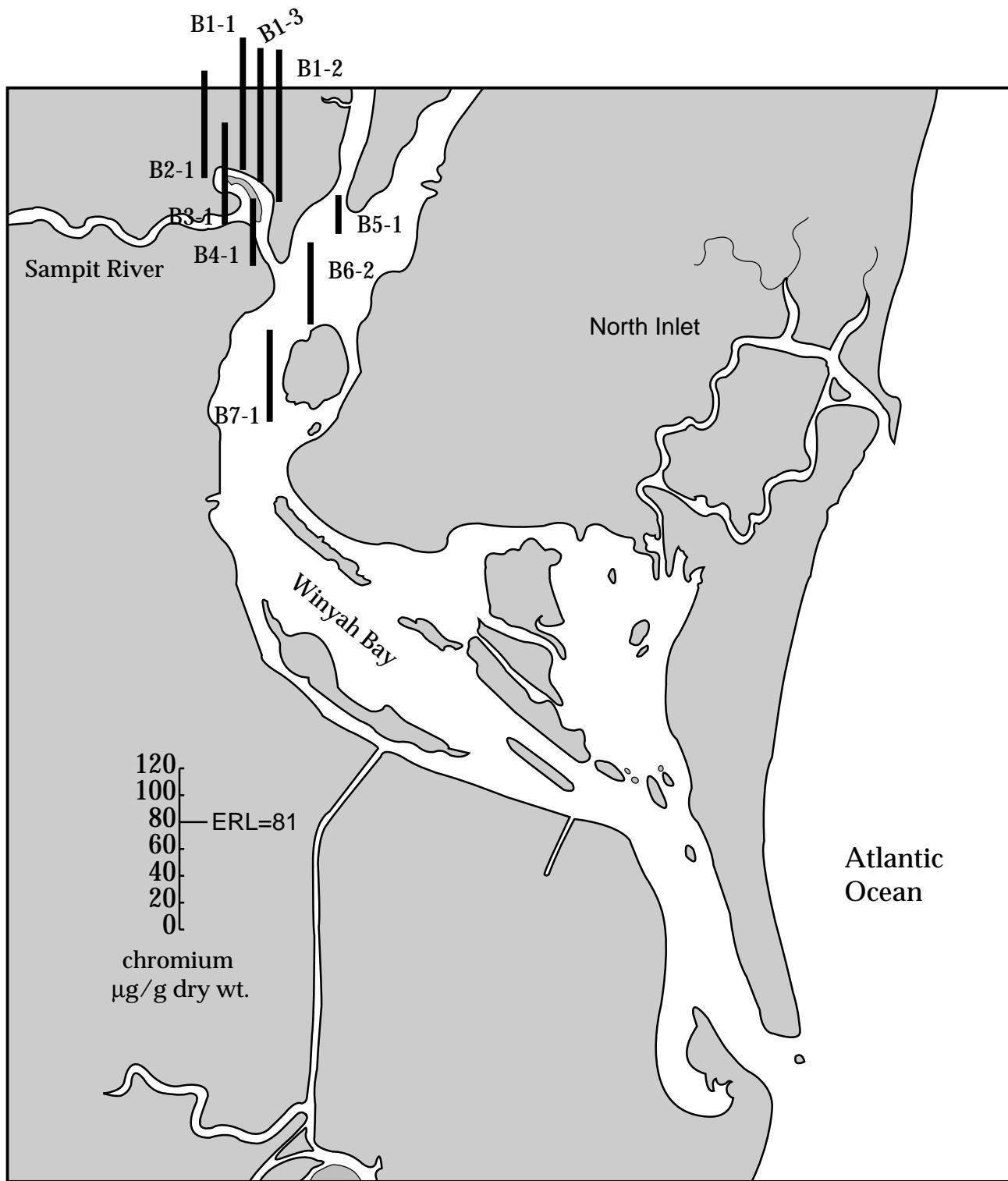


Figure 42. Concentrations of chromium in sediments from Winyah Bay.

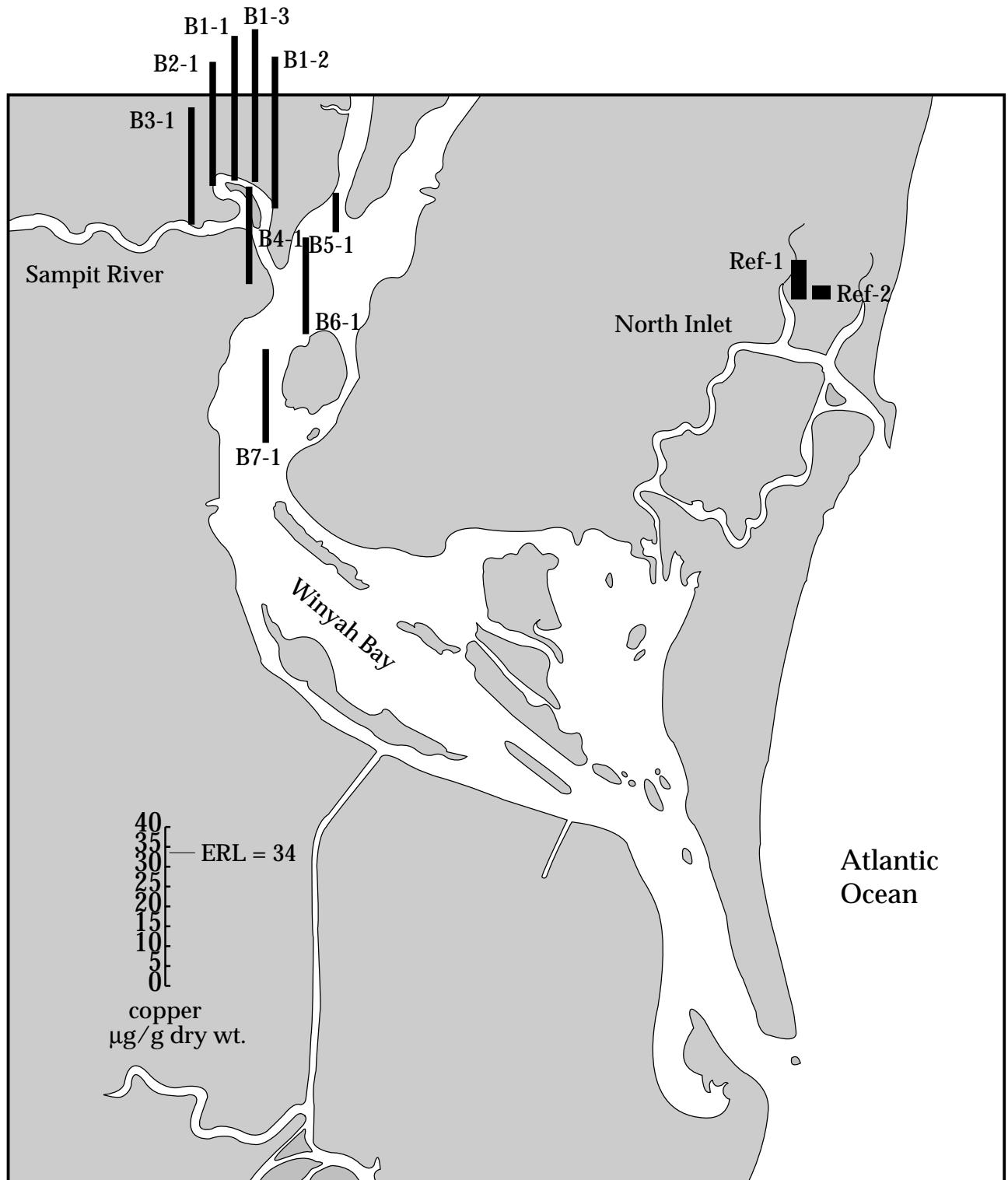


Figure 43. Concentrations of copper in sediments from Winyah Bay.

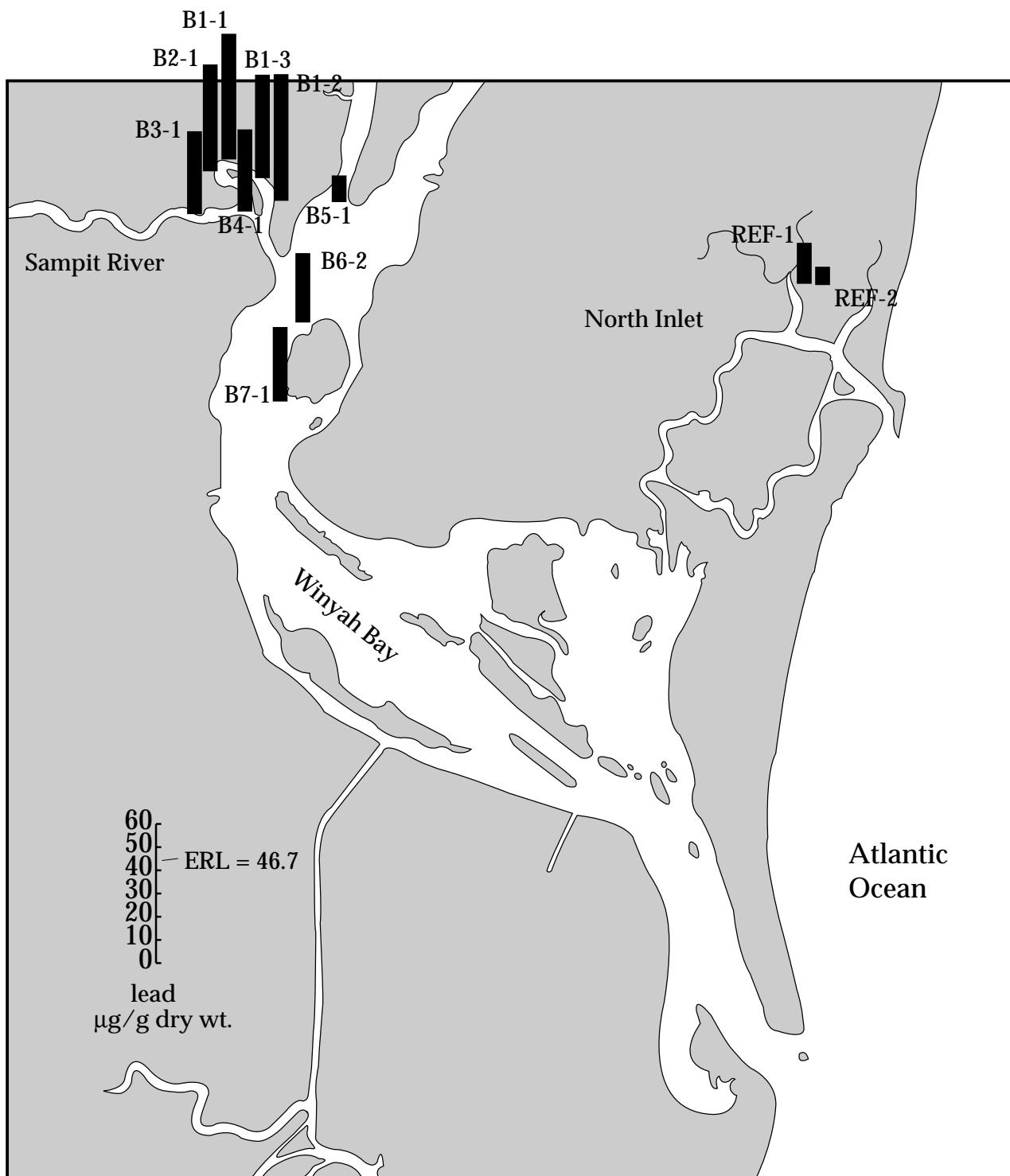


Figure 44. Concentrations of lead in sediments from Winyah Bay.

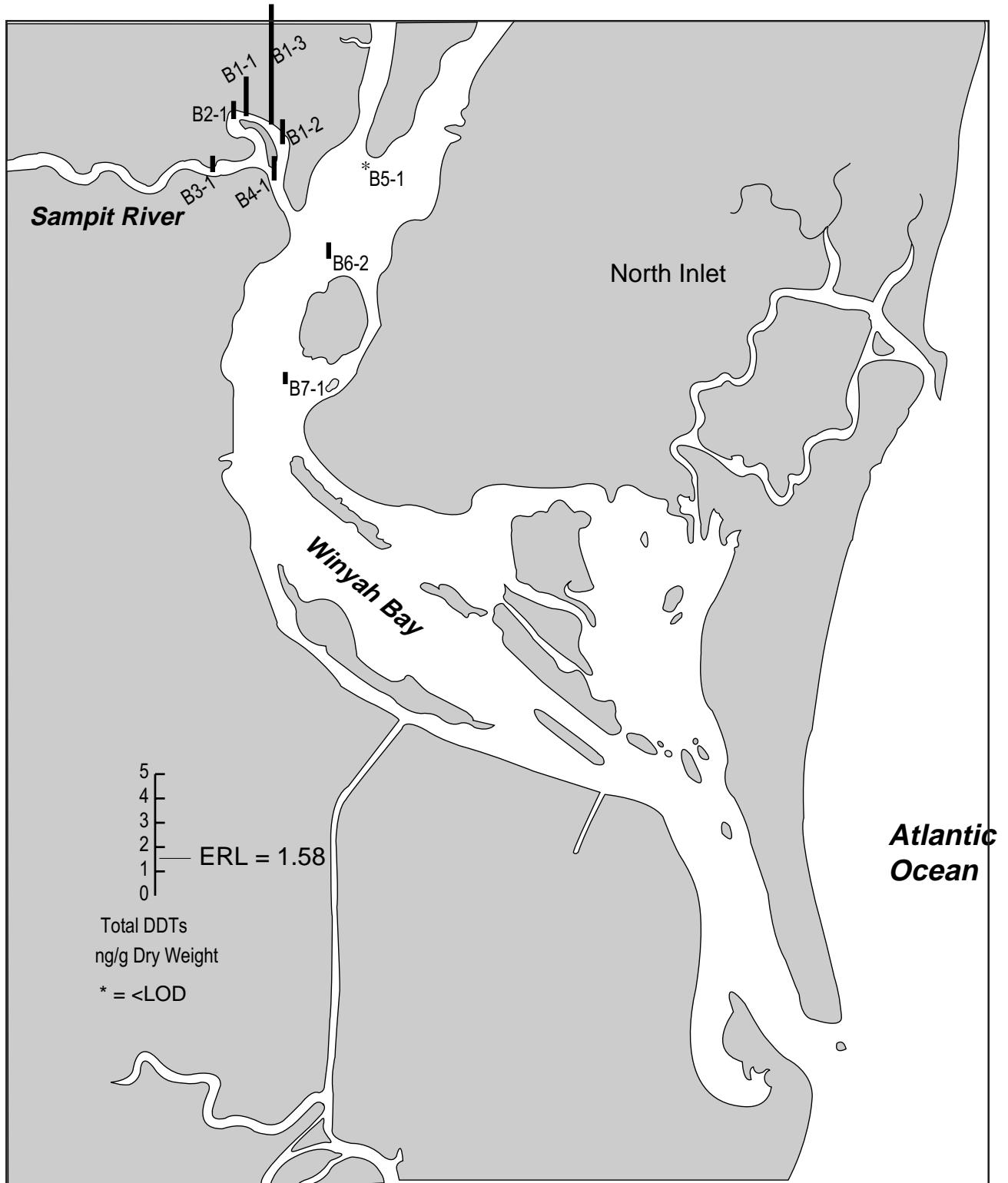


Figure 45. Concentrations of total DDTs in sediments from Winyah Bay.

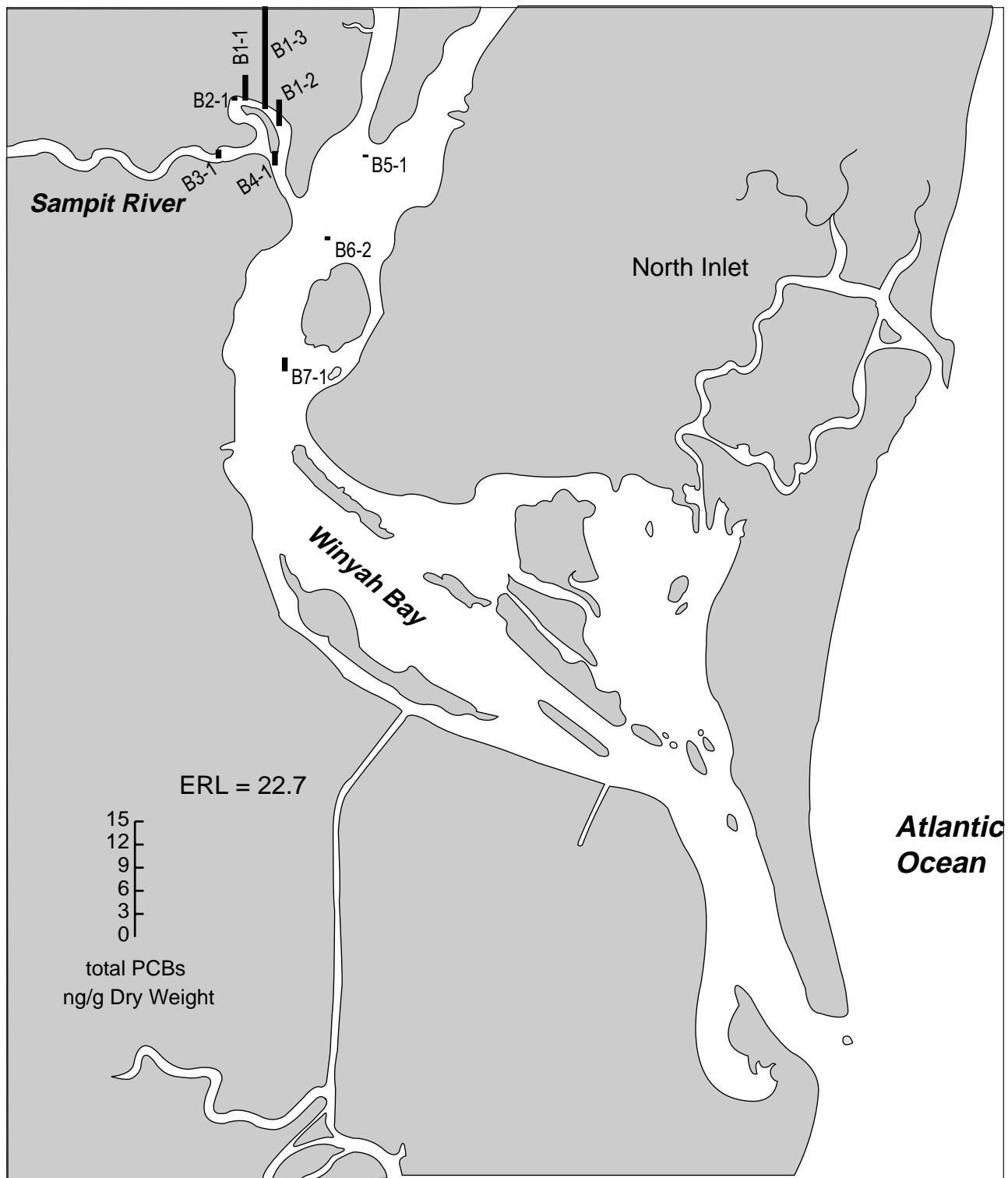


Figure 46. Concentrations of total PCBs in sediments from Winyah Bay.

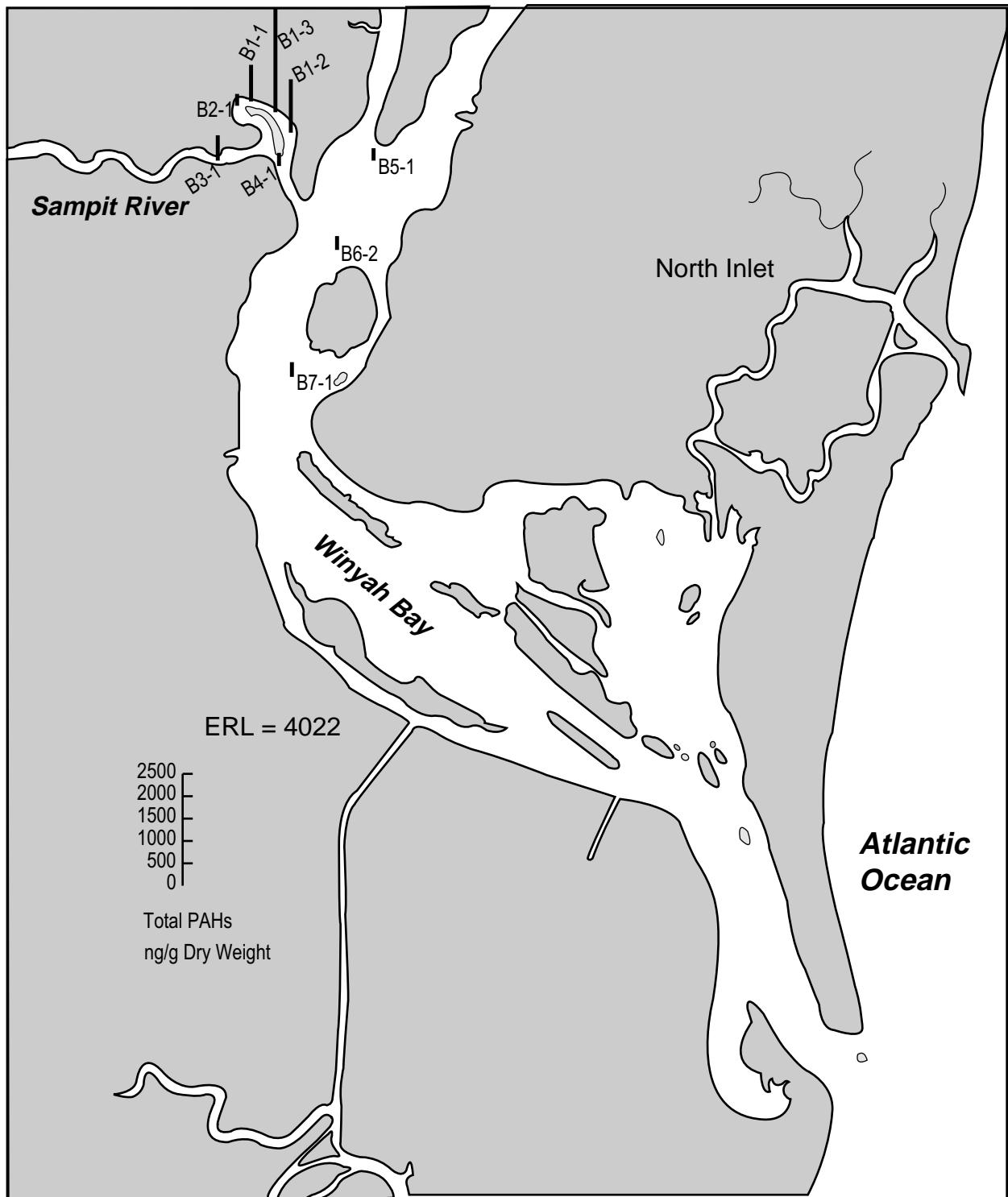


Figure 47. Concentrations of total PAHs in sediments from Winyah Bay.

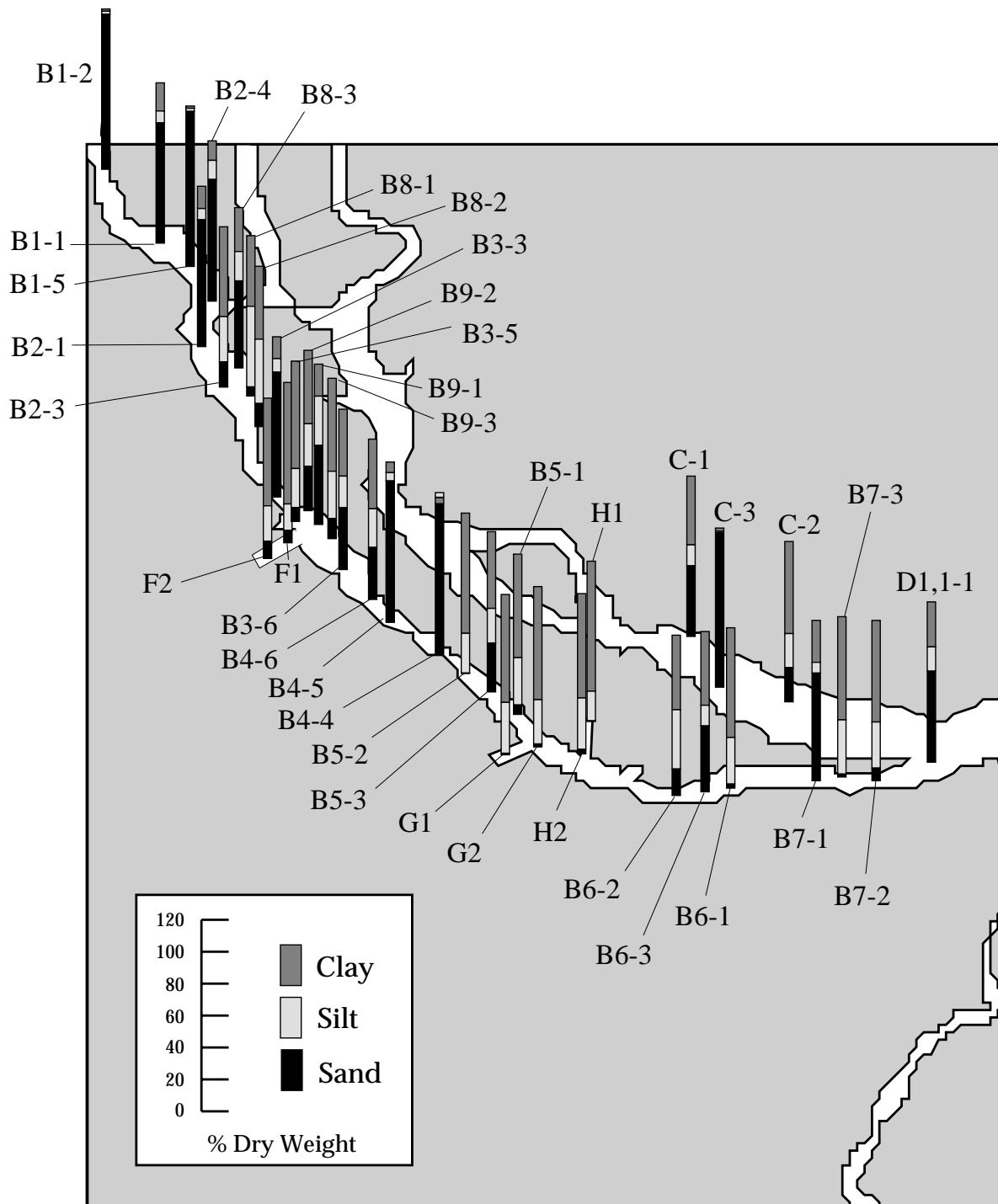


Figure 48. Concentrations of sand, silt, and clay in sediments from the upper Savannah River channel.

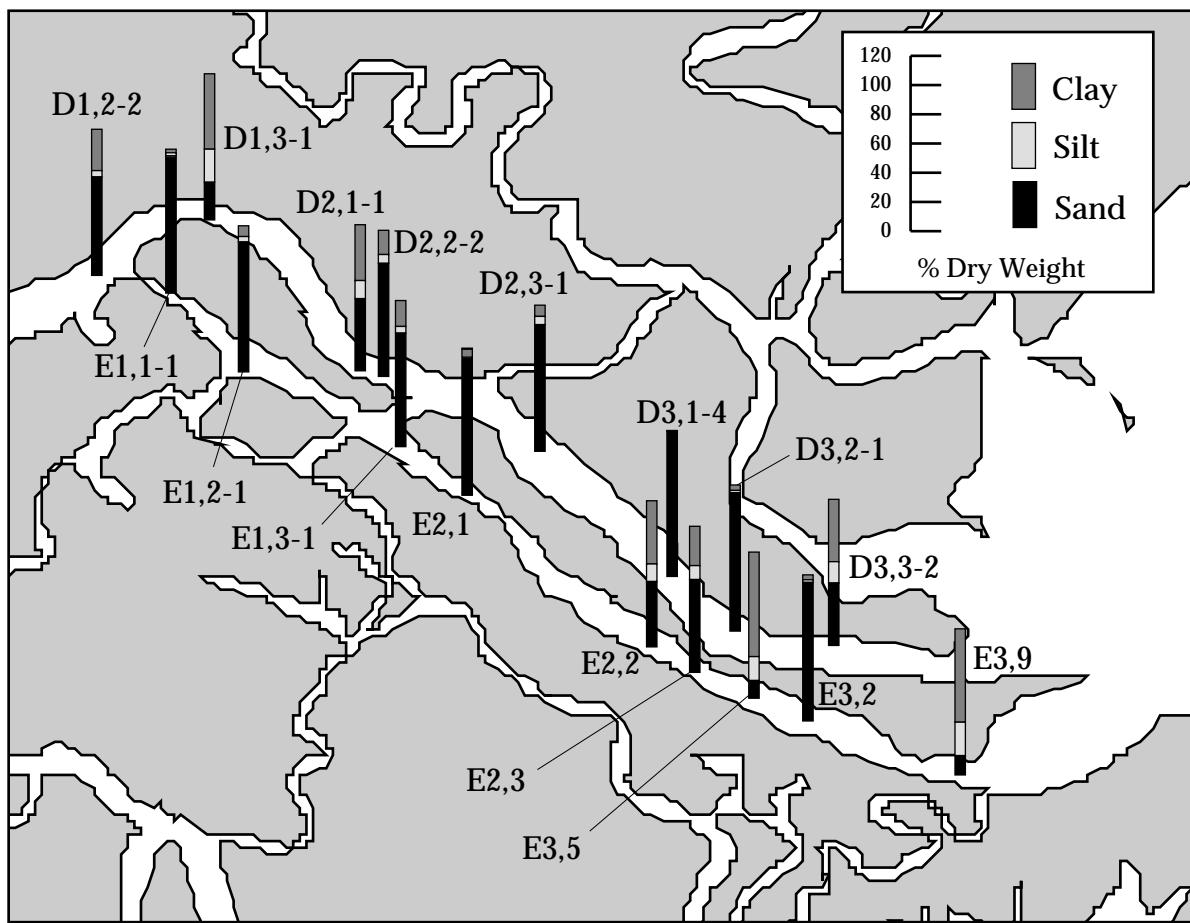


Figure 49. Concentrations of sand, silt, and clay in sediments from the lower Savannah River channel.

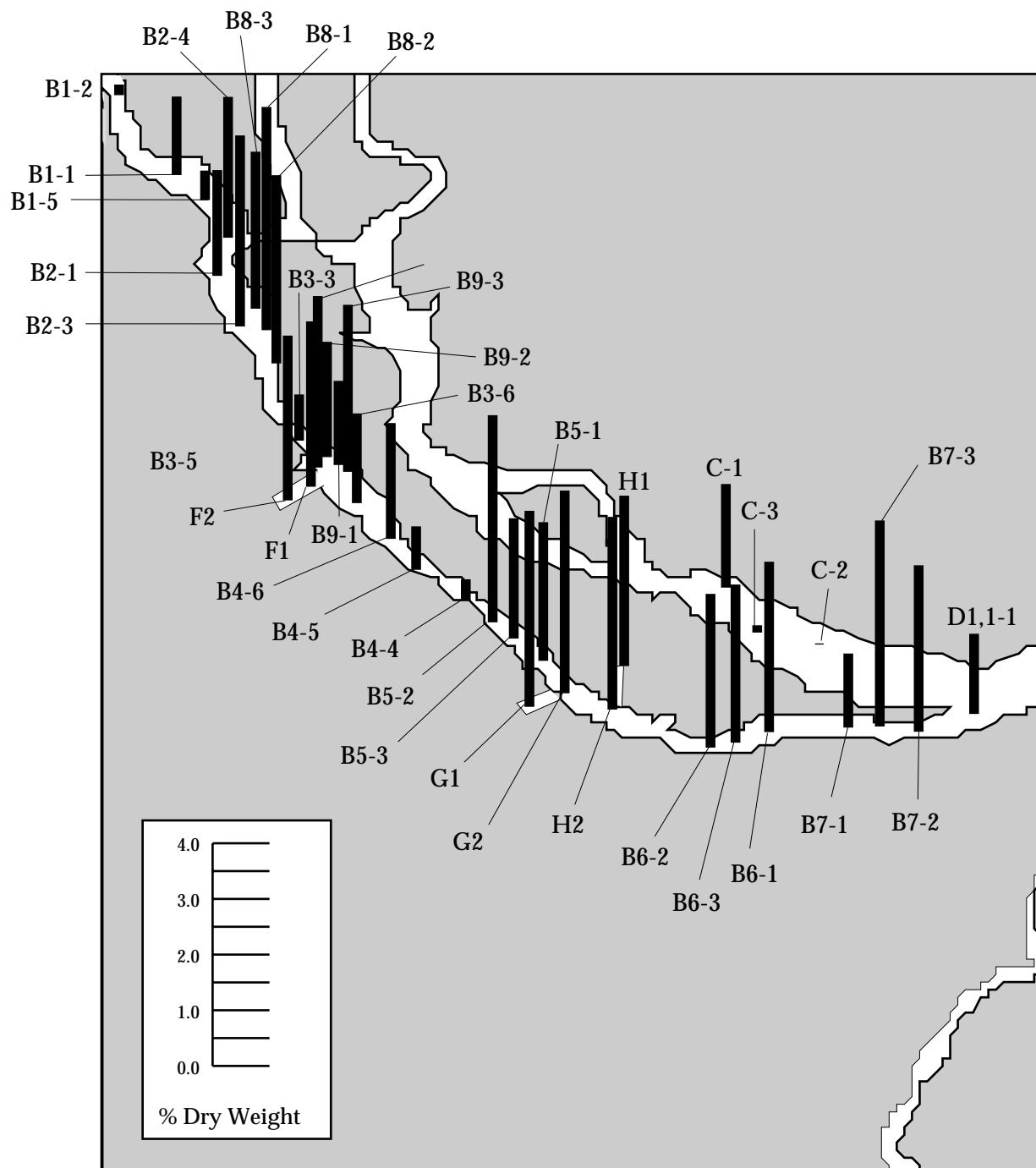


Figure 50. Concentrations of total organic carbon in sediments from the upper Savannah River channel.

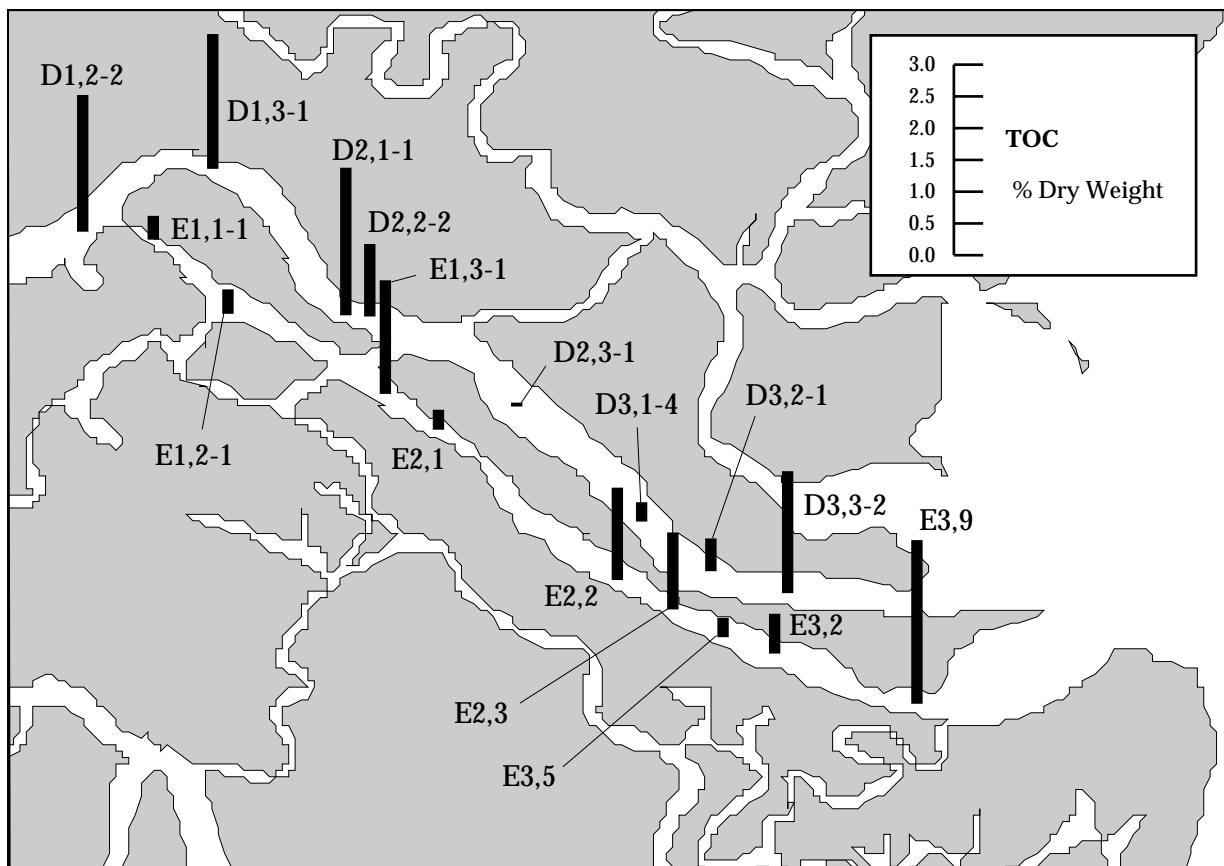


Figure 51. Concentrations of total organic carbon in sediments from the lower Savannah River channel.

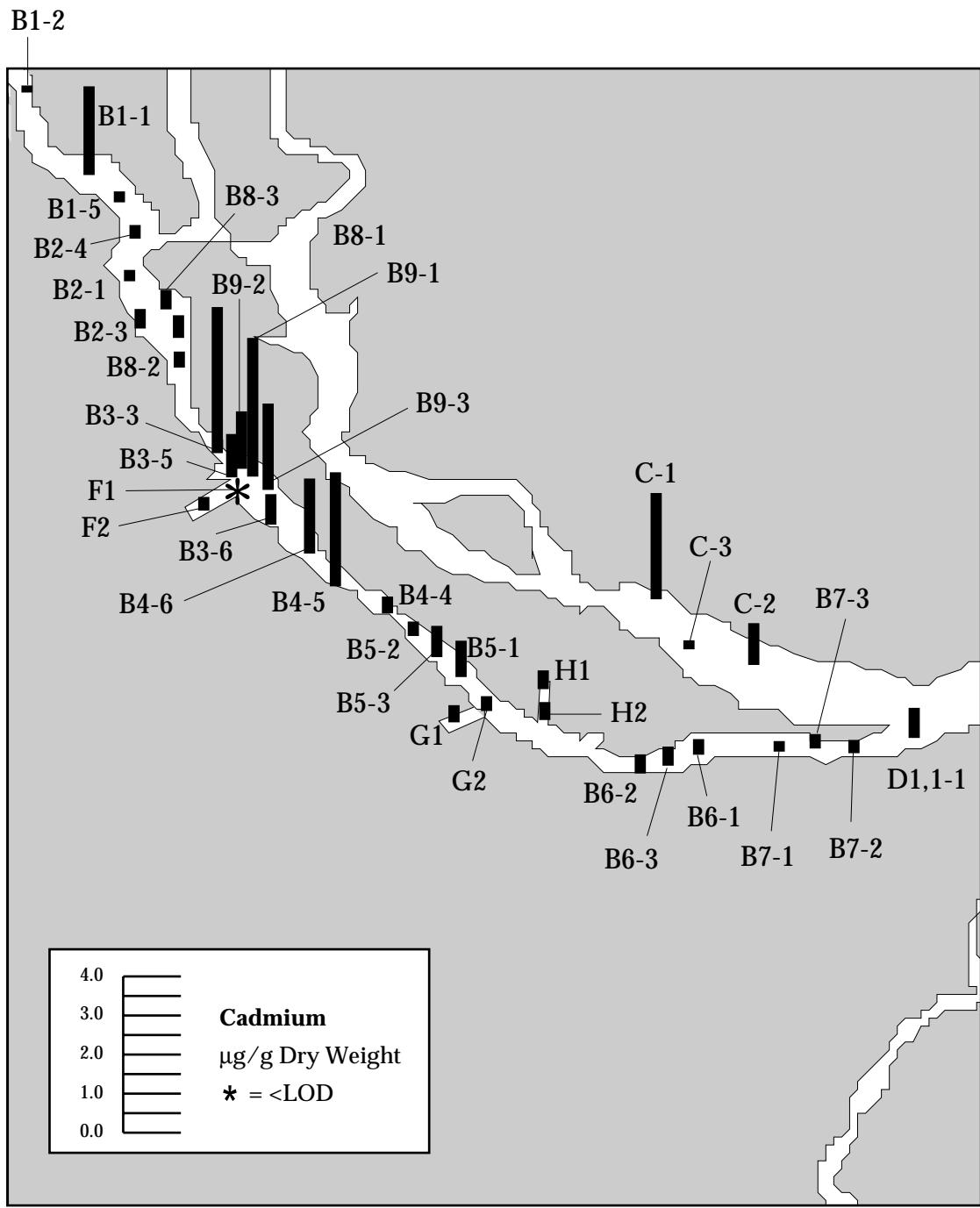


Figure 52. Concentrations of cadmium in sediments from the upper Savannah River channel.

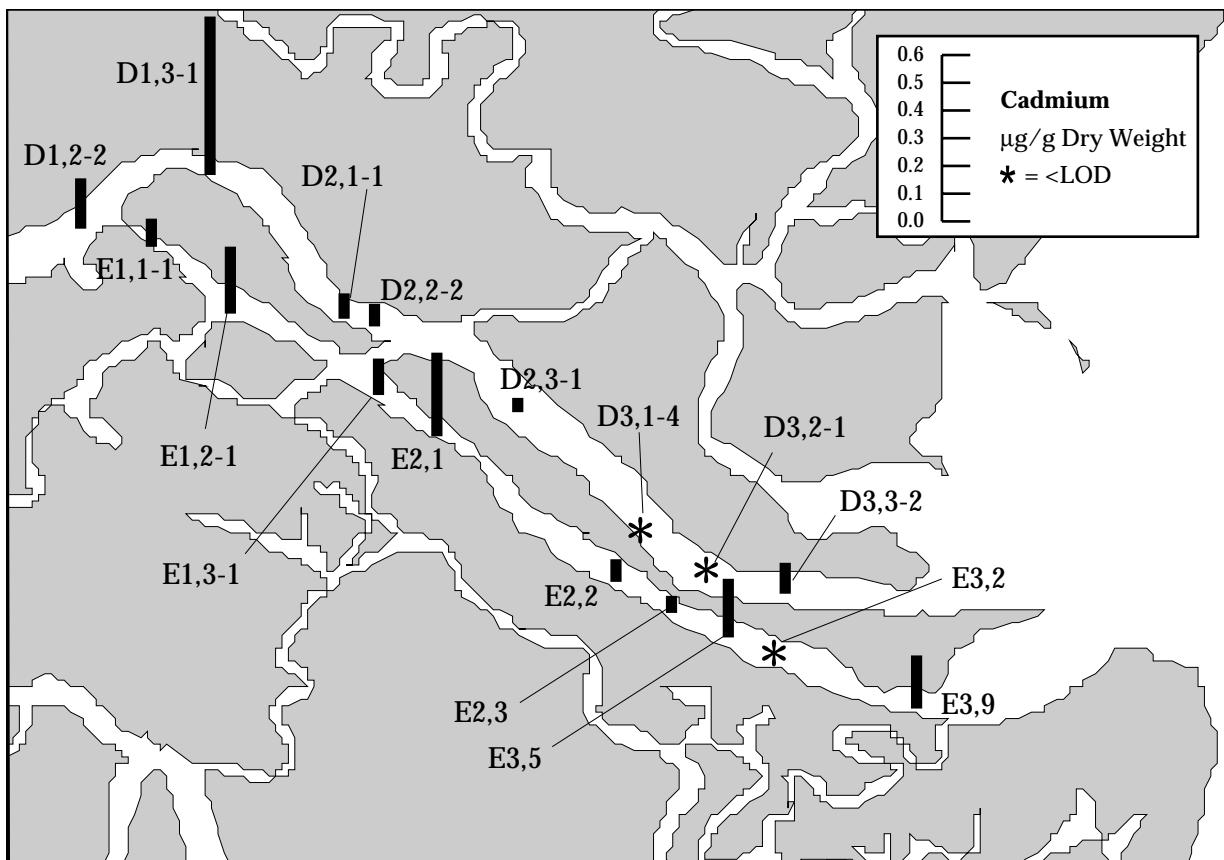


Figure 53. Concentrations of cadmium in sediments from the lower Savannah River channel.

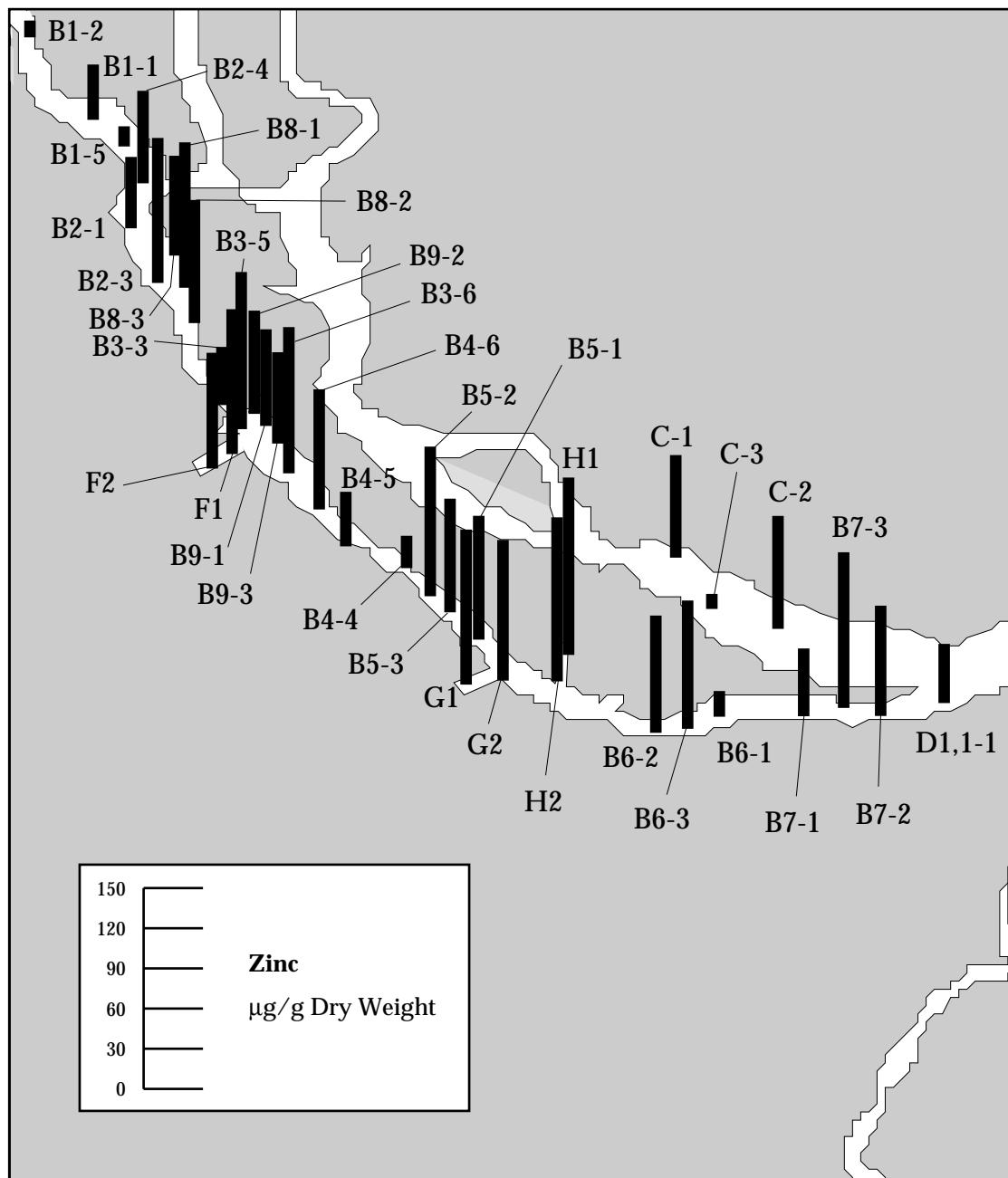


Figure 54. Concentrations of zinc in sediments from the upper Savannah River channel.

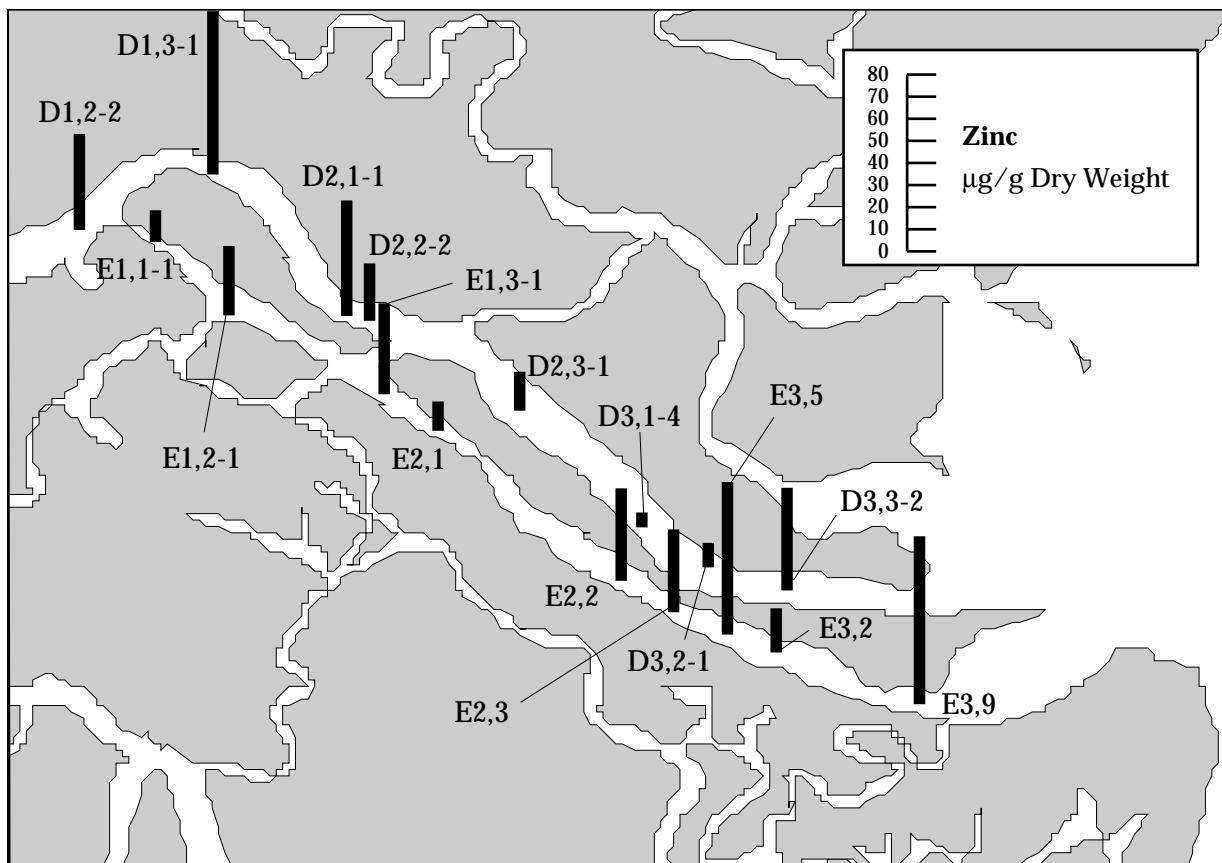


Figure 55. Concentrations of zinc sediments from the lower Savannah River channel.

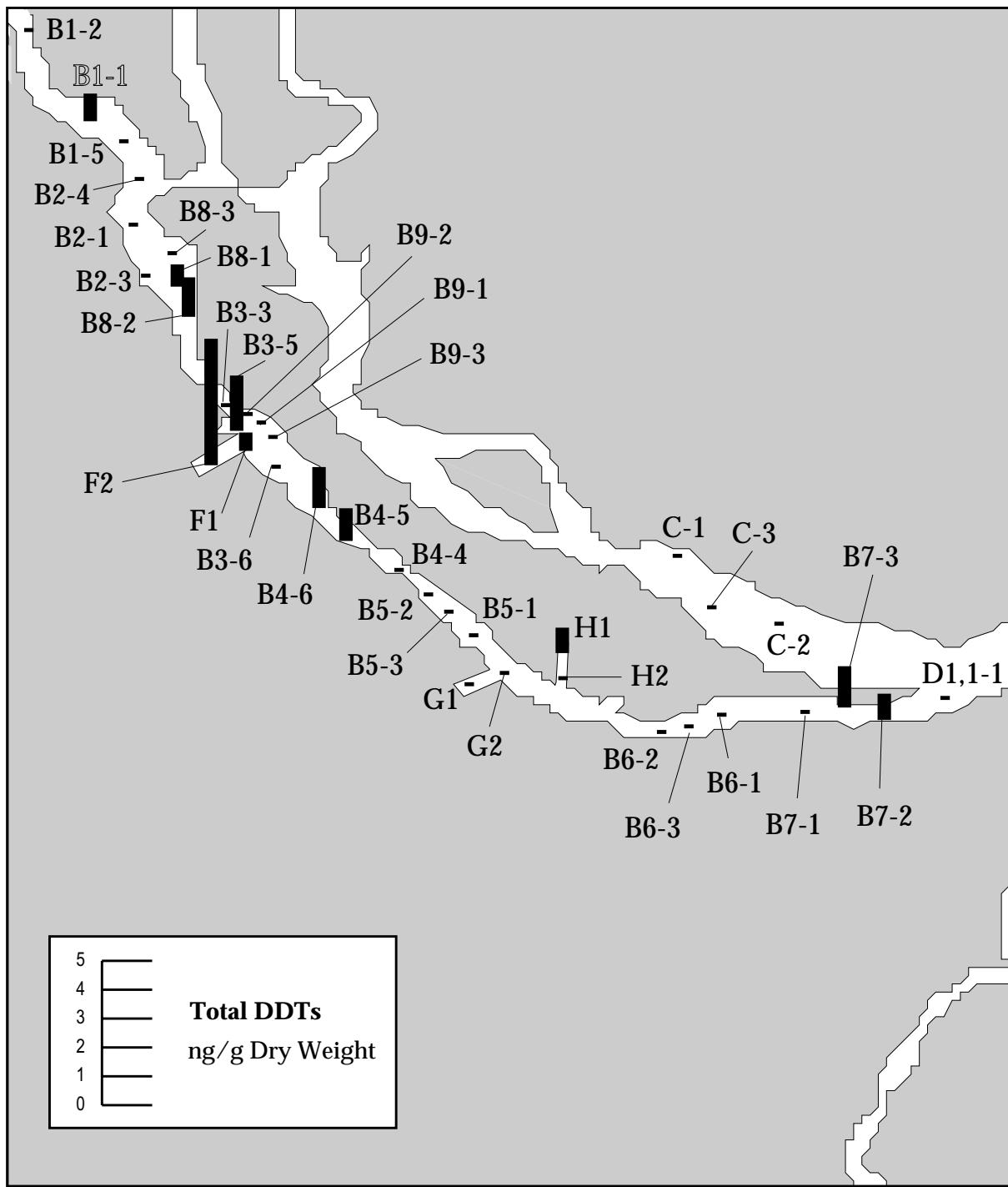


Figure 56. Concentrations of total DDTs in sediments from the upper Savannah River channel.

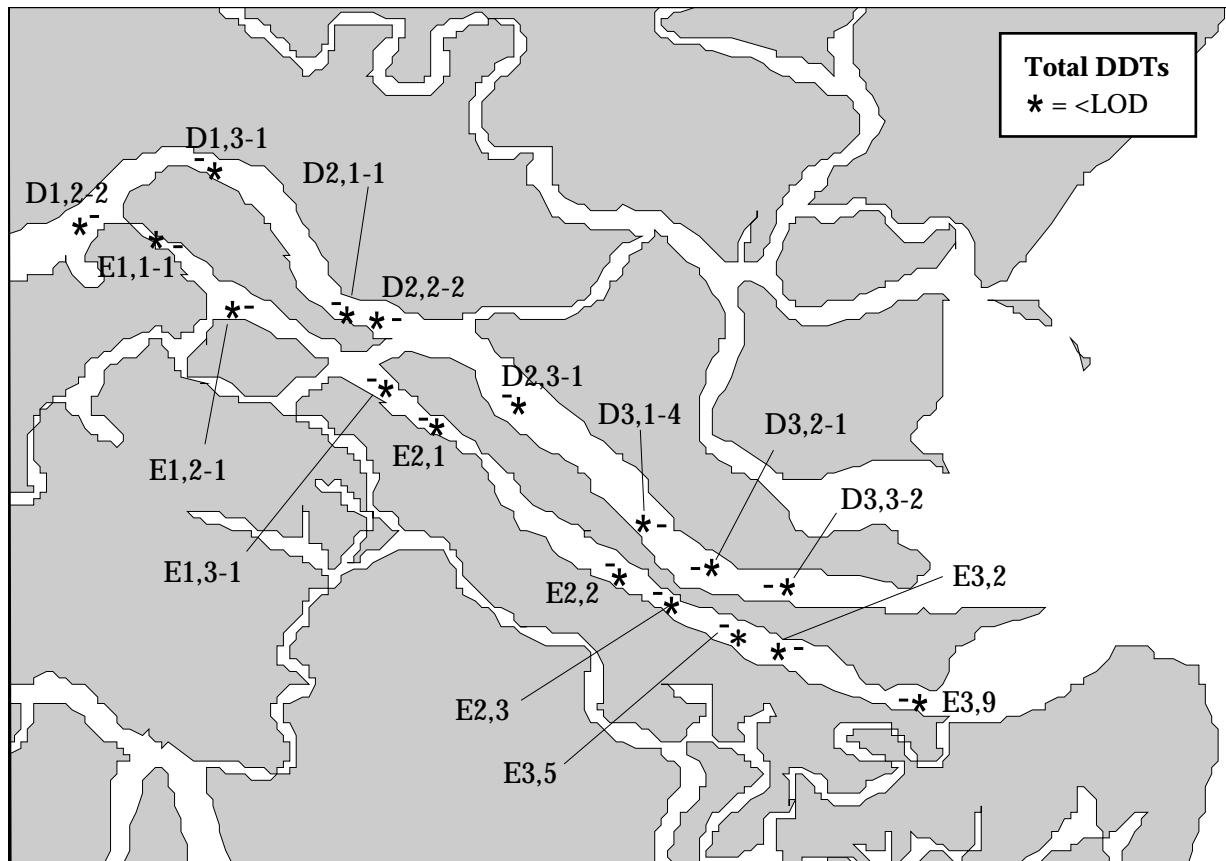


Figure 57. Concentrations of total DDTs in sediments from the lower Savannah River channel.

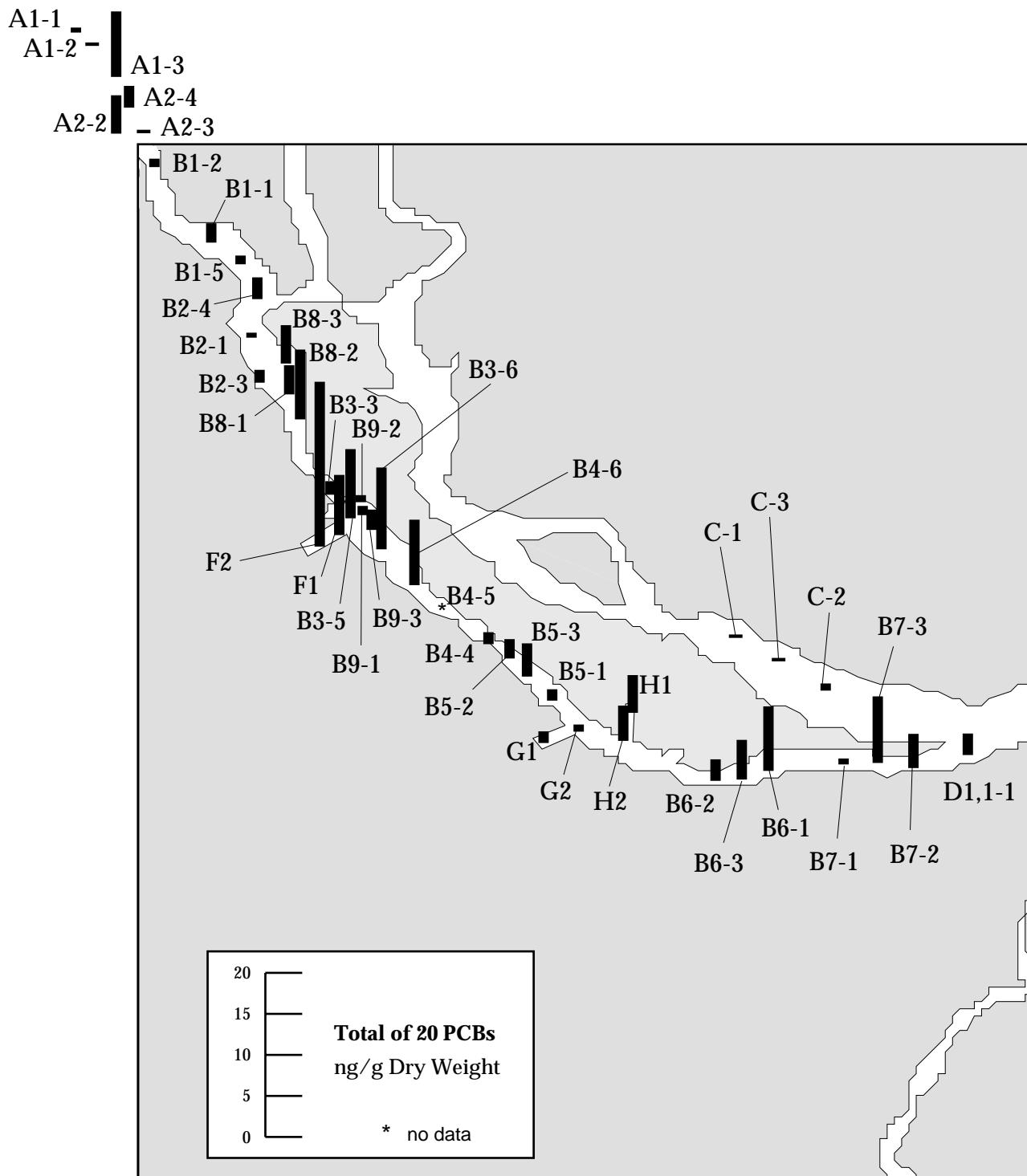


Figure 58. Total concentrations of 20 PCB congeners in sediments from the upper Savannah River channel.

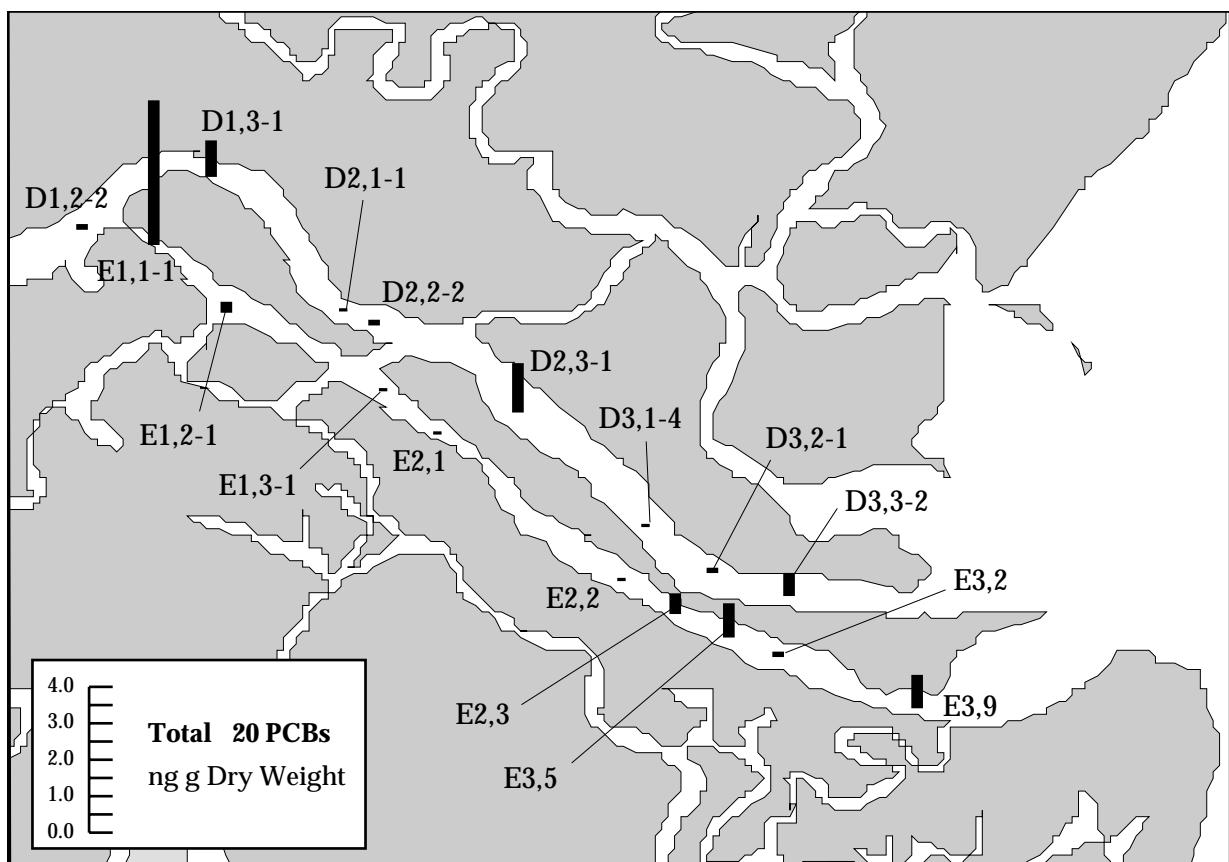


Figure 59. Total concentrations of total 20 PCB congeners in sediment from the lower Savannah River channel.

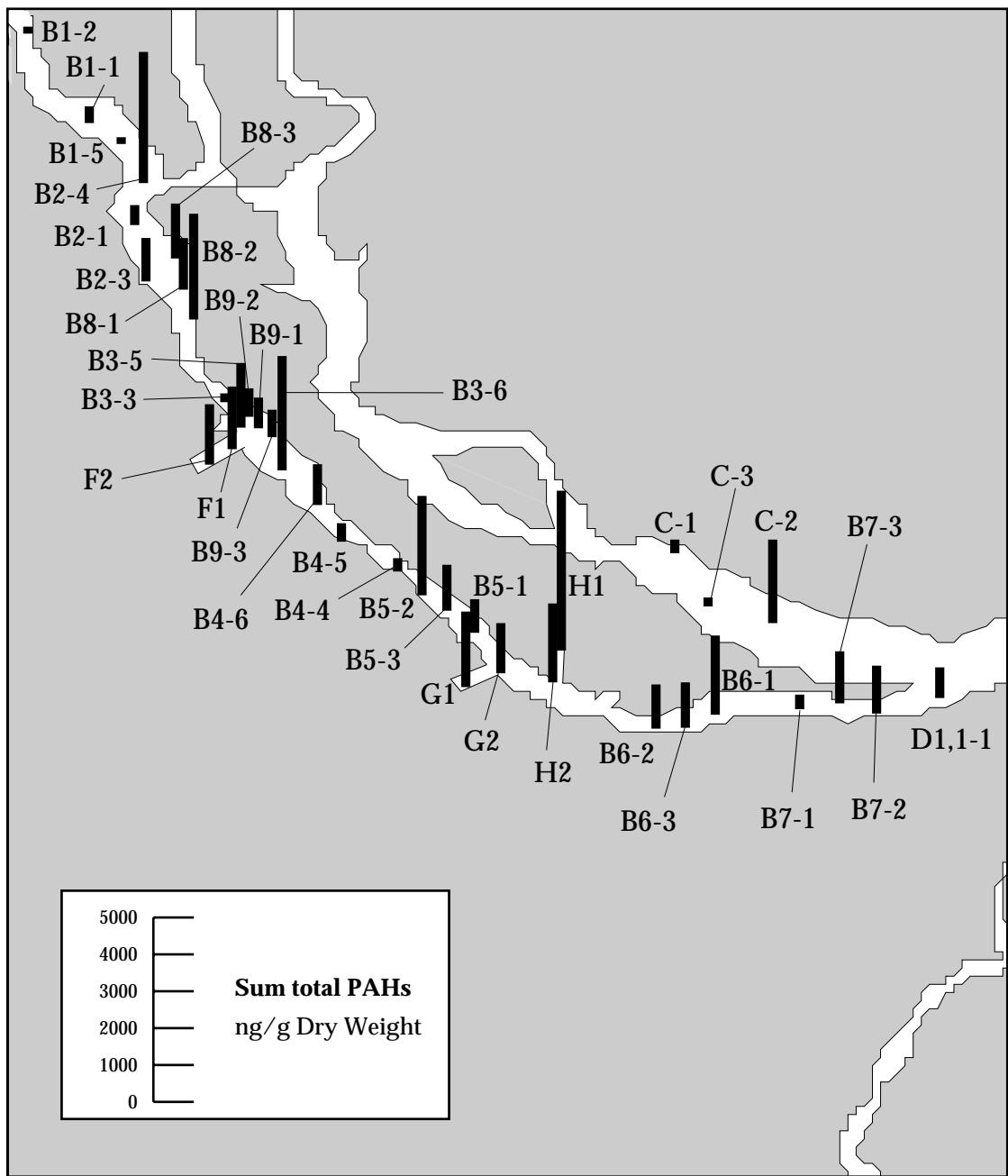


Figure 60. Concentrations of total PAH in sediments from the upper Savannah River channel.

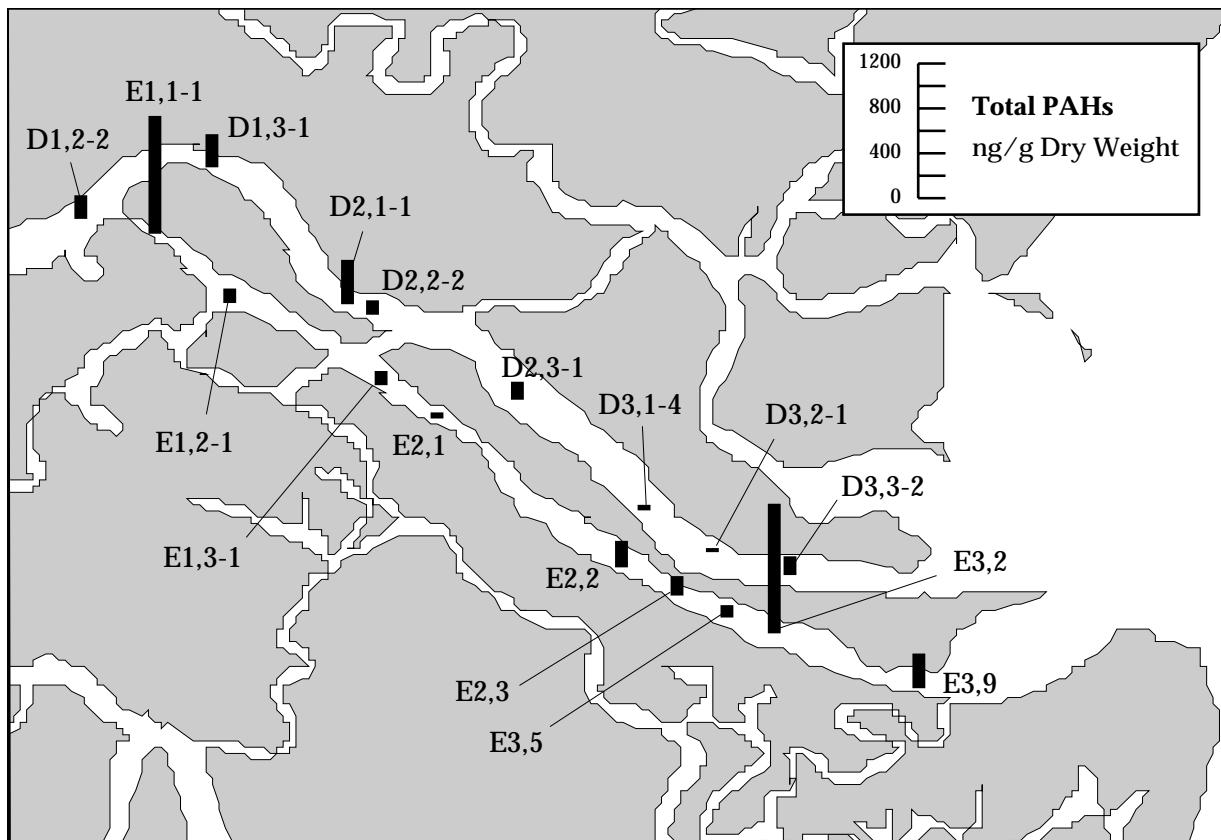


Figure 61. Concentrations of total PAHs in sediments from the lower Savannah River channel.

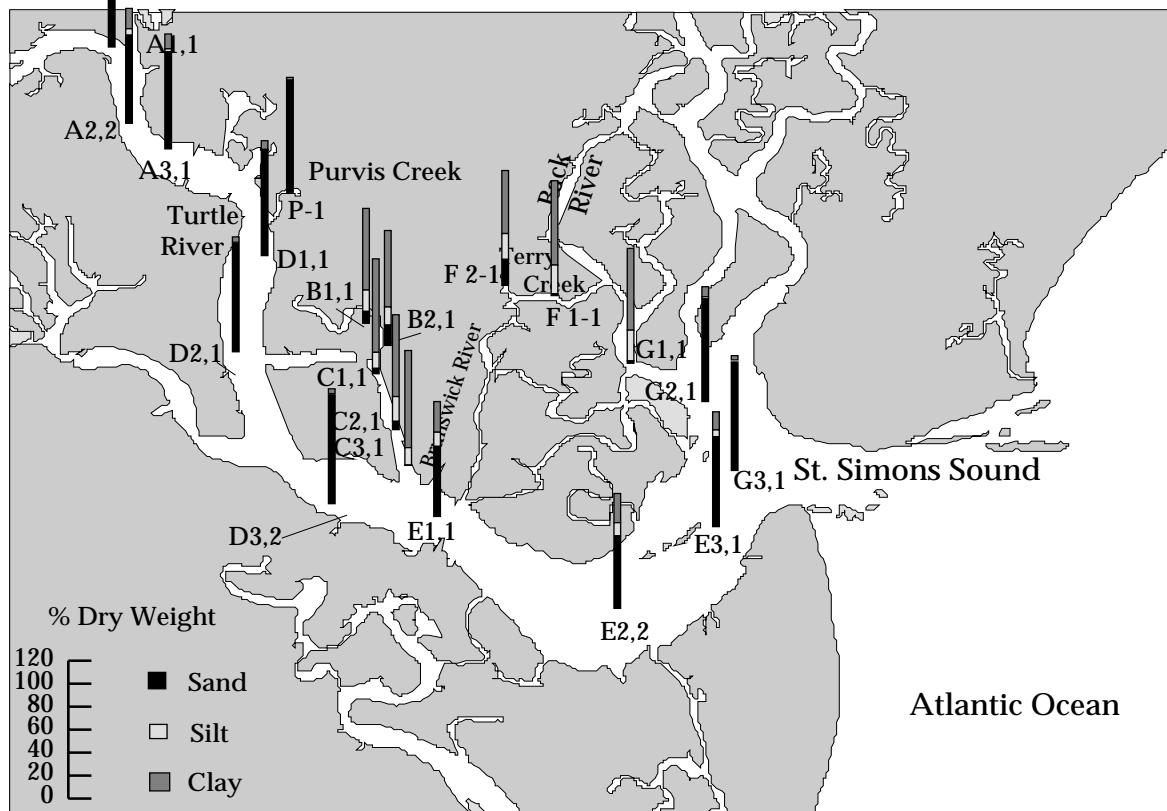


Figure 62. Concentrations of sand, silt, and clay in sediments from St. Simons Sound.

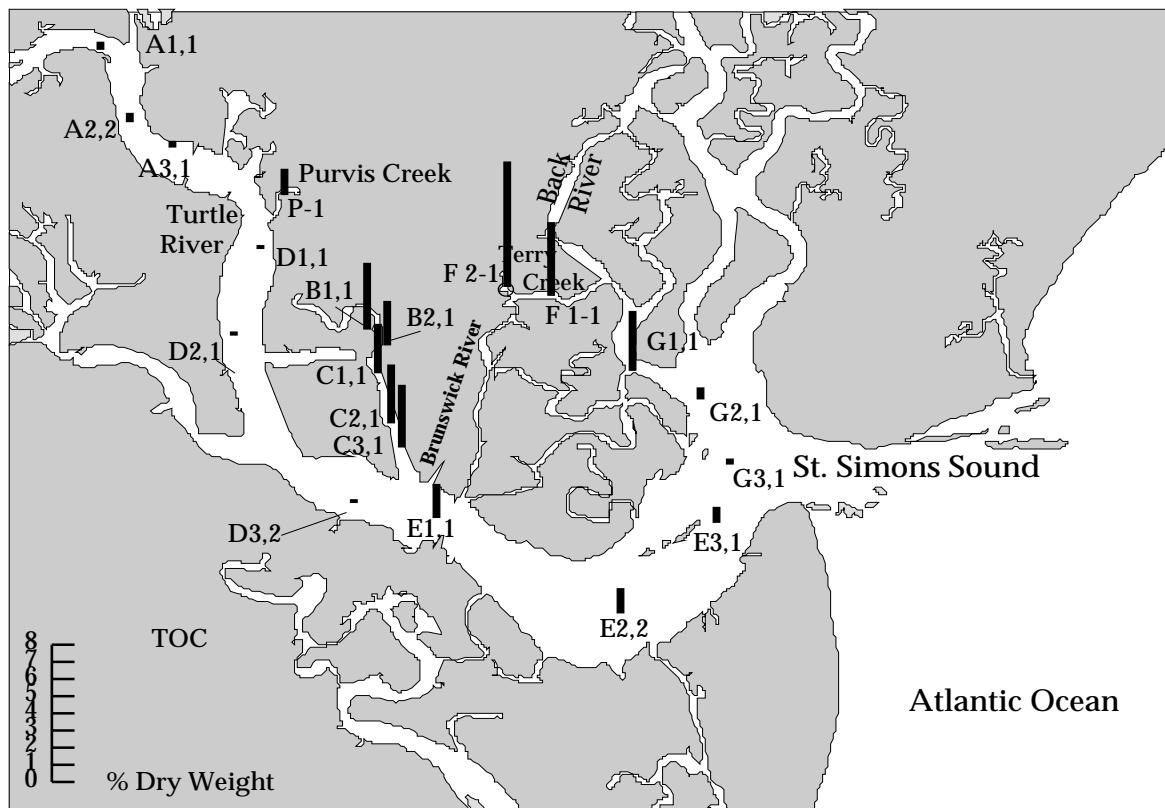


Figure 63. Concentrations of total organic carbon in sediments from St. Simons Sound.

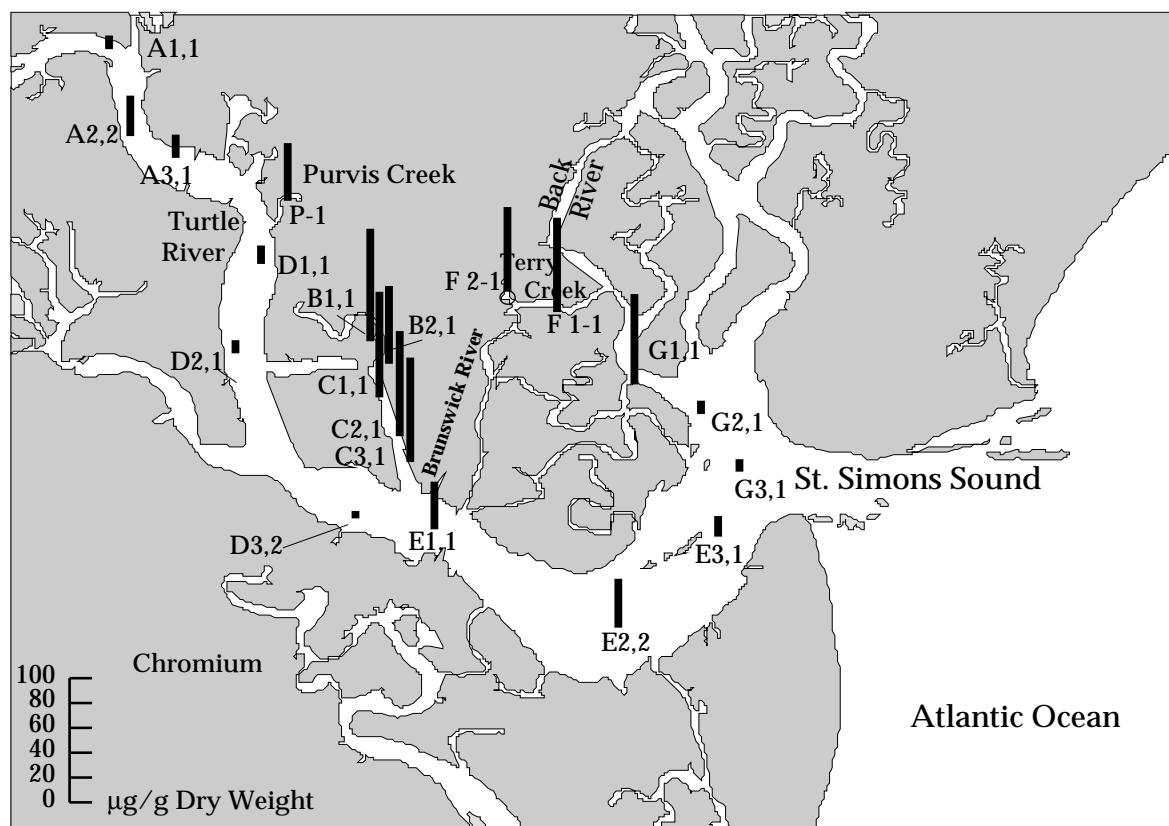


Figure 64. Concentrations of chromium in sediments from St. Simons Sound.

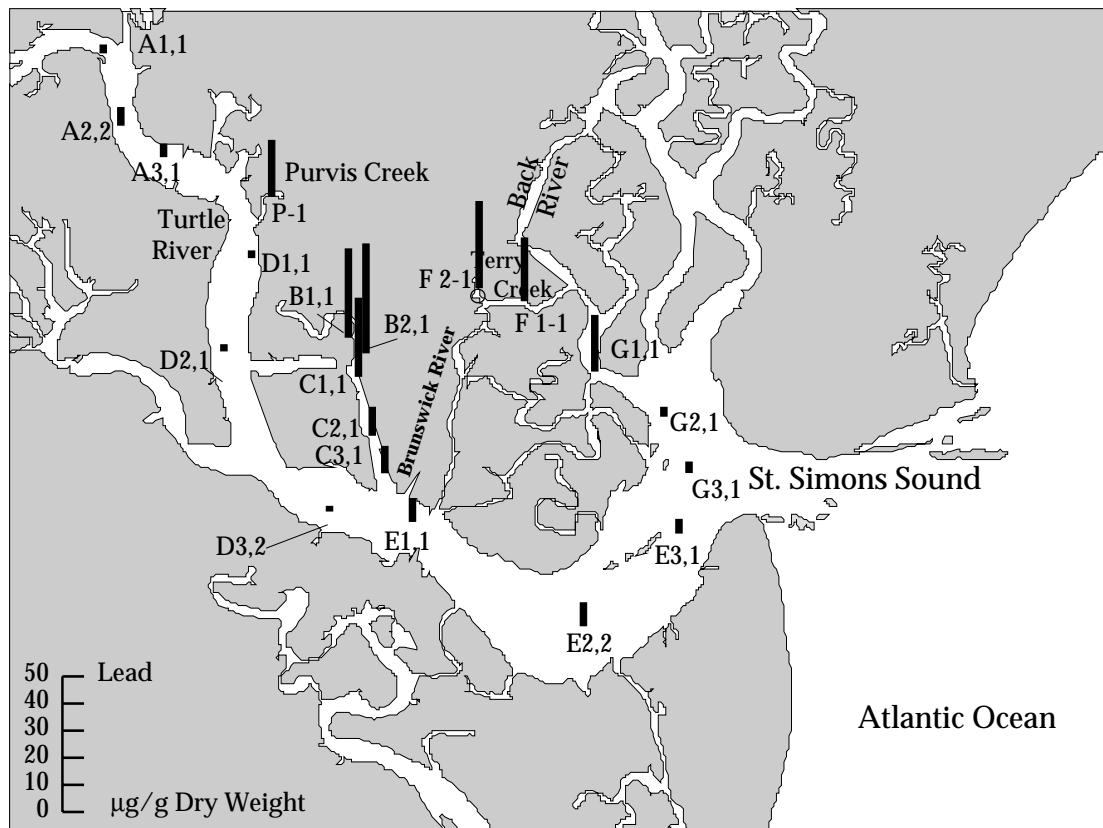


Figure 65. Concentrations of lead in sediments from St. Simons Sound.

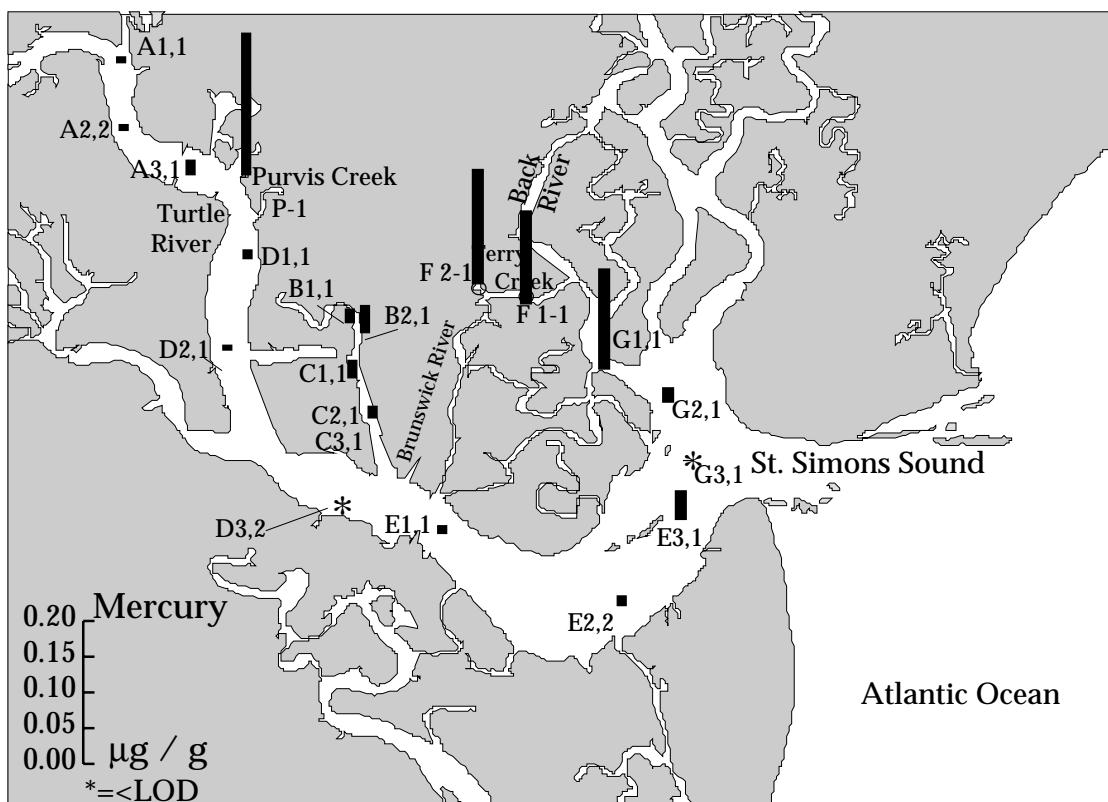


Figure 66. Concentrations of mercury in sediments from St. Simons Sound.

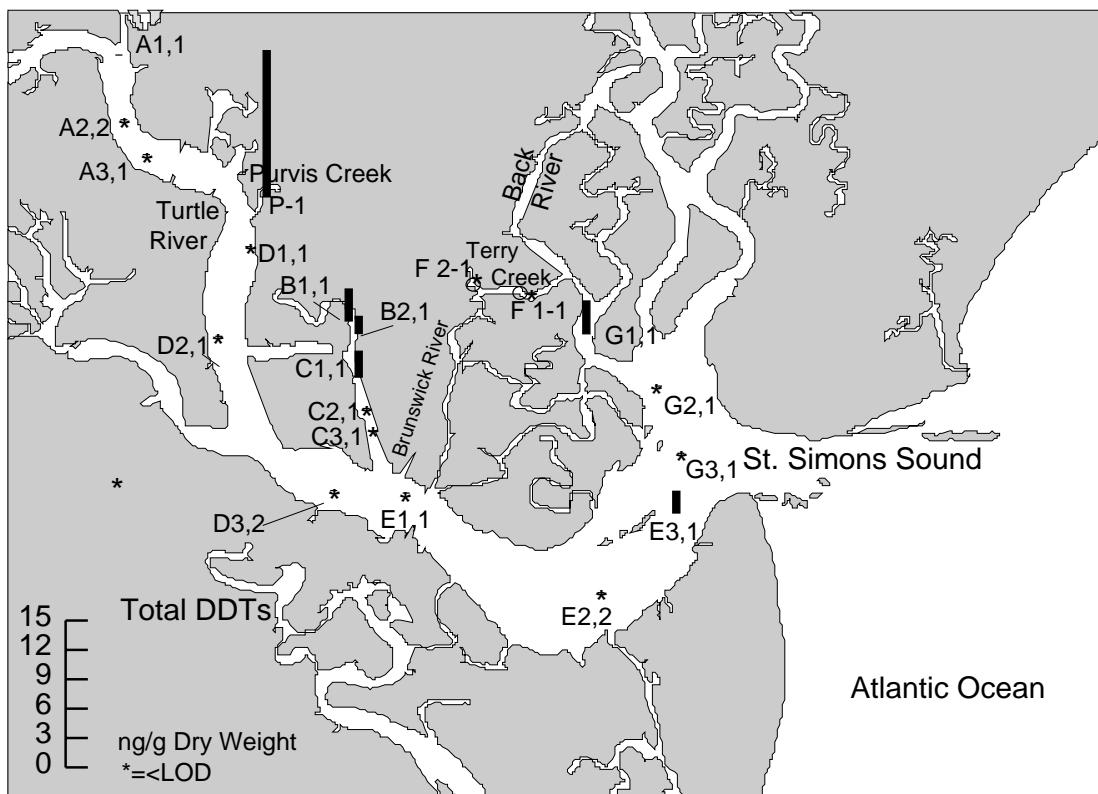


Figure 67. Concentrations of total DDTs in sediments from St. Simons Sound.

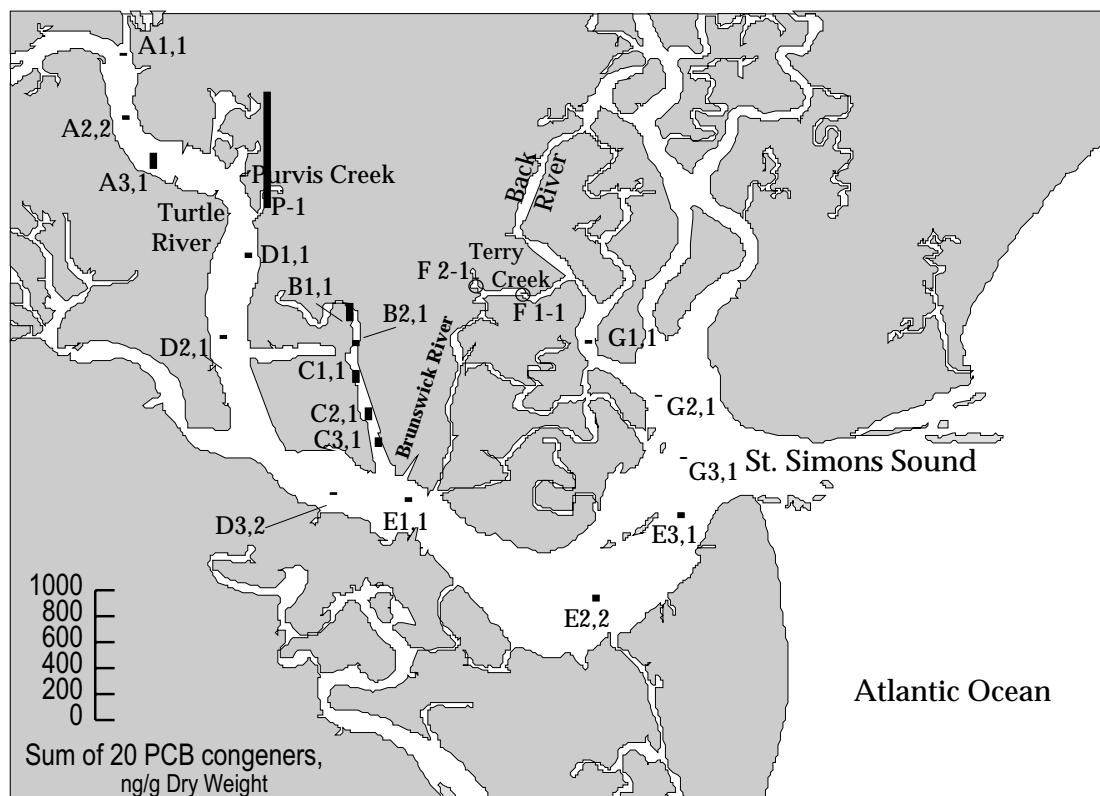


Figure 68. Concentrations of total PCBs (sum of 20 congeners) in sediments from St. Simons Sound.

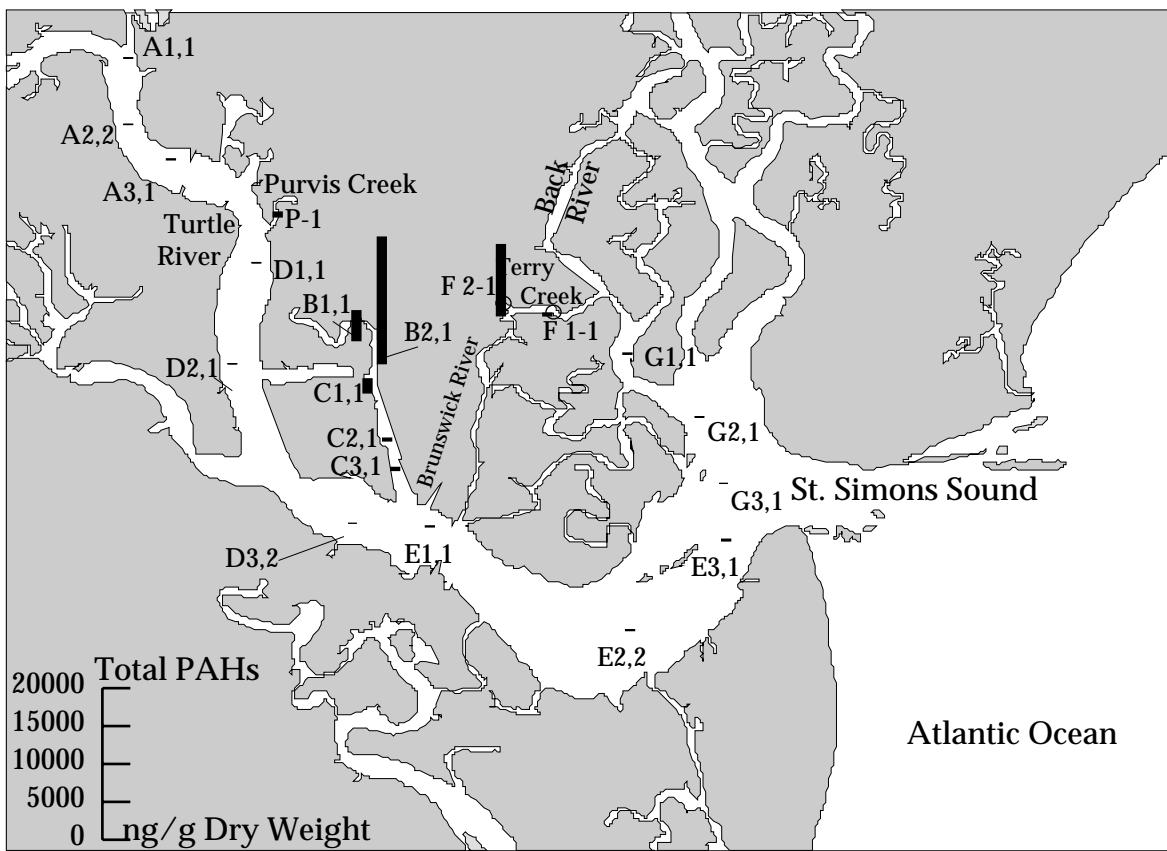


Figure 69. Concentrations of total PAHs in sediments from St. Simons Sound.



Figure 70. Distribution of Microtox test results in Charleston Harbor.

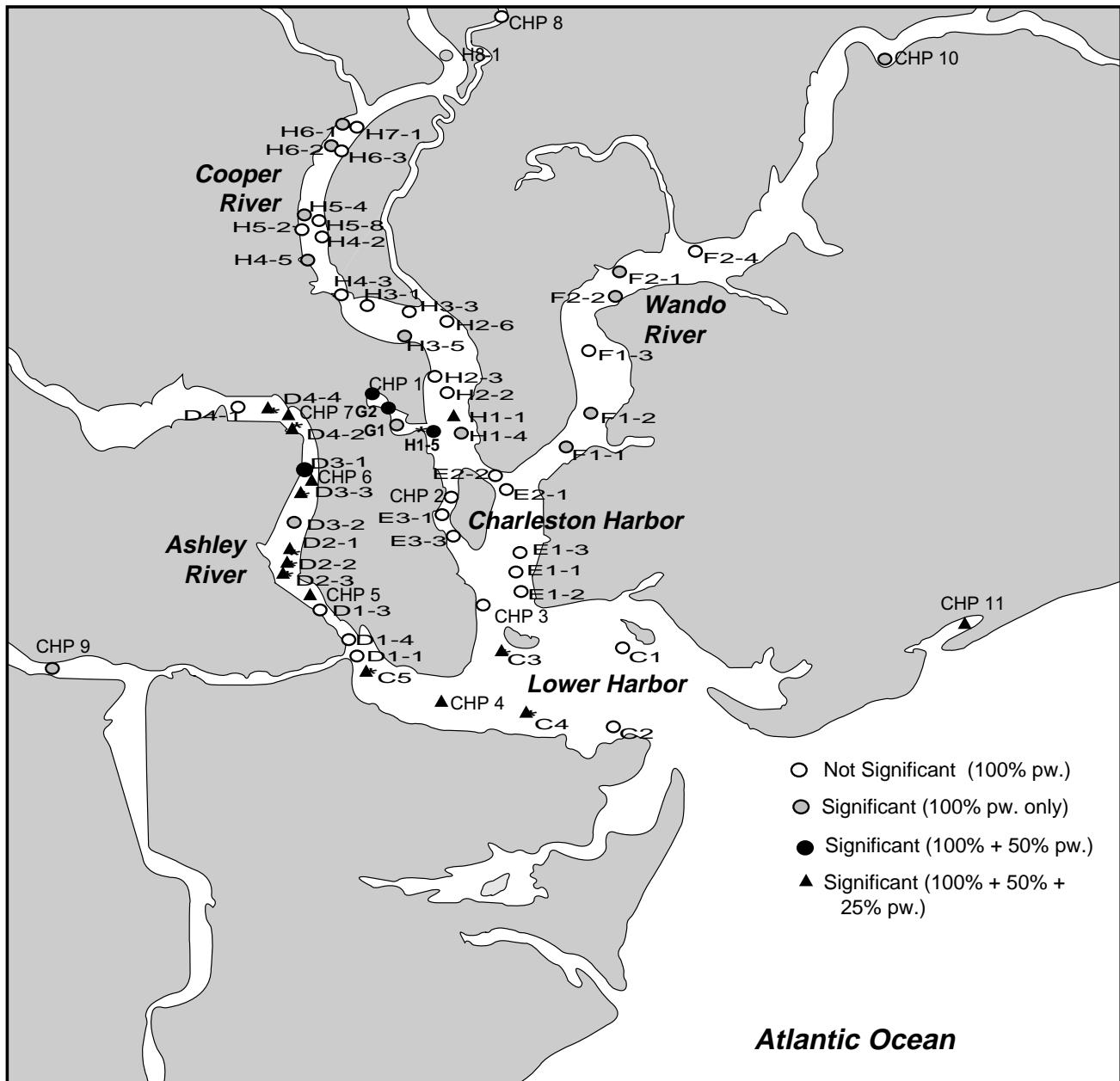


Figure 71. Distribution of results of urchin fertilization tests in 100%, 50%, and 25% porewater in Charleston Harbor.

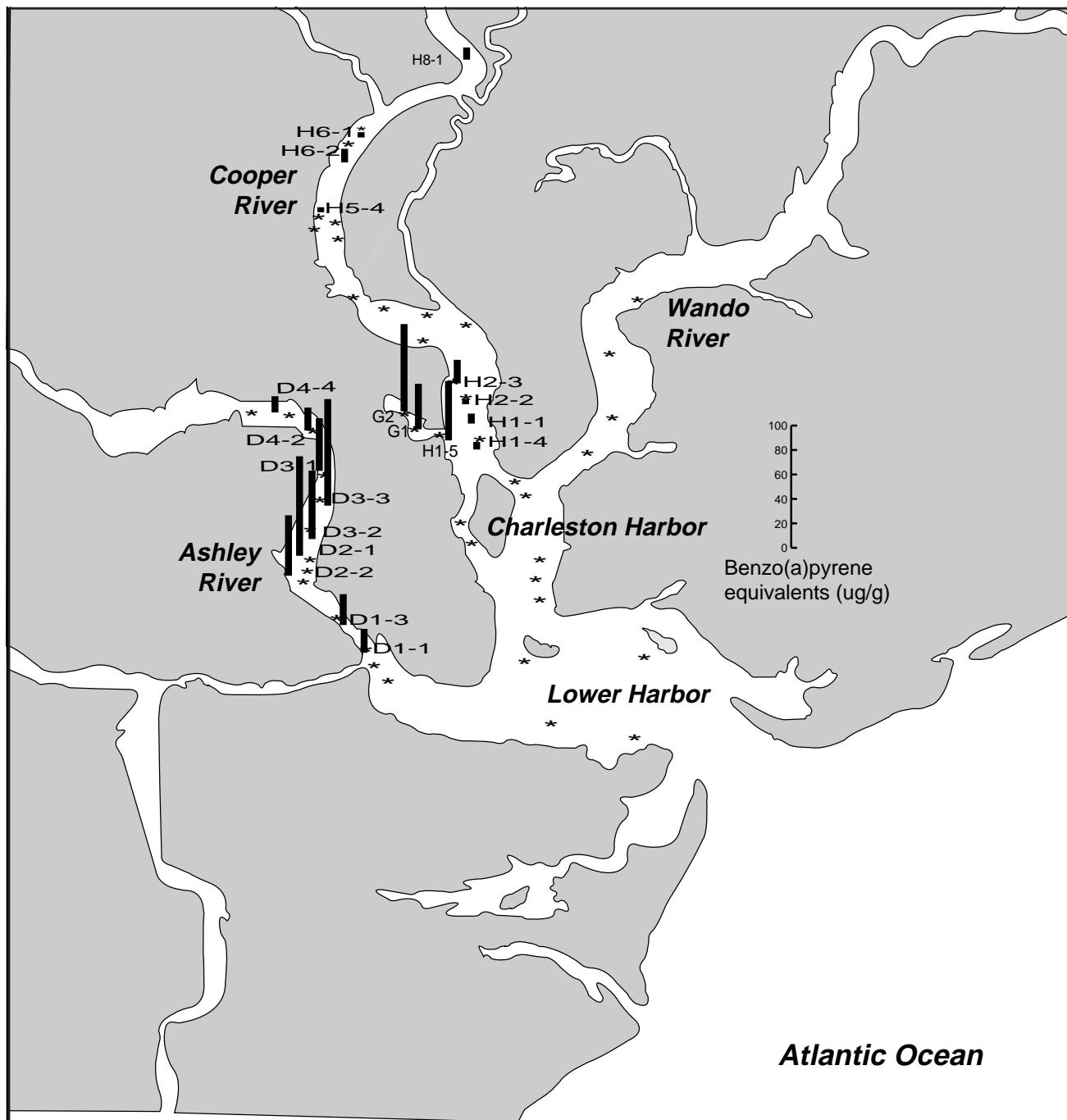


Figure 72. Results of cytochrome P-450 RGS assays of selected sediment samples from Charleston Harbor (ave. benzo(a)pyrene equivalents, ug/g).

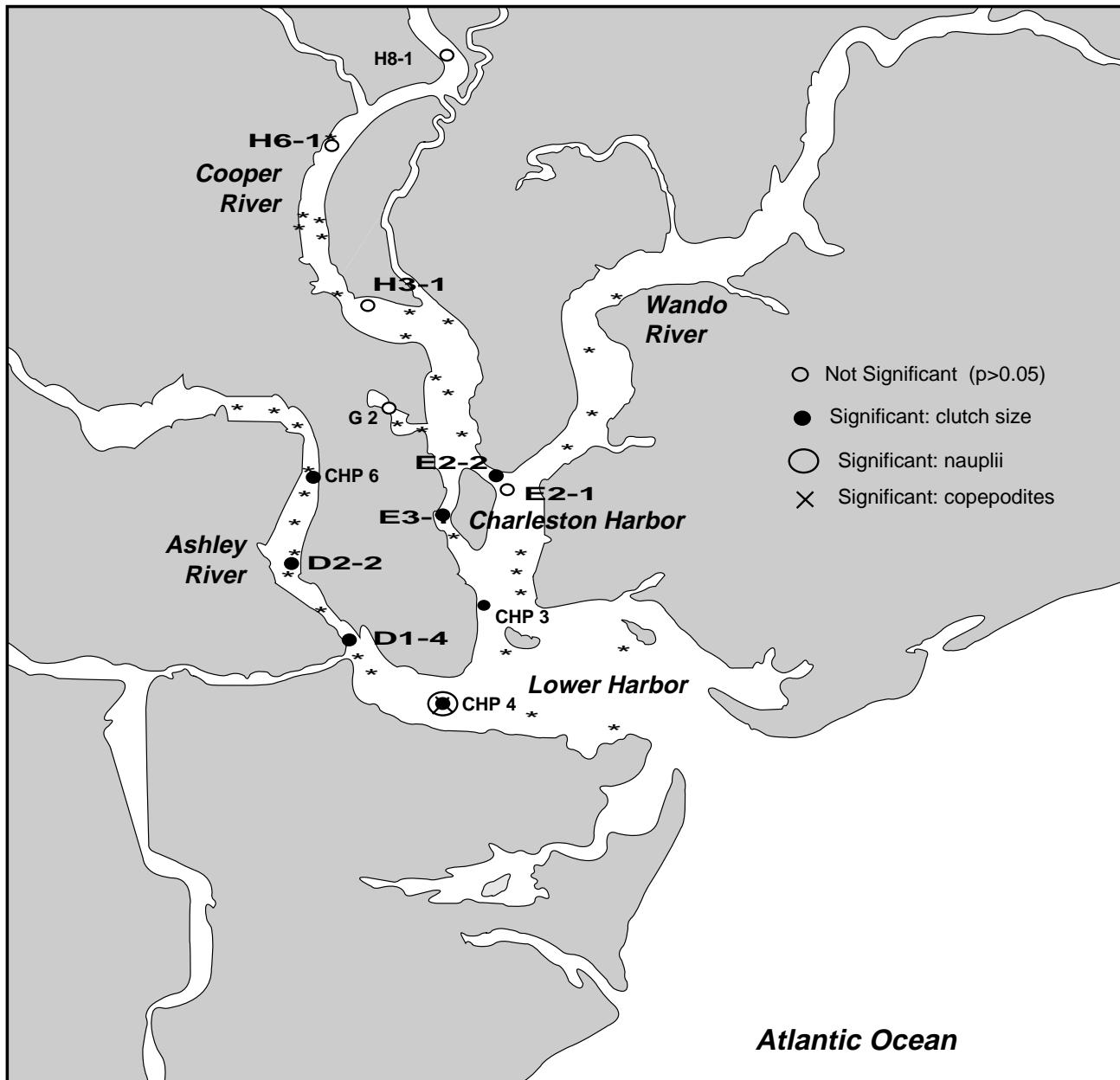


Figure 73. Distribution of copepod reproduction test results in Charleston Harbor.

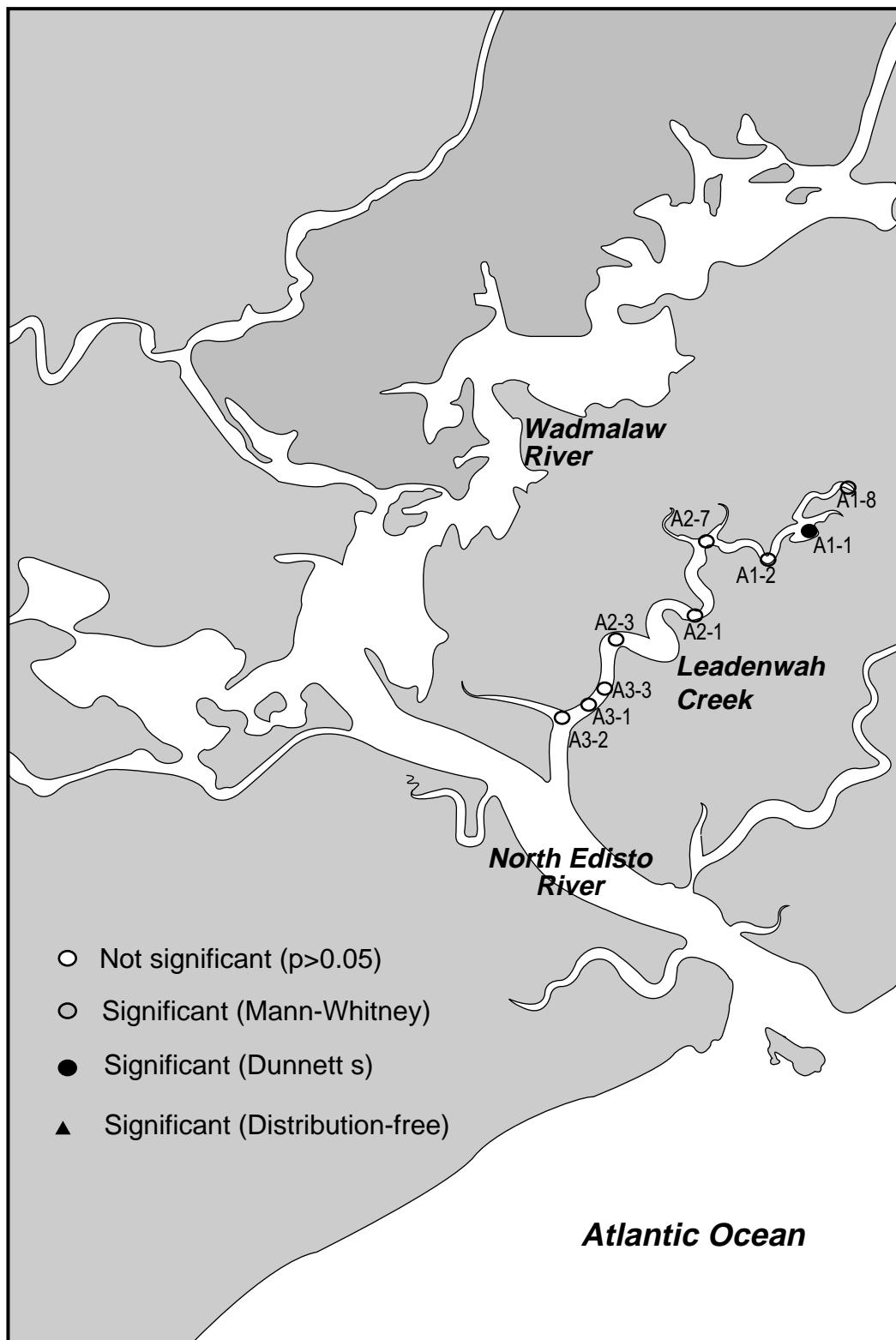


Figure 74. Distribution of Microtox test results in Leadenwah Creek.

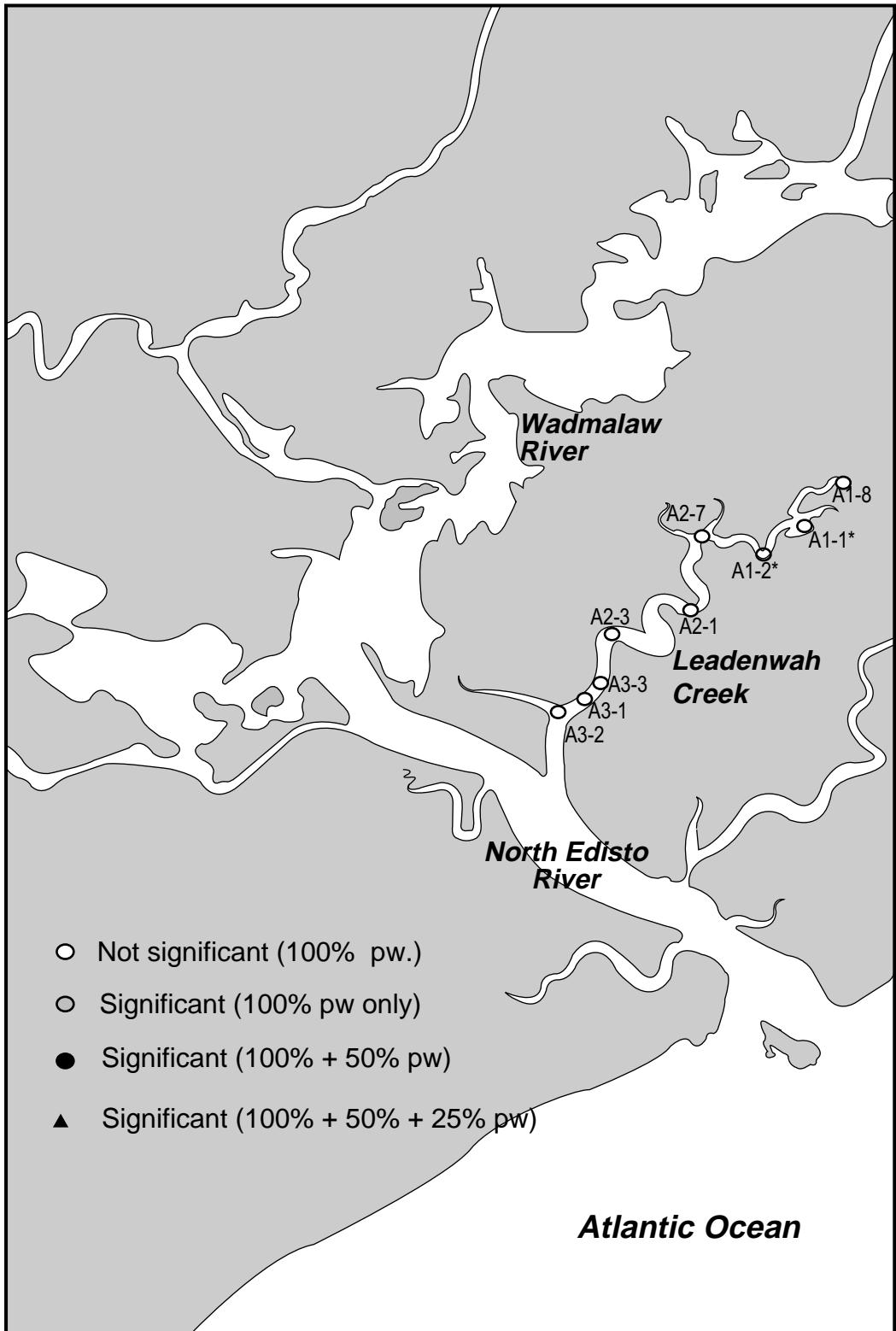


Figure 75. Distribution of urchin fertilization test results in Leadenwah Creek. (* significant results observed in 25% porewater only).

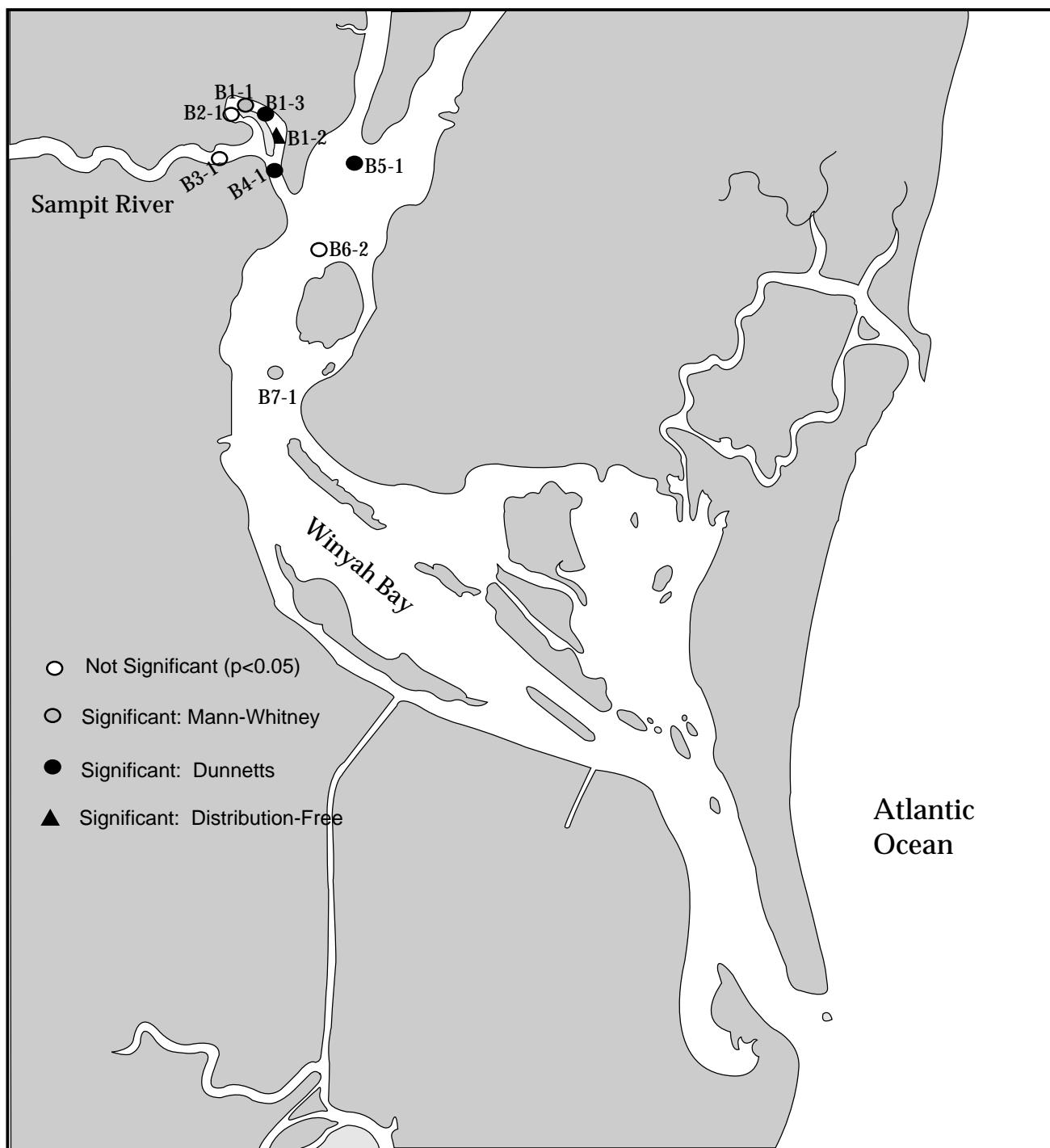


Figure 76. Distribution of Microtox test results for Winyah Bay.

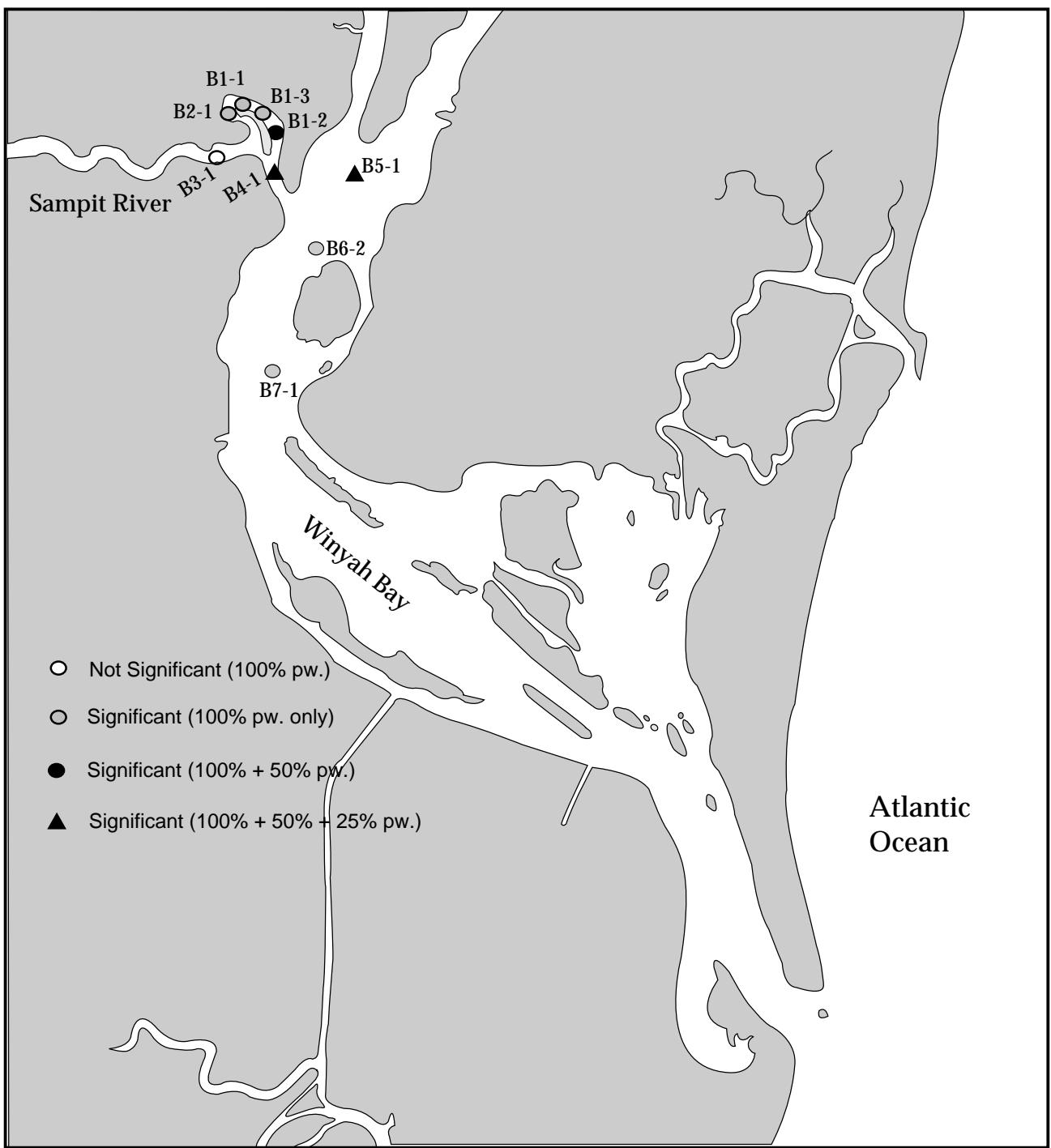


Figure 77. Distribution of results of urchin fertilization tests in 100%, 50%, and 25% porewater in Winyah Bay.

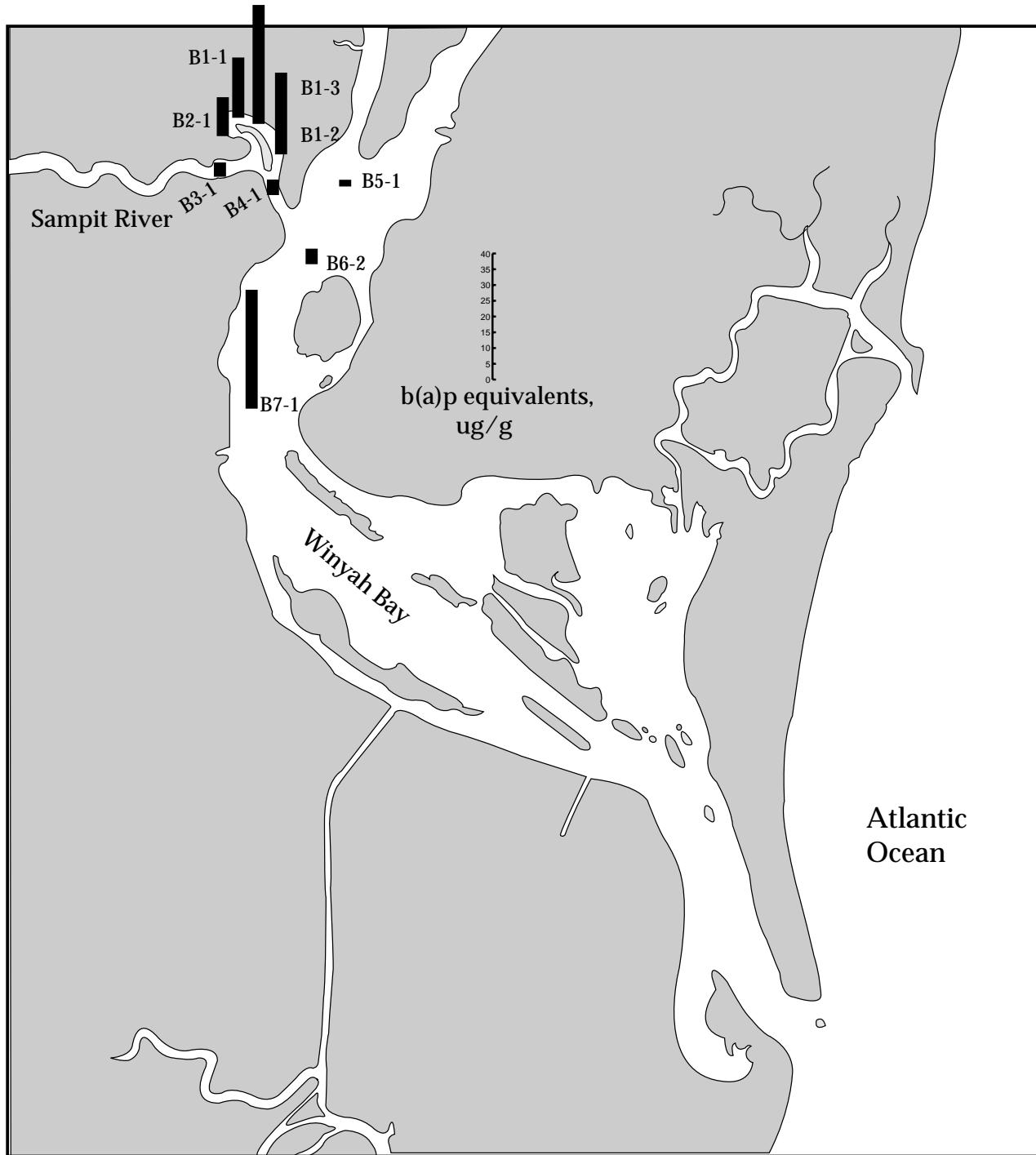


Figure 78. Results of cytochrome P-450 RGS assays of selected sediment samples from Winyah Bay (benzo(a)pyrene equivalents, ug/g).

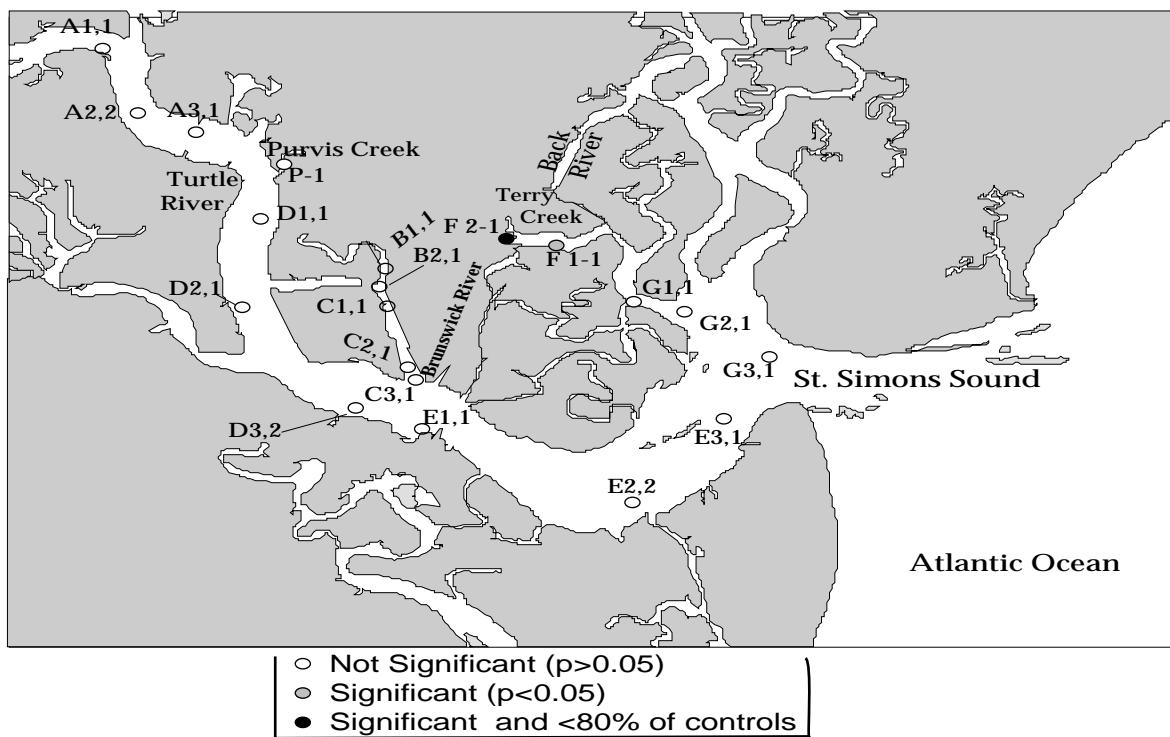


Figure 79. Distribution of amphipod test results in St. Simons Sound.

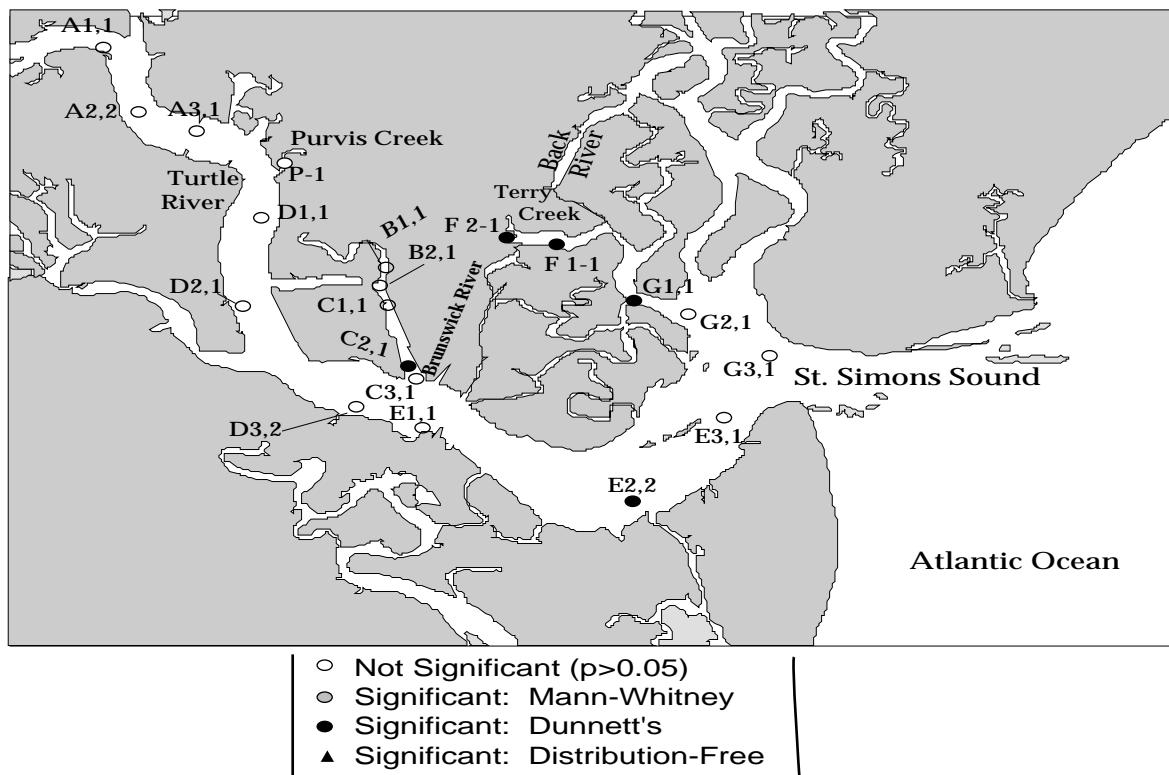


Figure 80. Distribution of Microtox test results in St. Simons Sound.

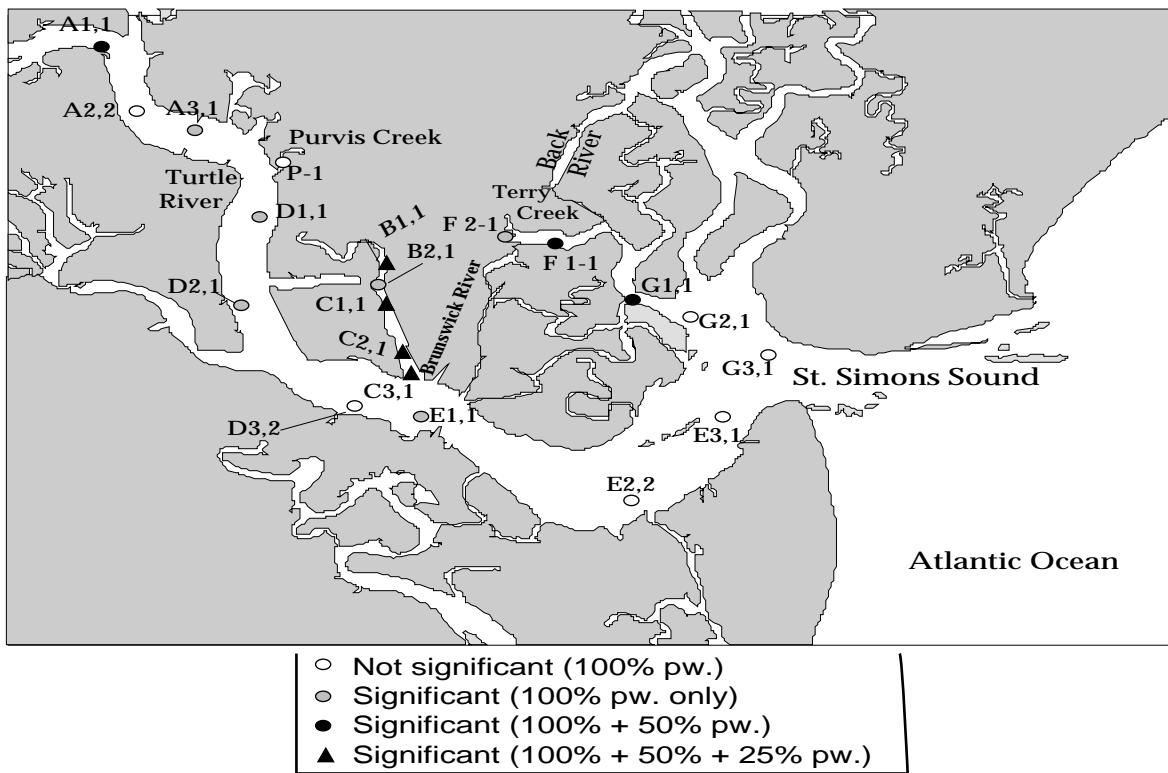


Figure 81. Distribution of urchin embryological development test results in St. Simons Sound.

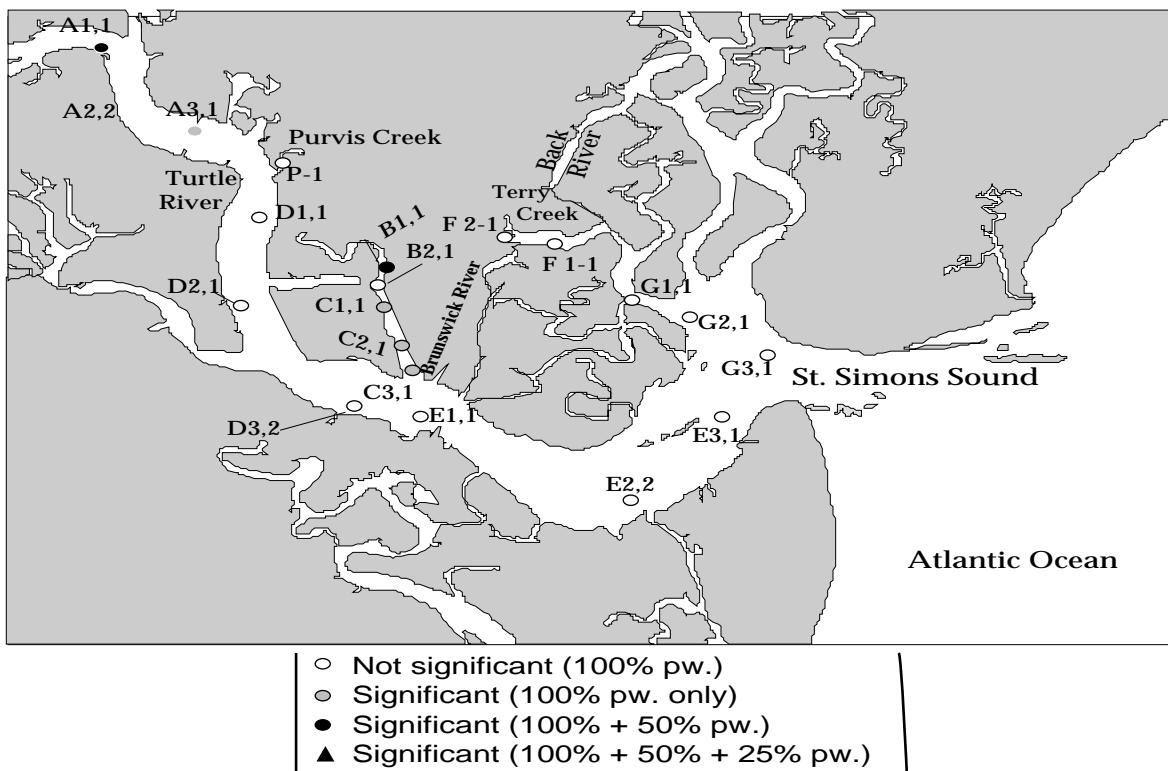


Figure 82. Distribution of urchin egg fertilization test results in St. Simons Sound.

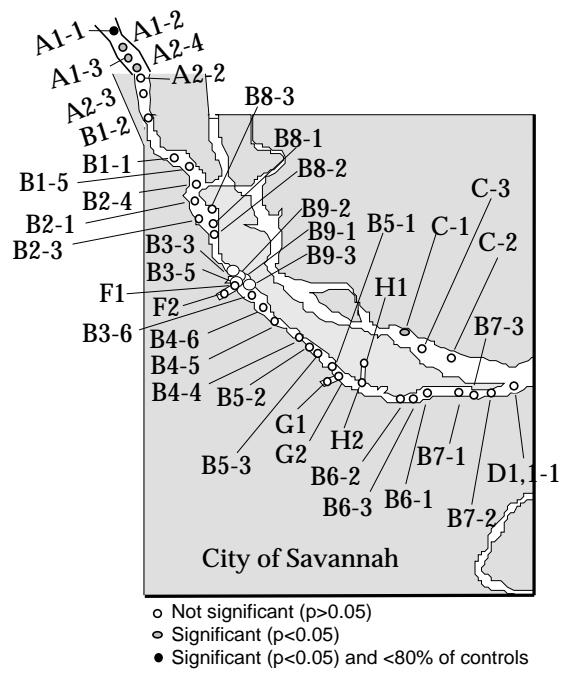


Figure 83. Distribution of Amphipod results in the upper Savannah River.

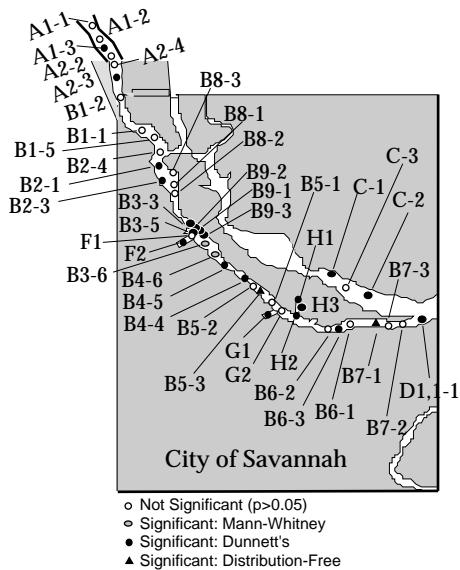


Figure 84. Distribution of Microtox test results in the upper Savannah River.

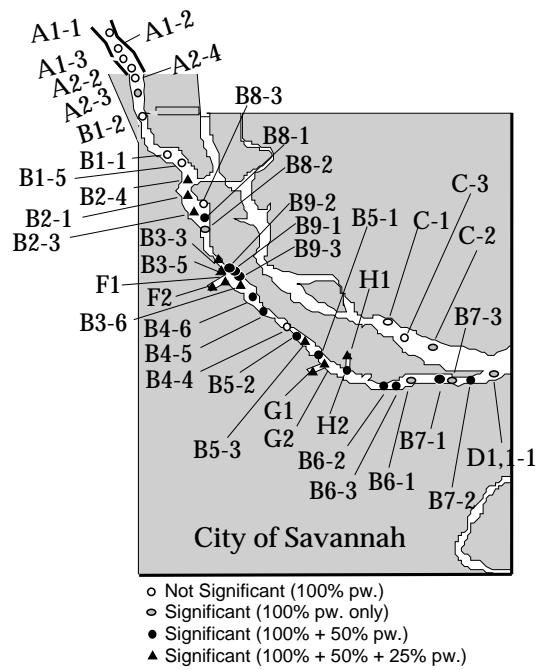


Figure 85. Distribution of urchin embryological development test results in the upper Savannah River.

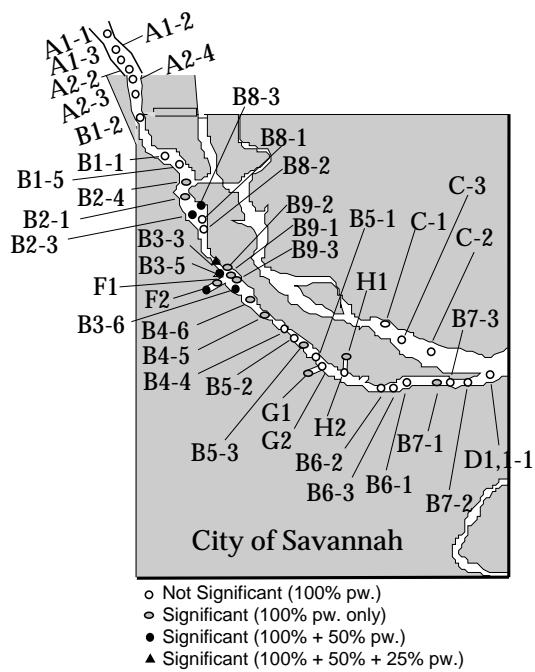


Figure 86. Distribution of urchin egg fertilization test results in the upper Savannah River.

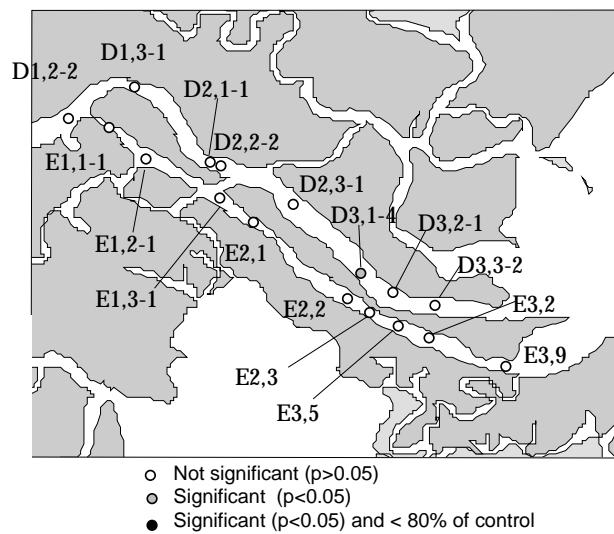


Figure 87. Distribution of amphipod test results in the lower Savannah River.

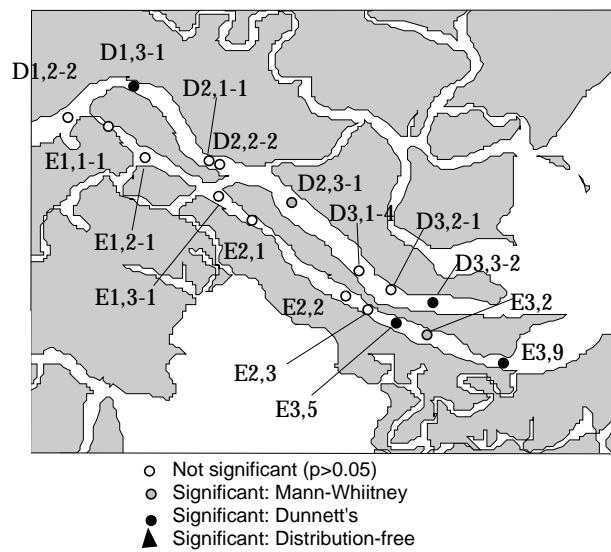


Figure 88. Distribution of Microtox test results in the lower Savannah River.

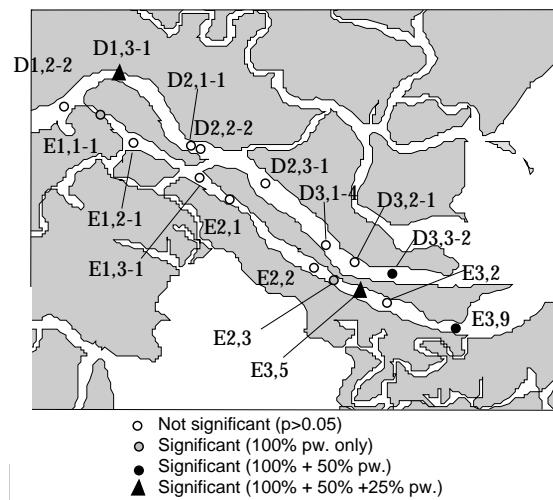


Figure 89. Distribution of urchin embryological development test results in the lower Savannah River.

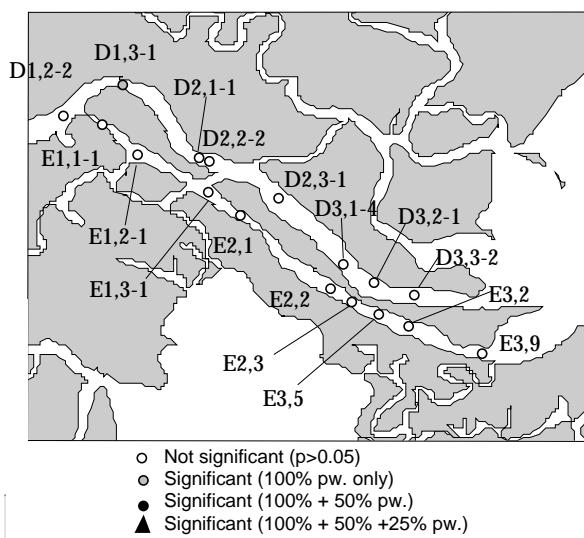


Figure 90. Distribution of urchin egg fertilization test results in the lower Savannah River.

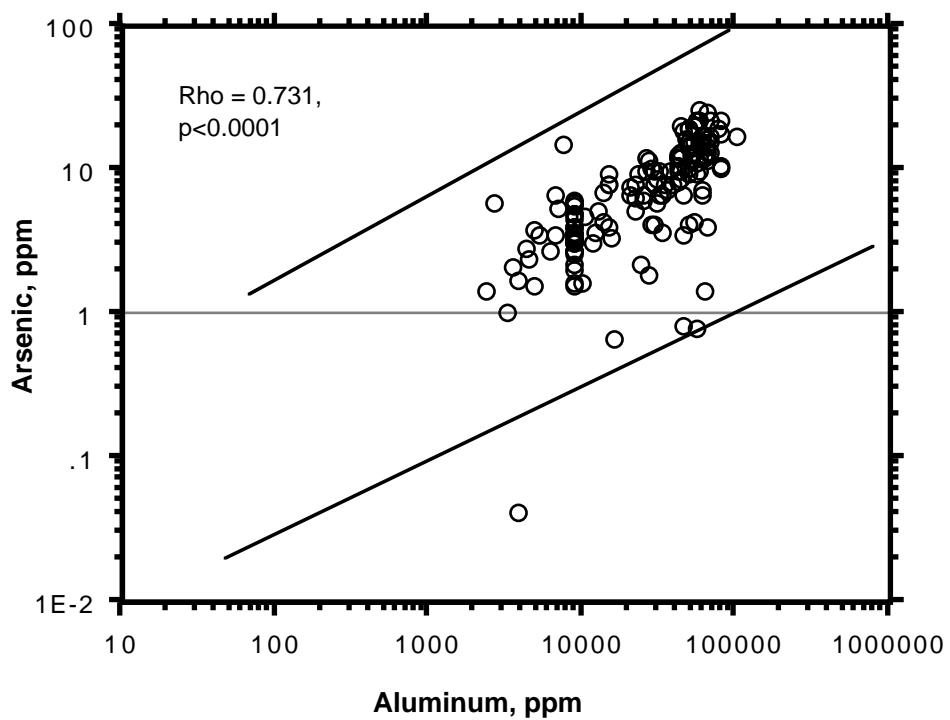


Figure 91. Relationship between the concentrations of aluminum and arsenic in sediment samples from South Carolina/Georgia.

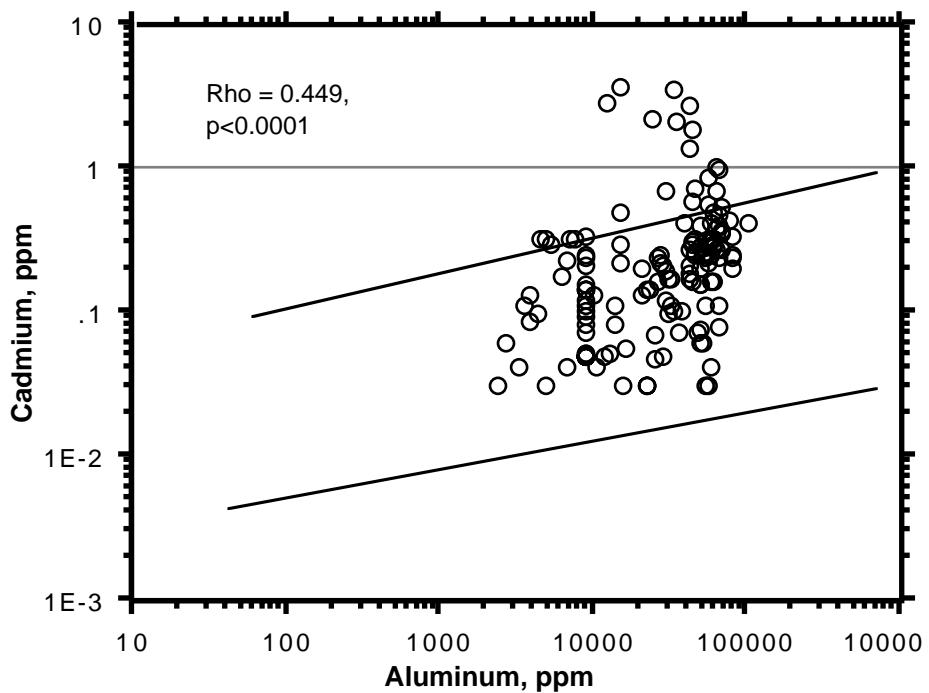


Figure 92. Relationship between the concentrations of aluminum and cadmium in sediment samples from South Carolina/Georgia.

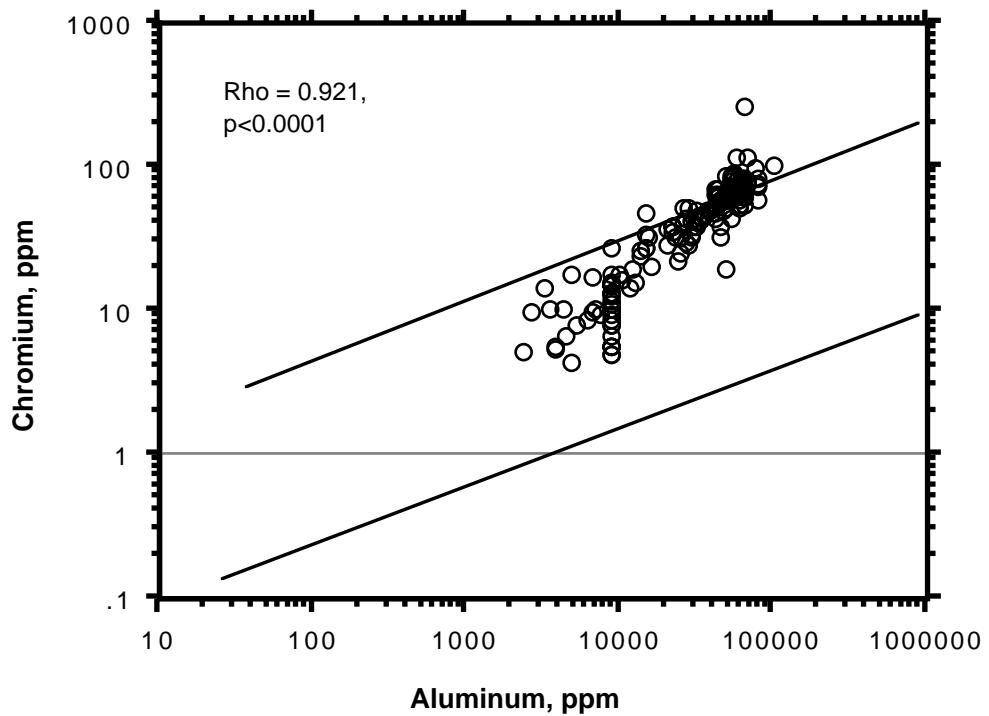


Figure 93. Relationship between the concentrations of aluminum and chromium in sediment samples from South Carolina/Georgia.

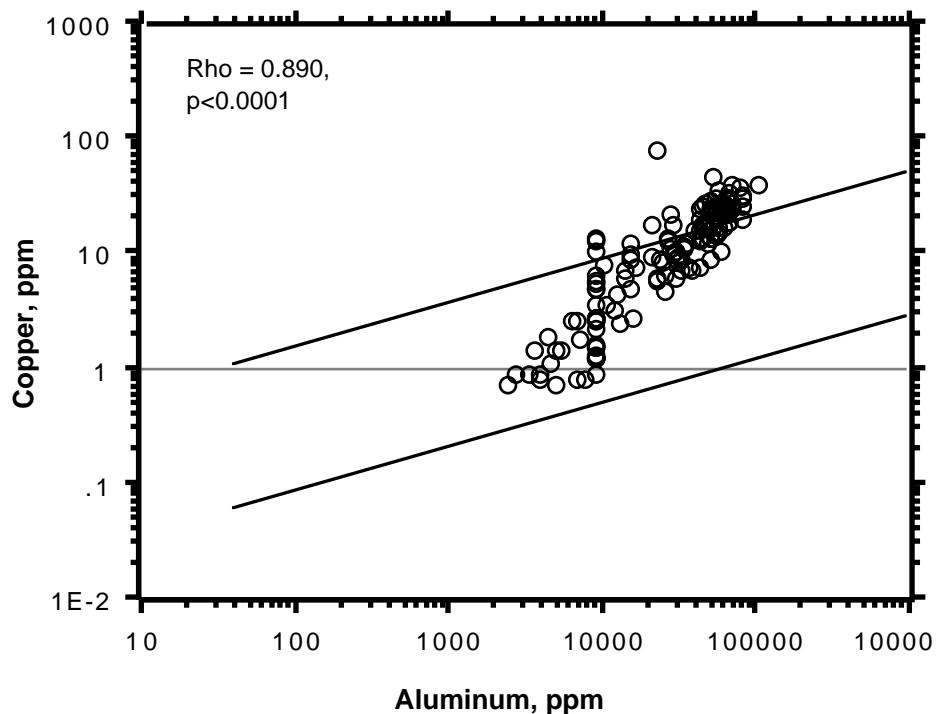


Figure 94. Relationship between the concentrations of aluminum and copper in sediment samples from South Carolina/Georgia.

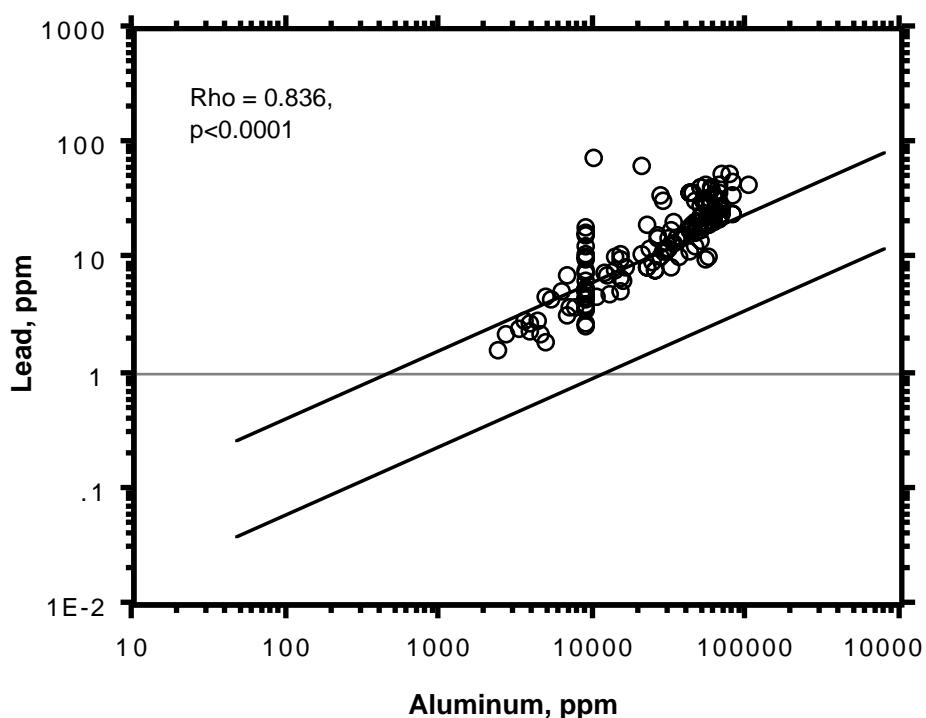


Figure 95. Relationship between the concentrations of aluminum and lead in sediment samples from South Carolina/Georgia.

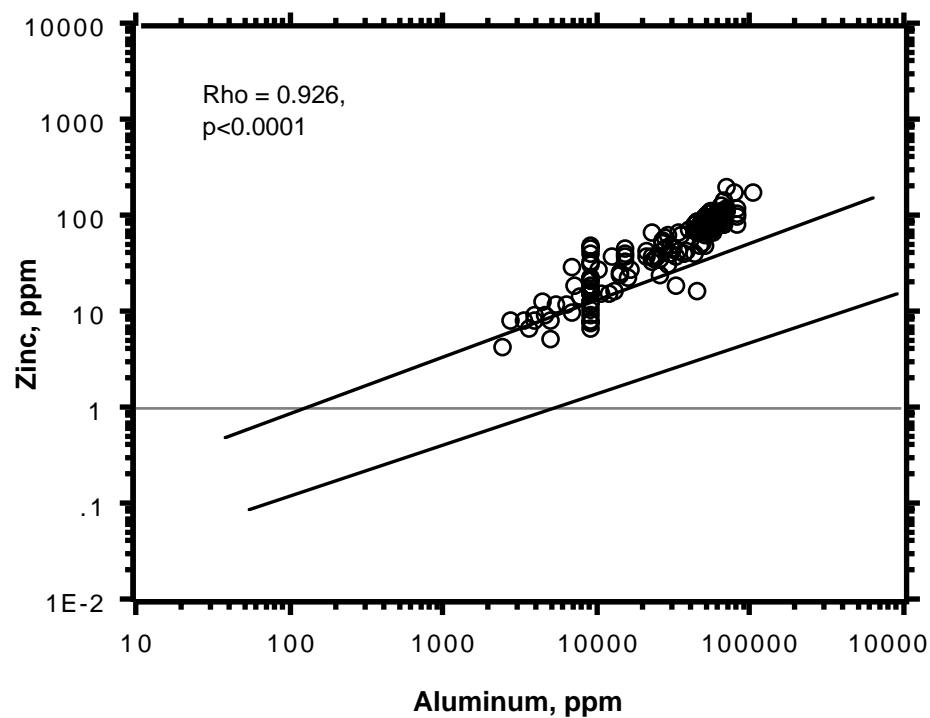


Figure 96. Relationship between the concentrations of aluminum and zinc in sediment samples from South Carolina/Georgia.

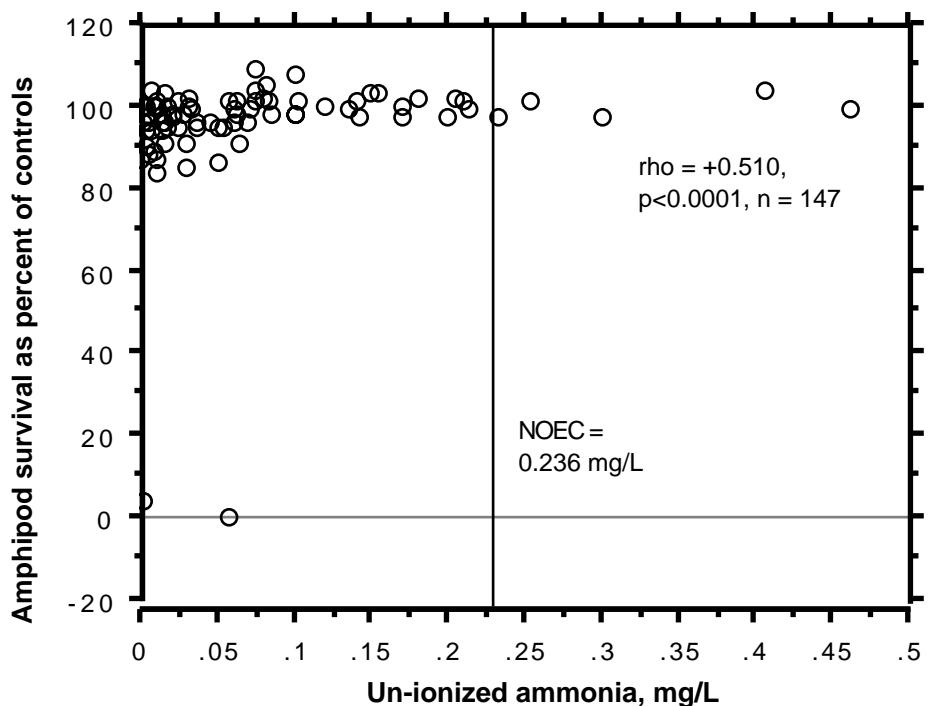


Figure 97. Relationship between the concentrations of un-ionized ammonia in overlying water of test chambers and amphipod survival.

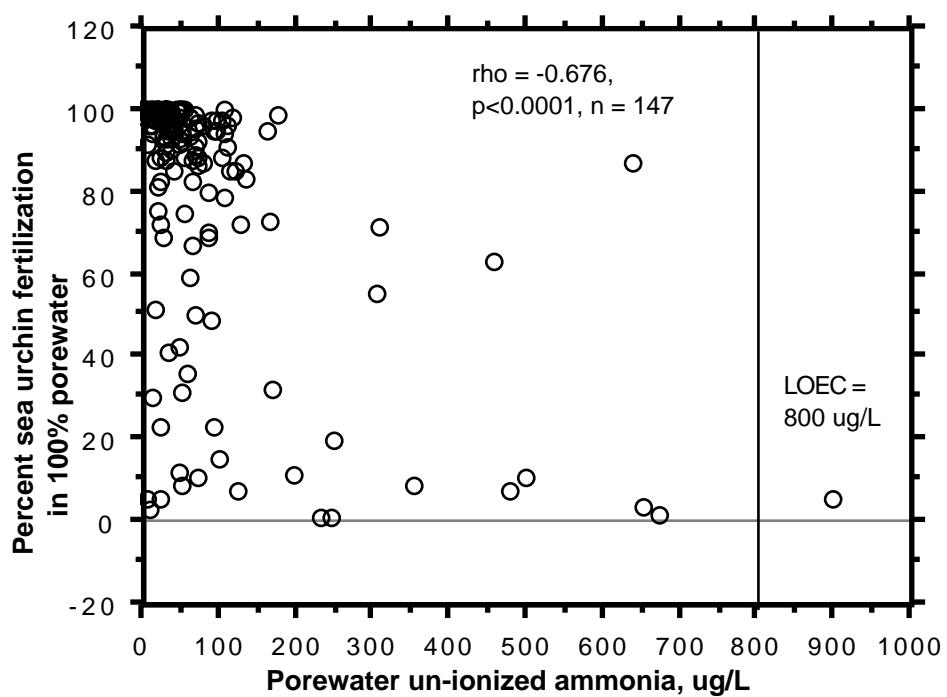


Figure 98. Relationship between un-ionized ammonia in porewater and percent sea urchin fertilization in 100% porewater.

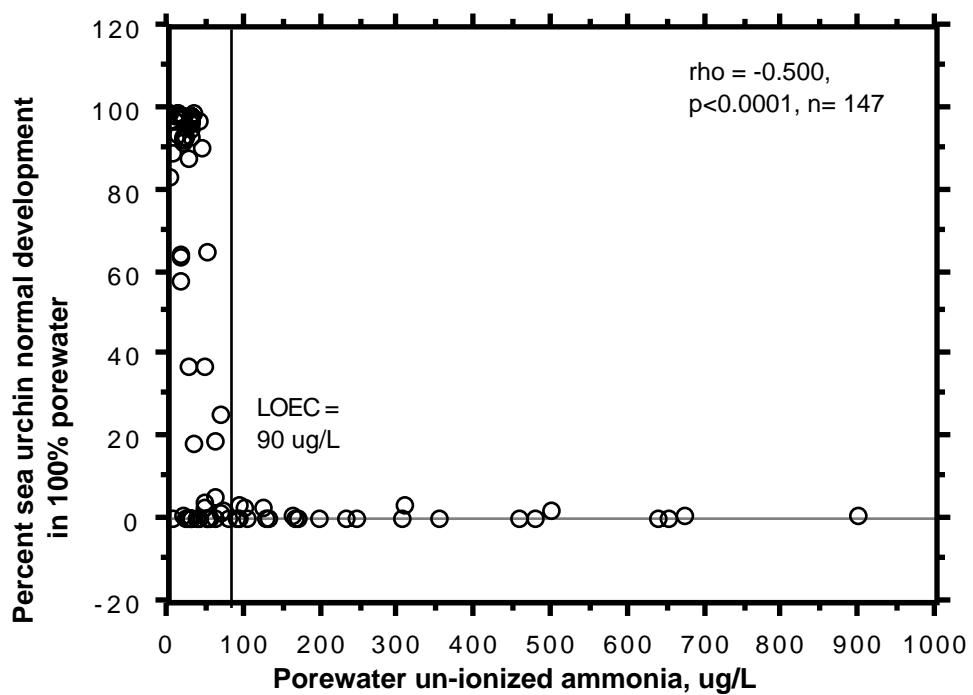


Figure 99. Relationship between the concentrations of un-ionized ammonia in porewater and percent sea urchin normal development in 100% porewater.

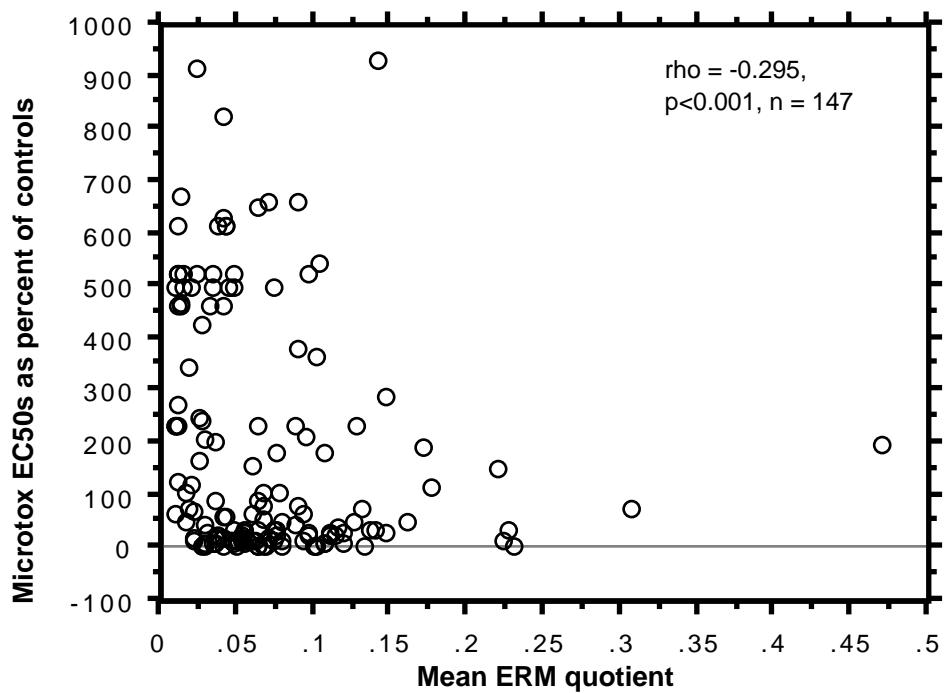


Figure 100. Relationship between mean ERM quotients for 25 substances and microbial bioluminescence.

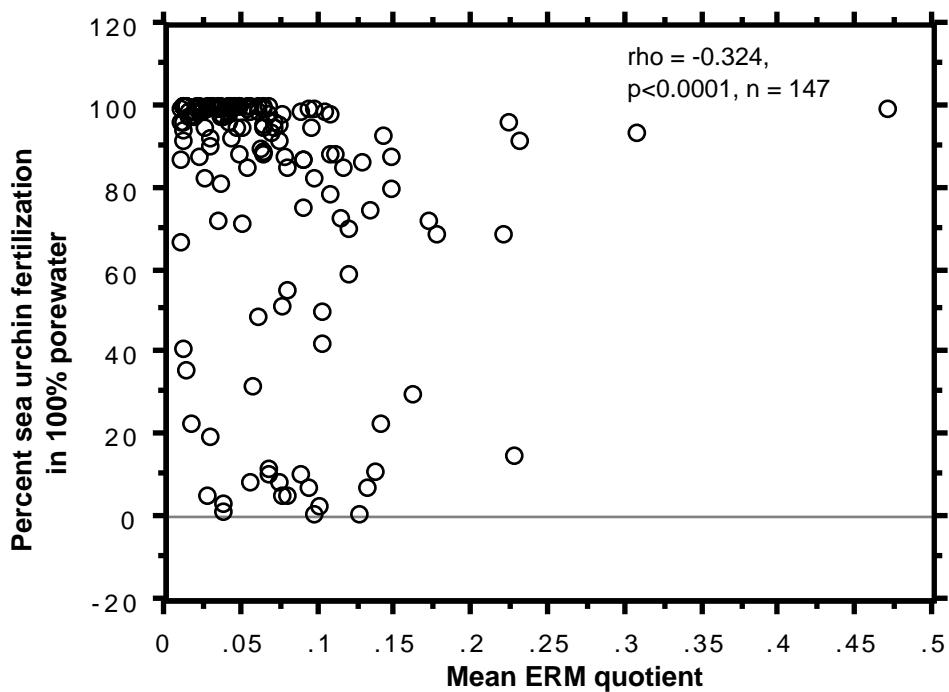


Figure 101. Relationship between mean ERM quotients for 25 substances and percent sea urchin fertilization in 100% porewater.

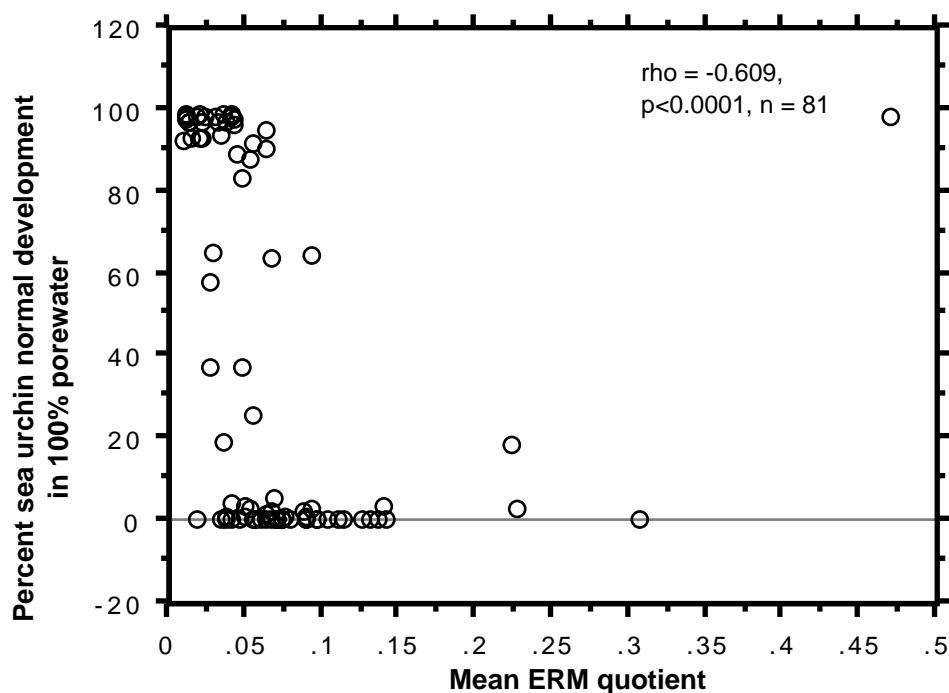


Figure 102. Relationship between mean ERM quotients for 25 substances and percent sea urchin normal development.

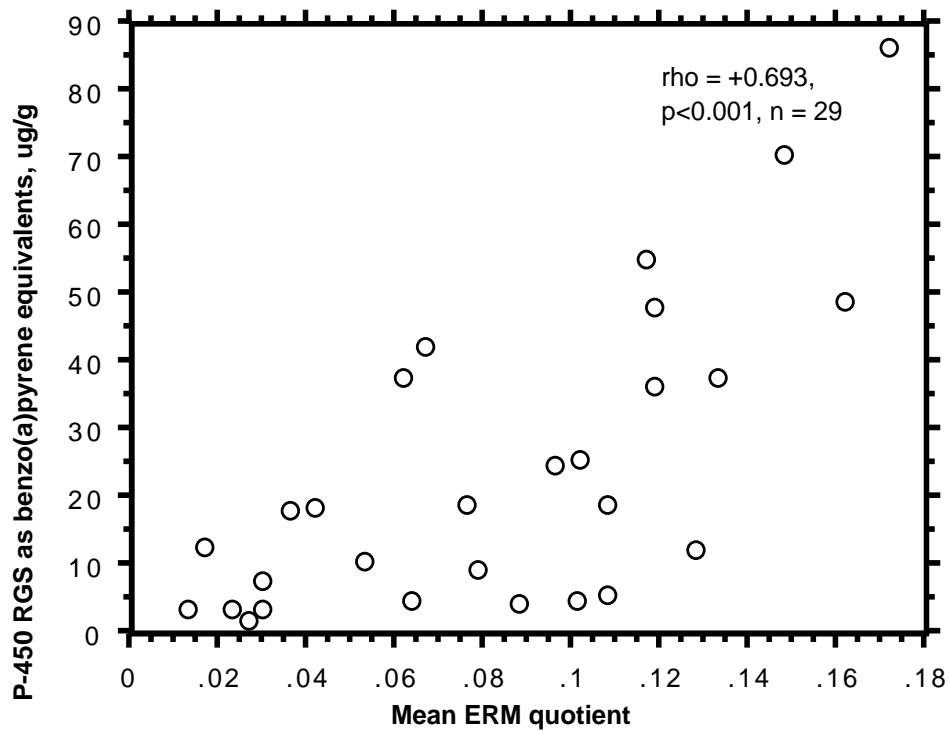


Figure 103. Relationship between mean ERM quotients and cytochrome P450 RGS assay results as benzo(a)pyrene equivalents.

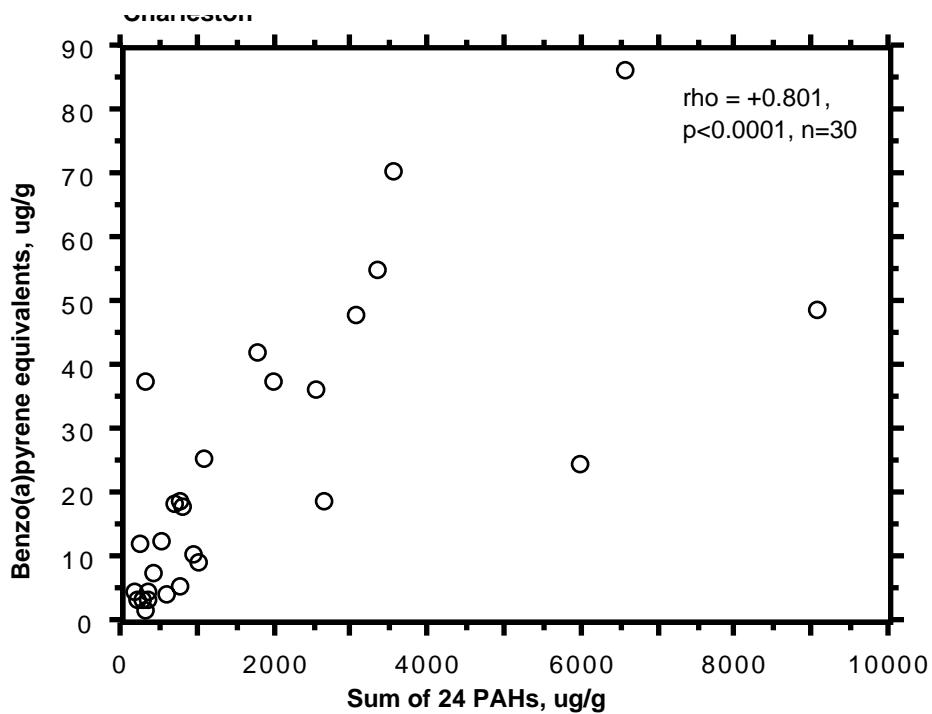


Figure 104. Relationship between the concentrations of total PAHs and cytochrome P-450 RGS assay results as benzo(a)pyrene equivalents.

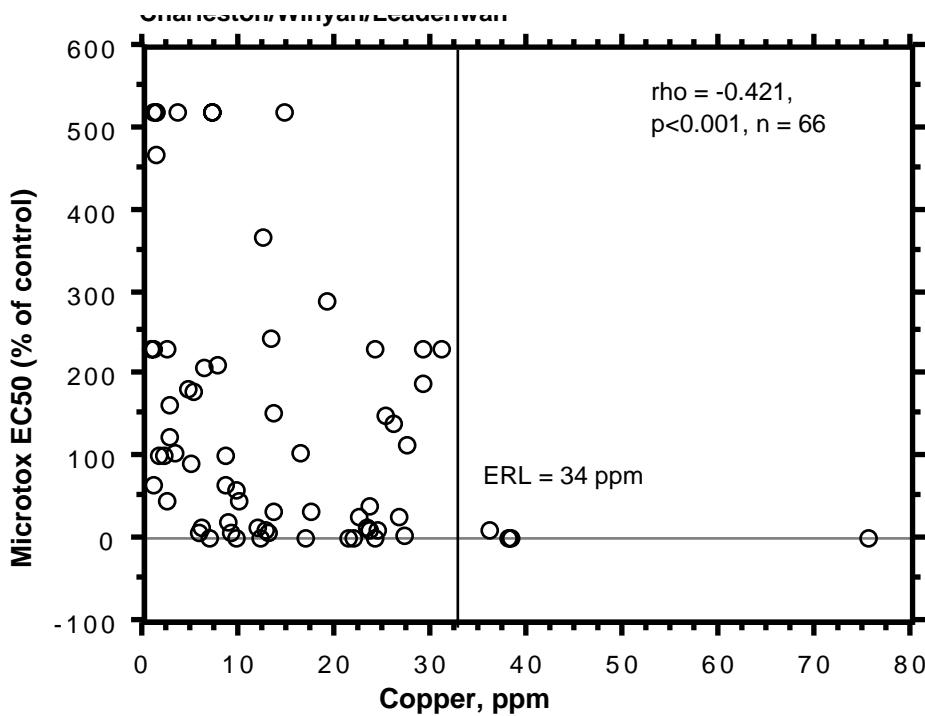


Figure 105. Relationship between microbial bioluminescence and the concentrations of copper in sediments from Charleston Harbor, Winyah Bay, and Leadenwah Creek.

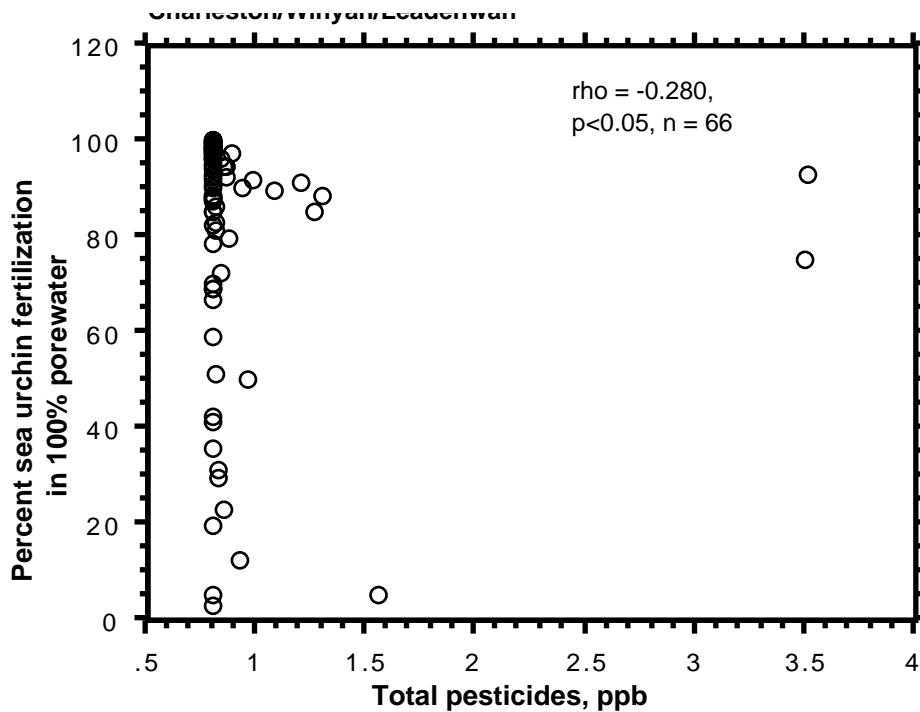


Figure 106. Relationship between sea urchin fertilization and the concentrations of total pesticides in sediments from Charleston Harbor, Winyah Bay, and Leadenwah Creek.

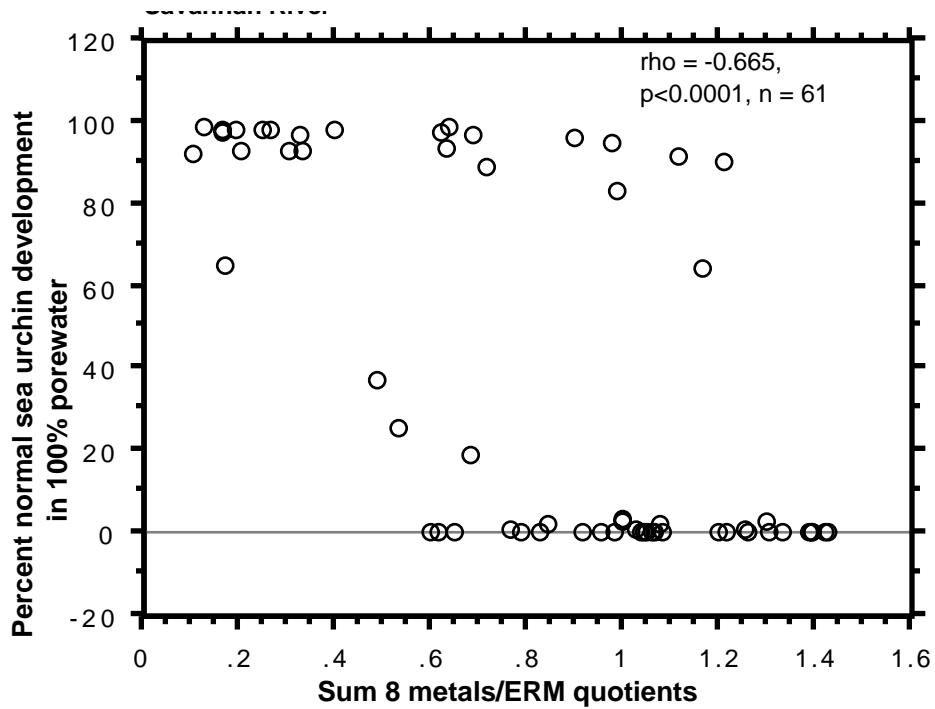


Figure 107. Relationship between the sum of 8 metals/ERM quotients and percent normal sea urchin development in sediments from Savannah River.

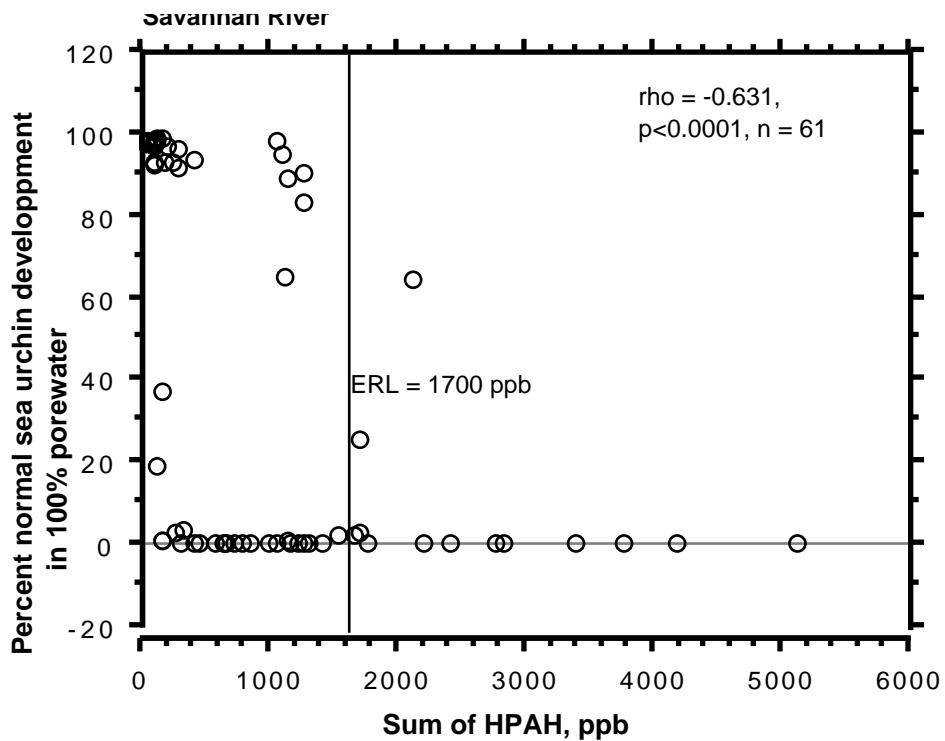


Figure 108. Relationship between percent normal sea urchin development and the sum of total HPAHs in sediments from the Savannah River.

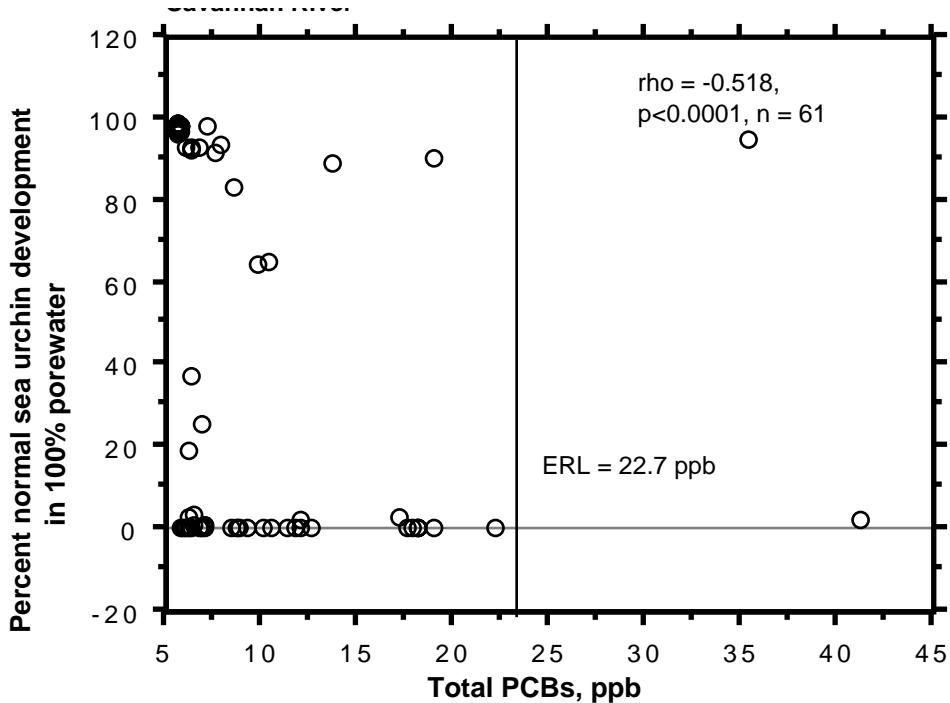


Figure 109. Relationship between percent normal sea urchin development and concentrations of total PCBs in sediments from the Savannah River.

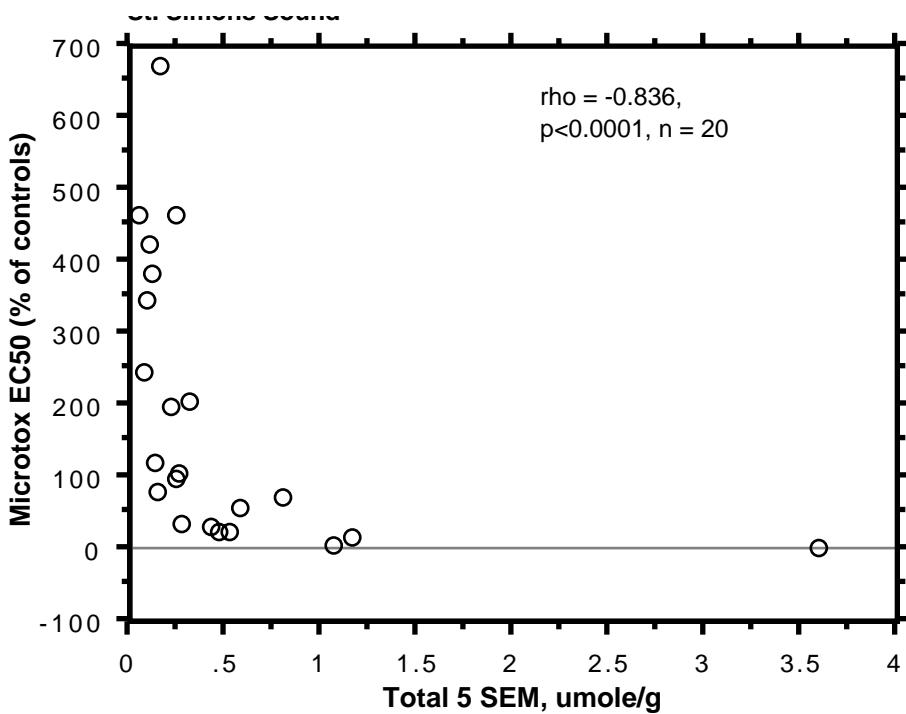


Figure 110. Relationship between microbial bioluminescence and the sums of five simultaneously-extracted metals (SEM) in sediments from St. Simons Sound.

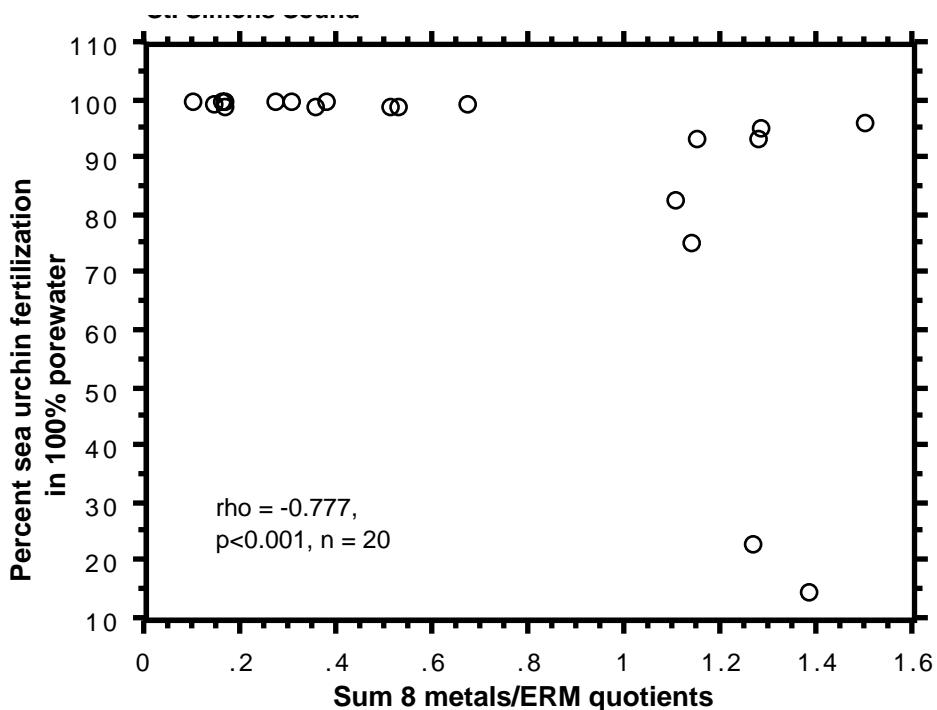


Figure 111. Relationship between percent sea urchin fertilization and the sum of 8 metals/ERM quotients in sediments from St. Simons Sound.

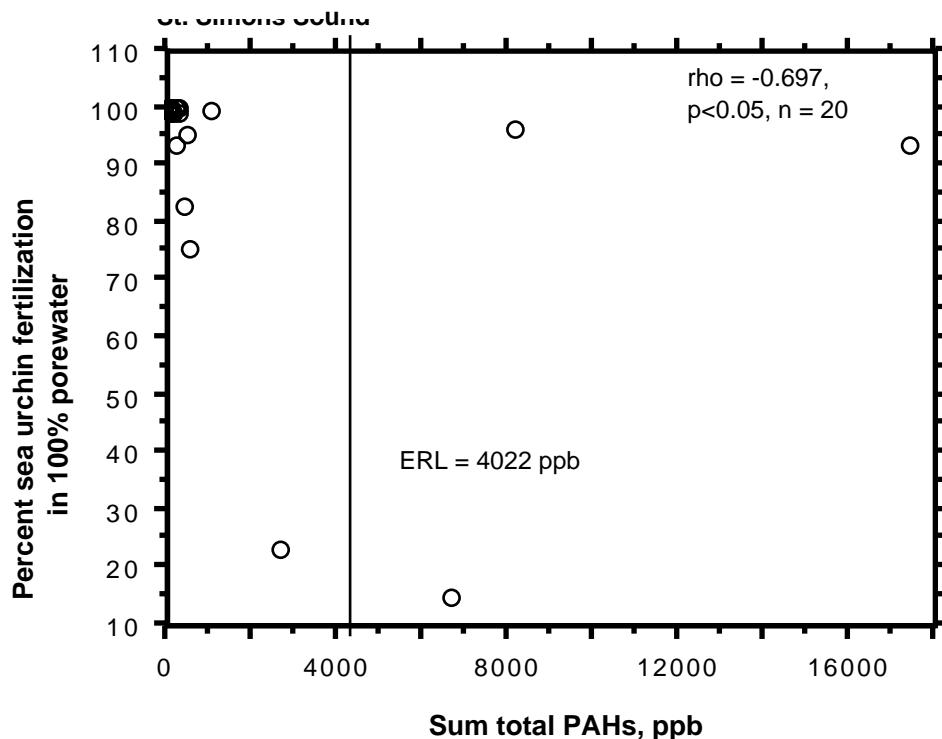


Figure 112. Relationship between sea urchin fertilization and the concentrations of total PAHs in sediments from St. Simons Sound.

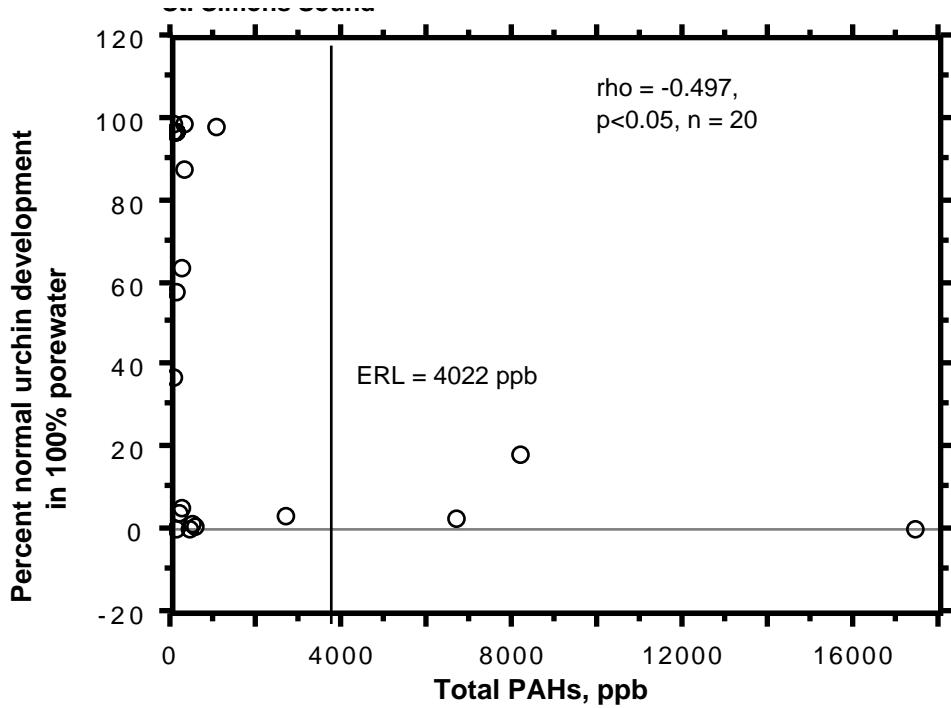


Figure 113. Relationship between normal sea urchin development and the concentrations of total PAHs in sediments from St. Simons Sound.

Table 1. Average, minimum and maximum concentrations of selected chemicals in sediments from different regions of the study area.

	Charleston Harbor n=65	St. Simons Sound n=20	Leadenwah Creek n=4 or 9*	Upper Savannah R. n=36	Lower Savannah R. n=18	Winyah Bay n=9
Silver (ug/g dry wt.)	0.12	0.08	<0.12	0.05	0.02	0.13
Chromium (ug/g dry wt.)	38.3	<0.02-0.48	<0.12-0.12	<0.02-0.12	0.02-0.05	<0.12-0.16
Lead (ug/g dry wt.)	4.8-258.3	42.9	46.9	43.5	3.2	74.4
Zinc (ug/g dry wt.)	2.5-73.9	5.0-89.5	13.8-66.6	5.3-75.9	4.3-64.4	28.1-112.8
Sum 5 SEM (umol/g dry wt.)	0.9	0.3	0.6	1	0.2	0.9
5SEM/AVS ratio	<0.01-15.3	<0.01-22.3	0.16-6.64	<0.01-126.6	<0.01-17.9	<0.01-2.6
Sum total PAHs (ng/g dry wt.)	2054	1974.4	178.4	1321.7	479.1	629.5
Sum DDTs (ng/g dry wt.)	19.1-9634	27-17544	33-248	77-4279	29-1185	157-1990
Sum PCBs (ng/g dry wt.)	0.7	3.7	1.3	2.8	<2.5	1.7
	<0.01-3.1	<2.5-15.9	0.5-3.0	<0.01-8.6	<2.5	0.5-5.2
	8	168.6	2.3	6.6	0.6	9
	<0.01-328.7	5.8-1775.9	<0.01-7.5	<0.01-29.3	<0.01-3.9	0.4-11.2

*trace metals concentrations for 4 samples and organics concentrations for 9 samples.

Table 2 Summary of toxicity test results and mean ERM quotients for sediment samples from Charleston Harbor, Winyah Bay, and Leadenwah Creek.

Station No.	% survival	<u>Ampelipod</u> % of control	Signif.	(mg/ml) signif.	Microtox	Sea urchin fertilization			Tox. tally	Mean ERM quotient
						100%	pw signif	50% pw signif		
Leadenwah Creek										
A 1-1	90	105.9	ns	0.02	**	9.6	ns	97	ns	87.8
A 1-2	91	107.1	ns	4.17	ns	97.4	ns	97.8	ns	92.2
A 1-8	82	96.5	ns	1.42	ns	97.4	ns	98.4	ns	98.6
A 2-1	87	102.4	ns	1.58	ns	99.4	ns	100	ns	99.6
A 2-3	87	102.4	ns	2.69	ns	99.6	ns	99.6	ns	98.8
A 2-7	88	103.5	ns	1.39	ns	99.2	ns	99	ns	99.4
A 3-1	86	101.2	ns	1.09	ns	97.8	ns	99.6	ns	99.8
A 3-2	87	102.4	ns	6.6	ns	9.9	ns	99.4	ns	99.2
A 3-3	86	101.2	ns	3.12	ns	9.6	ns	99.4	ns	99.2
Winyah Bay										
B 1-1	92	104.6	ns	0.29	*	88.4	++	96.2	ns	95.8
B 1-2	94	106.8	ns	0.00001	***	50.2	++	89	++	96.4
B 1-3	84	95.5	ns	0.01	**	74.8	++	95.4	ns	92.8
B 2-1	91	103.4	ns	7.66	ns	8.6	++	95.8	ns	97
B 3-1	93	105.7	ns	7.14	ns	98.4	ns	98.4	ns	97.4
B 4-1	94	106.8	ns	0.0001	**	2.8	++	23	++	63.6
B 5-1	88	100.0	ns	0.05	**	5	++	57.3	++	56.6
B 6-2	93	105.7	ns	7.03	ns	88.4	++	96.6	ns	95.2
B 7-1	92	104.6	ns	0.49	*	89.8	++	93.4	ns	86.6
Ashley River										
C 1-1	86	101.2	ns	9.8	ns	96.2	ns	97.8	ns	92.8
C 2-4	84	98.8	ns	2.92	ns	92.6	ns	95.6	ns	84.4
C 3-1	86	101.2	ns	8.46	ns	35.8	++	74.4	++	56.6
C 4-1	93	109.4	ns	9.43	ns	4.1	++	63.2	++	73.8
C 5-1	87	102.4	ns	5.83	ns	3.1	++	75.4	++	75.2
D 1-1	91	107.1	ns	0.63	ns	98.2	ns	99.4	ns	97.8
D 1-3	85	100.0	ns	2.94	ns	94.4	ns	97.6	ns	96.4

Table 2 (continued)

Station No.	Cooper River	<u>Amphipod</u>		<u>Microtox</u>		<u>Sea urchin</u>		<u>fertilization</u>		Tox. tally	Mean ERM quotient
		% survival	% of control	(mg/ml) signif.	100% pw signif	50% pw signif	25% pw signif				
D1-4	79	92.9	ns	7.45	ns	97.6	ns	99.6	ns	98.6	ns
D2-1	88	103.5	ns	2.03	ns	6.9	++	9.4	+	84.4	++
D2-2	90	105.9	ns	0.59	ns	29.6	++	55.8	++	54	++
D2-3	77	90.6	ns	2.32	ns	5.1	++	72.6	++	47	++
D3-1	81	95.3	ns	0.0004	* *	1.2	++	89.2	++	95.8	ns
D3-2	91	107.1	ns	0.49	ns	85.2	++	96.4	ns	96.6	ns
D3-3	88	103.5	ns	2.71	ns	72.2	++	83.2	++	6.9	++
D4-1	84	98.8	ns	6.79	ns	94.2	ns	9.8	ns	95.4	ns
D4-2	89	104.7	ns	1.16	ns	8.1	++	92.6	++	87	++
D4-4	87	102.4	ns	0.65	ns	2.3	++	58.8	++	40.8	++
E1-1	81	95.3	ns	3.29	ns	9.5	ns	96.2	ns	95.2	ns
E1-2	85	100.0	ns	6.71	ns	98.4	ns	99	ns	99	ns
E1-3	90	105.9	ns	7.49	ns	9.9	ns	99.6	ns	98.4	ns
E2-1	85	100.0	ns	0.06	* *	96.4	ns	9.4	+	95	ns
E2-2	84	98.8	ns	0.81	ns	92.2	ns	94.6	+	94	ns
E3-1	90	105.9	ns	7.24	ns	99.8	ns	99.6	ns	99.4	ns
E3-3	81	95.3	ns	0.89	ns	98.4	ns	98.8	ns	97.6	ns
G1-1	91	103.4	ns	0.28	*	58.8	++	9.4	ns	93.6	+
G2-1	91	103.4	ns	0.78	ns	79.6	++	91.4	+	94.2	ns
H1-1	92	104.6	ns	0.04	* *	19.4	++	80.4	++	83.8	++
H1-4	87	98.9	ns	5.8	ns	78.2	++	93.2	ns	94.4	++
H1-5	86	97.7	ns	0.84	ns	70	++	92.4	+	95.6	ns
H2-2	86	97.7	ns	7.55	ns	91.8	ns	96.6	ns	95.2	ns
H2-3	88	100.0	ns	0.62	*	9.8	ns	98.4	ns	98.8	ns
H2-6	86	97.7	ns	0.17	*	98.2	ns	98	ns	96	ns
H3-1	91	103.4	ns	7.12	ns	97.4	ns	98.2	ns	98.8	ns
H3-3	89	101.1	ns	0.47	*	94.4	ns	97.8	ns	97.6	ns

Table 2 (continued)

Station No.	% survival	<u>Amphipod</u>	% of control	Signif.	Microtox				Sea urchin fertilization				Tox. tally	Mean ERM quotient
					(mg/ml)	signif	100%	pw signif	50%	pw signif	25%	pw signif		
H3-5	93	105.7	ns	0.89	ns	82.8	++	97.2	ns	98	ns	ns	2	0.108
H4-2	91	103.4	ns	3.03	ns	96.8	ns	97.2	ns	95.8	ns	0	0.119	
H4-3	91	103.4	ns	7.22	ns	94	ns	95	ns	94.8	ns	0		
H4-5	94	106.8	ns	4.14	ns	91	+	96.2	ns	96.6	ns	1	0.01	
H5-2	93	105.7	ns	3.84	ns	92.8	ns	97	ns	92.6	++	2	0.013	
H5-4	90	102.3	ns	0.24	*	90.2	+	95.2	ns	94.4	ns	2	0.119	
H5-8	96	109.1	ns	1.58	ns	96.6	ns	95.8	ns	97.6	ns	0	0.148	
H6-1	92	104.6	ns	0.45	*	87.6	++	96.8	ns	93.8	+	2	0.177	
H6-2	88	100.0	ns	0.32	*	84.8	++	96.4	ns	98	ns	3	0.065	
H6-3	93	105.7	ns	7.29	ns	96.2	ns	97.6	ns	96.4	ns	0		
H7-1	88	100.0	ns	0.22	*	97	ns	99	ns	98.6	ns	2	0.108	
H8-1	93	105.7	ns	0.33	*	85.2	++	97.4	ns	98.4	ns	3	0.088	
Wando River														
F1-1	93	105.7	ns	5.29	ns	82.2	++	93.4	ns	86.8	++	4	0.133	
F1-2	97.5	110.8	ns	0.19	*	88.4	++	96.4	ns	95	ns	3	0.102	
F1-3	93	105.7	ns	7.3	ns	94.4	ns	95.6	ns	94.8	ns	0	0.078	
F2-1	87	98.9	ns	4.38	ns	90.6	+	92	ns	84.6	++	3	0.062	
F2-2	90	102.3	ns	7.53	ns	87.2	++	95	ns	93.2	++	4	0.064	
F2-4	91	103.4	ns	3.63	ns	94	ns	97.4	ns	94.8	ns	0	0.027	
North Inlet														
NOL	84	95.5	ns	3.02	ns	97.6	ns	97.2	ns	93.2	+	1	0.101	
NOL	90	105.9	ns	1.46	ns	99.8	ns	99.8	ns	99.2	ns	0	0.128	
NOL	85	88	ns			97.6	ns	98.2	ns	98.8	ns	0		
NOL	89	88	ns			97.4	ns	99.2	ns	98.8	ns	0		

Table 2 (continued)

Station No.	Charleston Harbor Project	Amphipod			Microtox			Sea urchin			fertilization			Tox. tally	Mean ERM quotient
		% survival	% of control	Signif.	(mg/ml)	signif.	100%	pw	signif.	50%	pw	signif.	25%	pw	
CHP1	91	103.4	ns		3.76	ns	68.8	++	91.2	++	94.6	ns	4	0.172	
CHP2	90	105.9	ns		7.48	ns	99.8	ns	98.6	ns	99.4	ns	0	0.117	
CHP3	84	98.8	ns		0.02	* *	91.2	ns	98.6	ns	97.6	ns	2	0.026	
CHP4	83	97.7	ns		1.16	ns	66.6	++	71.4	++	68.8	++	6	0.043	
CHP5	85	100.0	ns		3.88	ns	87.6	++	92.4	++	91	++	6		
CHP6	82	96.5	ns		4.92	ns	42.4	++	66.4	++	56.8	++	6	0.018	
CHP7	85	100.0	ns		0.03	* *	5	++	70.6	++	89.8	++	8		
CHP8	87	102.4	ns		2.58	ns	92.2	ns	93	++	90.8	++	4	0.096	
CHP9	91	103.4	ns		1.01	ns	91.4	+	95.4	ns	97.2	ns	1	0.016	
CHP10	95	108.0	ns		0.002	* *	89.2	++	95.2	ns	93.4	+	5	0.067	
CHP11	88	100.0	ns		3.45	ns	87.6	++	91.6	++	84.3	++	6	0.08	

ns non-significant

^ NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnets

*** statistically significant for Mann-Whitney, Dunnets, and Distribution-free

Table 3. Summary results of cytochrome P-450 RGS assays and meiobenthic copepod reproduction bioassays on selected samples from Charleston Harbor and vicinity.

STATION No.	P-450 RGS Assay CAS Lab.Id.	P-450 RGS Average fold (10ul)	B (a) P Equiv. (ug/g)	copepodite production	Signif naupliar production	Signif total production	Signif clutch size	Signif
<u>Leadenwah Creek</u>								
A1-8		61.67		327.00	388.67	14.00	*	
<u>Winyah Bay</u>								
B1-1	21	12.14	18.8					
B1-2	22	18.43	25.57					
B1-3	23	24.71	37.39					
B2-1	24	9.81	11.97					
B3-1	25	4.19	4.22	2.00	5.33	95.33		
B4-1	26	5.10	4.59					
B5-1	27	4.29	1.76					
B6-2	28	4.76	4.57					
B7-1	29	26.62	37.4					
<u>Charleston Harbor</u>								
CHP3		59.67		434.33	494.00	13.96	*	
CHP4		1.75	*	162.00	* 163.75	8.79	*	
CHP6		13.00		129.67	142.67	12.06	*	
D1-1	19	37.71	18.68					
D1-3	20	48.67	24.44					
D1-4				11.33	223.00	234.33	10.48	*
D2-2	18	118.38	48.55	23.00	235.00	258.00	13.56	*
D3-1	14	67.70	42.19					
D3-2	16	50.00	55.14					
D3-3	15	67.90	86.33					
D4-1	17	56.52	80.48					
D4-2	13	38.80	17.93					
D4-4	12	36.40	12.42					

Table 3 continued

STATION No.	P-450 Assay CAS Lab.Id.	RGS Average fold (10u)	P-450 RGS (ug/g)	P-450 RGS (ug/g)	B (a) P Equiv. production	copepodite Signif production	naupliar Signif production	total Signif production	clutch size	Signif
E2-2										*
E3-1	2	32.90	36.21		1.00	116.67	117.67	9.93		*
G1-1	3	59.70	70.39	18.00	0.00	101.00	101.00	11.90		
G2-1										
H1-1	1	15.67	7.31							
H1-4	4	14.67	5.31							
H1-5	5	37.90	47.81							
H2-2	6	10.00	3.5							
H2-3	7	26.10	18.42							
H3-1										
H4-5										
H5-4	8	7.76	3.19							
H5-8										
H6-1	9	9.19	3.38	15.33	140.00	155.33	155.33	13.41		
H6-2	10	10.38	10.32							
H8-1	11	7.43	9.09	7.00	175.00	182.00	182.00	13.17		
NIOL Ref 1	batch 1			58.00	319.67	377.67	377.67	17.80		
NIOL Ref 1	batch 2				17.25	261.75	261.75	13.55		
NIOL Ref 1	batch 3				14.67	250.67	250.67	15.39		
USC lab cts.	batch 2				3.33	203.00	206.33	13.92		
USC lab cts.	batch 3				44.25	329.75	385.00	12.06		

* significantly different from controls ($p<0.05$)

Table 4. Summary of toxicity test results and mean ERM quotients for sediment samples from the Savannah River.

Station No.	Amphipod % control	Microtox mg/ml	Sea urchin fertilization						Sea urchin development						Tox. tally	Mean ERM quotient
			signif	100% pw	signif	50% pw	signif	25% pw	signif	100% pw	signif	50% pw	signif	25% pw		
Upper Savannah River																
A1-1	4	@~	3.89	ns	99.6	ns	99.2	ns	98.6	ns	83.2	ns	87.8	ns	88.4	ns
A1-2	89	@	3.87	ns	99	ns	99.4	ns	98.8	ns	93	ns	94	ns	91.4	ns
A1-3	84	@	0.23	**	99	ns	98.4	ns	99.4	ns	95	ns	94.6	ns	92.4	ns
A2-2	87	ns	0.68	ns	99.8	ns	99.2	ns	99	ns	90.4	ns	90.4	ns	91.6	ns
A2-3	95	ns	0.23	**	98.4	ns	99.4	ns	98.4	ns	25.2	++	92.4	ns	94	ns
A2-4	90	@	3.98	ns	99.6	ns	99.2	ns	99.8	ns	88.8	ns	92.6	ns	95.6	ns
Downtown Savannah																
B1-1	101	ns	3.82	ns	99	ns	99	ns	98.6	ns	93.2	ns	94.8	ns	95.2	ns
B1-2	100	ns	3.79	ns	99.2	ns	99	ns	98.2	ns	92	ns	91.4	ns	91.4	ns
B1-5	103	ns	3.84	ns	98.8	ns	99.6	ns	98.6	ns	92.8	ns	93.4	ns	93.6	ns
B2-1	99	ns	0.17	**	3.4	++	86.7	ns	96.8	ns	0	++	0	++	0	++
B2-3	99	ns	0.18	**	5.2	++	67	++	95	ns	0.8	++	0	++	49.6	++
B2-4	99	ns	0.57	ns	7.4	++	88	ns	98.2	ns	0	++	0	++	87.2	ns
B3-3	101	ns	0.12	**	1.4	++	78.6	++	98.4	ns	0.8	++	0.2	++	88	ns
B3-5	104	ns	0.20	**	0.8	++	53.2	++	96.6	ns	0	++	0	++	0.2	++
B3-6	97	ns	0.34	*	0.6	++	72.8	++	96.8	ns	0	++	0	++	16.4	++
B4-4	104	ns	0.15	**	98.6	ns	99.4	ns	98.4	ns	92.8	ns	95.6	ns	93.6	ns
B4-5	108	ns	0.15	**	63	++	94.2	ns	96.2	ns	0	++	0.6	++	95.4	ns
B4-6	102	ns	0.38	*	55.4	++	95	ns	98.8	ns	0	++	0	++	93.6	ns
B5-1	102	ns	5.08	ns	95	ns	99.4	ns	99	ns	0	++	76.8	+	94.8	ns
B5-2	104	ns	6.77	ns	93	ns	98.2	ns	98.2	ns	0	++	62.4	++	91.2	ns
B5-3	101	ns	0.08	***	8.6	++	87.4	ns	94.8	ns	0	++	0	++	2.8	++
B6-1	101	ns	6.38	ns	98.4	ns	98.8	ns	99.6	ns	0	++	92.5	ns	89.8	ns
B6-2	96	ns	0.80	ns	97.8	ns	99.8	ns	99.2	ns	0	++	75	++	88.8	ns
B6-3	98	ns	0.12	**	95.8	ns	98.2	ns	98.8	ns	0	++	0	++	88.8	ns
B7-1	101	ns	0.07	***	71.8	++	96.6	ns	96.4	ns	0	++	0.6	++	93.6	ns
B7-2	96	ns	3.93	ns	95	ns	99.2	ns	98.6	ns	0	++	73	++	93	ns
B7-3	109	ns	5.16	ns	87	ns	98	ns	99	ns	0	++	92.6	ns	95.4	ns
B8-1	98	ns	3.82	ns	95.4	ns	98	ns	99.6	ns	0	++	25.4	++	93.2	ns
B8-2	99	ns	4.22	ns	98.4	ns	99.2	ns	98.8	ns	0.4	++	85.2	ns	93.8	ns

Table 4. (continued)

Station No.	Amphipod % control	Microtox (mg/ml) signif	Sea urchin fertilization						Sea urchin development						Tox. tally	Mean ERM quotient		
			100% pw	50% pw	signif	25% pw	signif	100% pw	signif	2.2	0.2	+++#	56.6	++#				
B8-3	97	ns	0.40	ns	10.4	+++#	80.2	ns	99.4	ns#	0	+++#	0	+++#	10	0.067		
B9-1	100	ns	0.11	**	48.4	+++#	96.8	ns	99.8	ns#	0	0	0	91.4	ns#	8	0.061	
B9-2			0.21	**														
B9-3	100	ns	0.10	**	31.6	+++#	97.4	ns#	99.0	ns#	0	+++#	0	+++#	94.2	ns#	8	0.057
Back River																		
C1	91	@	0.07	**	99.8	ns	99.6	ns	95.2	ns	91.4	ns	98	ns	98.6	ns	3	0.056
C2	91	ns	0.08	**	99.2	ns	99.8	ns	98.8	ns	64.4	++	97.6	ns	96.6	ns	4	0.094
C3	97	ns	5.03	ns	100.0	ns	99.6	ns	100.0	ns	98.6	ns	98	ns	96.4	ns	0	0.012
Lower Savannah River																		
D1,1-1	91	ns	0.09	**	100.0	ns	100.0	ns	99.4	ns	0	++	91.6	ns	97.6	ns	4	0.047
D1,2-2	101	ns	4.97	ns	97.2	ns	99.2	ns	99.2	ns	96.4	ns	98.8	ns	98.8	ns	0	0.038
D1,3-1	97	ns	0.09	**	71.2	++	98.6	ns	98.6	ns	3.6	++	0	++	14.6	++	10	0.051
D2,1-1	94	ns	5.04	ns	99.0	ns	99.6	ns	99.4	ns	96.2	ns	97.8	ns	98.4	ns	0	0.044
D2,2-2	96	ns	7.53	ns	98.6	ns	99.2	ns	99.4	ns	98.2	ns	98.2	ns	98.2	ns	0	0.024
D2,3-1	87	ns	0.61	*	97.6	ns	98.4	ns	96.8	ns	98.2	ns	97	ns	98.2	ns	1	0.019
D3,1-4	88	@	3.88	ns	99.6	ns	100.0	ns	99.8	ns	97.4	ns	98.4	ns	96.8	ns	1	0.013
D3,2-1	100	ns	2.25	ns	99.6	ns	99.6	ns	99.4	ns	98	ns	97.8	ns	98.8	ns	0	0.013
D3,3-2	96	ns	0.16	**	98.0	ns	99.4	ns	99.6	ns	19	++	0	++	97.2	ns	6	0.037
South Channel																		
E1,2-1	100	ns	0.56	ns	100.0	ns	100.0	ns	99.4	ns	96.8	ns	98.6	ns	98	ns	0	0.023
E1,3-1	100	ns	4.84	ns	100.0	ns	99.6	ns	99.6	ns	97.6	ns	98.2	ns	97.6	ns	0	0.044
E1-1	100	ns	0.38	ns	99.8	ns	99.8	ns	99.6	ns	64.6	++	98.4	ns	97.4	ns	2	0.029
E2-1	94	ns	5.24	ns	99.6	ns	99.4	ns	99.6	ns	97.8	ns	98.6	ns	98.2	ns	0	0.041
E2-2	102	ns	3.82	ns	99.6	ns	99.4	ns	99.4	ns	98.4	ns	98.2	ns	98	ns	0	0.041
E2-3	99	ns	0.29	ns	100.0	ns	99.2	ns	98.8	ns	36.8	++	96.8	ns	97.6	ns	2	0.048
E3-2	94	ns	0.24	*	99.6	ns	99.8	ns	99.0	ns	97.8	ns	95.8	ns	98.8	ns	1	0.032
E3-5	95	ns	0.01	**	95.0	ns	99.6	ns	99.2	ns	1	++	0	++	79.2	++	8	0.051
E3-9	96	ns	0.12	**	99.8	ns	99.4	ns	100.0	ns	3	++	0	++	97.2	ns	6	0.053

Table 4. (continued)

Station No.	% control	Amphipod signif	Microtox (mg/ml)	Sea urchin fertilization				Sea urchin development				Tox. tally	Mean ERM quotient	
				100% pw	signif	50% pw	signif	25% pw	signif	100% pw	signif	50% pw	signif	
Savannah harbors														
F1	101	ns	0.47	ns	7.2	++#	95.2	ns#	99.4	ns#	2.6	++#	29	++# 8
F2	101	ns	0.32	**	10.4	++#	93.2	ns#	97.0	ns#	2	++#	0.2	++# 10
G1	97	ns	0.16	**	72.8	++	97.2	++	98.8	ns	0	++	0	0.088
G2	102	ns	0.59	ns	86.8	ns	94.4	ns	98.2	ns	0.2	++	0	0.114
H1	103	ns	0.25	**	11	++	94.6	ns	98.2	ns	0	++	0	0.091
H2	98	ns	0.21	**	88.2	ns	96.4	ns	99.4	ns	0	++	0.6	0.137
H3			0.19	**								++	89.8	ns 6
North Inlet														
Ref. No. Inlet 216			0.84	ns										
Ref. No. Inlet 242			0.82	ns										
Ref. No. Inlet 273			0.77	ns										
Ref. No. Inlet (Lower Savannah)														
Ref. No. Inlet (upper Savannah)														
Ref. Redfish Bay (Lower Savannah)														
Ref. Redfish Bay (upper Savannah)														

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.

MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnets, and Distribution-free

@ sample mean was statistically different from control

~ sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Table 5. Summary of toxicity test results and mean ERM quotients for sediment samples from the St. Simons Sound.

Station No.	Amphipod % control	Microtox mg/ml)	Sea urchin fertilization						Sea urchin development			Tox. tally	Mean ERM quotient
			signif	100% pw	signif	50% pw	signif	25% pw	signif	100% pw	signif		
upper Turtle River													
A1 -1	95	ns	2.68	ns	98.8	ns	99.6	ns	99.6	ns	52.8	++	98.6
A2 -2	96	ns	3.70	ns	99.0	ns	99.4	ns	96.6	ns	97.80	ns	98.2
A3 -1	95	ns	0.63	ns	100.0	ns	100.0	ns	63.4	++	96.4	ns	97.4
Brunswick Harbor													
B1 -1	98	ns	0.27	ns	14.8	++	65.4	++	99.0	ns	2.6	++	61.6
B2 -1	100	ns	0.58	ns	93.4	ns	99.2	ns	99.6	ns	0	++	85.4
East River													
C1 -1	101	ns	0.25	ns	23.0	++	93.2	ns	99.2	ns	3.25	++	7.4
C2 -1	105	ns	0.17	**	82.6	++	98.6	ns	99.6	ns	0	++	0
C3 -1	101	ns	3.27	ns	75.4	++	98.2	ns	99.6	ns	1	++	0
mid-Turtle River													
D1 -1	98	ns	3.45	ns	99.4	ns	99.2	ns	99.6	ns	58	++	96.6
D2 -1	98	ns	1.97	ns	99.8	ns	99.2	ns	99.4	ns	37.25	++	95.6
D3 -2	96	ns	3.80	ns	99.6	ns	99.2	ns	99.0	ns	96.4	ns	97.4
lower Turtle River													
E1 -1	95	ns	0.46	ns	99.0	ns	100.0	ns	99.8	ns	3.8	++	98.2
E2 -2	97	ns	0.17	**	99.0	ns	99.8	ns	98.8	ns	87.8	ns	97.4

Table 5. (continued)

E3-1	85	ns	1.68	ns	99.8	ns	100.0	ns	99.2	ns	98.6	ns	98.2	ns	98.6	ns	0	0.037
Terry Creek																		
F1-1	86	@	0.01	**	95.2	ns	99.4	ns	99.4	ns	1.6	++	60.6	++	98.4	ns	7	0.065
F2-1	0	@~	0.12	**	96.2	ns	99.8	ns	99.2	ns	18.5	++	97.6	ns	98.4	ns	6	0.225
St. Simons Sd.																		
G1-1	103	ns	0.02	**	93.2	ns	98.4	ns	100.0	ns	5.4	++	0	++	95.8	ns	6	0.069
G2-1	99	ns	5.32	ns	99.8	ns	100.0	ns	99.6	ns	96.6	ns	97.2	ns	99	ns	0	0.014
G3-1	96	ns	1.00	ns	99.8	ns	99.6	ns	97.2	ns	98.6	ns	98.6	ns	98.4	ns	0	0.022
Purvis Creek																		
P1	98	ns	1.61	ns	99.4	ns	99.6	ns	99.8	ns	98	ns	99	ns	98.4	ns	0	0.470
North Inlet																		
Ref. No. Inlet	0.84	ns																
Ref. No. Inlet	0.82	ns																
Ref. Redfish Bay																		
Ref. No. Inlet																		

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnnett's

*** statistically significant for Dunnnett's, and Distribution-free

@ sample mean was statistically different from control

~ sample mean 80% or less than the control

Table 6. Incidence (percent of total samples tested) of significantly toxic samples from each of six estuarine regions.

Region	Amphipod survival* bioluminescence**	Microbial			Urchin fertilization***			All tests Urchin development*** All tests Urchin development*** All tests Urchin development*** All tests Urchin development*** All tests Urchin development*** All tests Urchin development***
		100% pw.	50% pw.	25% pw.	100% pw.	50% pw.	25% pw.	
Leadenwah Creek (n=9)	0.0	11.1	0.0	0.0	22.2	nd	nd	6.7
Winyah Bay (n=9)	0.0	66.6	88.9	33.3	44.4	nd	nd	46.7
Charleston Harbor (n=65)	0.0	25.4	50.8	33.8	40.0	nd	nd	30.5
Savannah River (n=60)	10.0	49.2 ^a	31.7	8.3	0.0	63.9	47.5	21.3
St. Simons Sound (n=20)	10.0	25.0	20.0	5.0	0.0	65.0	35.0	20.0
North Inlet (n=4)	0.0	0.0	0.0	25.0	100 ^b	0.0 ^b	0.0 ^b	11.5

* one-way t-test ($p<0.05$)

** Mann-Whitney test ($p<0.05$)

*** Dunnett's one-tailed test ($p<0.05$)

nd = no data

^a data for station B9-2 excluded

^b for urchin development tests, n=2

Table 7. Spearman-rank correlations (rho, corrected for ties) among toxicity tests performed on sediments from Charleston Harbor, Winyah Bay, Leadenwah Creek, Savannah River, and St. Simons Sound.

Charleston Harbor/Winyah Bay/Leadenwah Creek

	Amphipod survival	Microtox	Urchin fertilization
Microtox	0.115 ns		
Urchin fertilization	0.221 ns	0.389 * *	
P450 RGS (B[a]p equivalents)	-0.045 ns	0.082 ns	-0.112 ns
Copepod:			
• clutch size	0.265 ns	-0.260 ns	0.101 ns
• nauplii production	-0.389 ns	-0.377 ns	-0.224 ns
• copepodite production	-0.230 ns	-0.425 ns	-0.361 ns

Savannah River

	Amphipod survival	Microtox	Urchin fertilization
Microtox	0.158 ns		
Urchin fertilization	-0.329 *	0.312 *	
Urchin development	-0.345 *	0.280 *	0.664 * * *

St. Simons Sound

	Amphipod survival	Microtox	Urchin fertilization
Microtox	0.012 ns		
Urchin fertilization	-0.447 *	0.502 *	
Urchin development	-0.315 ns	.390 ns	0.732 * *

All areas combined

	Amphipod survival	Microtox	Urchin fertilization
Microtox	-0.031 ns		
Urchin fertilization	-0.308 * * *	0.388 * * *	
Urchin development	-0.368 * *	0.335 *	0.701 * * *

ns: p>0.05, *p<0.05, ** p<0.001, *** p<0.0001

Table 8. Estimates of the spatial extent of toxicity, expressed as kilometer² and percent of each area, based upon results of three toxicity tests*.

Survey Region	Total survey area (km ²)	Amphipod survival	Urchin fertilization	Microbial bioluminescence
Winyah Bay	7.3	0	3.1 (42.2%)	5.1 (70.0%)
Charleston Harbor	41.1	0	12.5 (30.4%)	17.6 (42.9%)
Leadenwah Creek	1.7	0	0	0.3 (20.1%)
Savannah River	13.1	0.2 (1.2%)	2.4 (18.4%)	7.5 (57.1%)
St. Simons Sound	24.6	0.1 (0.4%)	0.7 (2.6%)	11.4 (46.4 %)
Total area	87.8	0.3 (0.3%)	18.7 (21.3%)	41.9 (47.7%)

* Based upon bioassay responses <80% of controls

Table 9. Spearman rank correlations (Rho, corrected for ties) between measures of toxicity and chemical concentrations in sediments from Charleston Harbor, Winyah Bay, and Leadenwah Creek (n=66).

Chemical	Amphipod survival	Microbial bioluminescence	Urchin fertilization	P-450 RGS bioassay
P450 RGS	-.165	ns	.146	ns
UAN (amphipod-start)	-.096	ns	na	na
UAN (amphipod-end)	-.094	ns	na	na
UAN (urchin)	na	na	-.151	ns
silver	.040	ns	-.124	ns
aluminum	.228	ns	-.375	*
arsenic	.217	ns	-.301	*
cadmium	.055	ns	-.237	ns
chromium	.217	ns	-.343	*
copper	.219	ns	-.421	*
iron	.290	ns	-.354	*
mercury	.173	ns	-.195	ns
manganese	.273	ns	-.267	*
nickel	.215	ns	-.355	*
lead	.186	ns	-.318	*
selenium	.216	ns	-.299	*
tin	.269	ns	-.277	*
zinc	.218	ns	-.373	*
percent fines	.253	ns	-.412	*
percent toc	.240	ns	-.418	*
total 5 SEM	.069	ns	-.298	*
SEM:AVS ratio	.120	ns	.679	***
total gc PAHs	.015	ns	-.276	*
sum 8 LPAHs	.001	ns	-.320	*
sum 16 HPAHs	.022	ns	-.262	*
total PCBs	.071	ns	-.345	*
total pesticides	.076	ns	-.308	*
total DDTs	.083	ns	-.239	*
sum 9 metals/ERMs	.160	ns	-.356	*
sum 3 coh/ERMs	.024	ns	-.315	*
sum 13 pahs/ERMs	.031	ns	-.269	*
mean of 25 ERM quot.	.031	ns	-.269	*
ns = non-significant (p>0.05)				

* p<0.05; ** p<0.001; *** p<0.0001

UAN = un-ionized ammonia

SEM = simultaneously extracted metals

AVS = acid-volatile sulfides

TOC = total organic carbon

PAHs = polynuclear aromatic hydrocarbons

PCBs = polychlorinated biphenyls

ERMs = effects range median values

Table 10. Spearman rank correlations (Rho, corrected for ties) between measures of toxicity and chemical concentrations in sediments from the Savannah River (n=61).

Chemical	Amphipod survival	Microbial		Urchin fertilization	Urchin development
		bioluminescence	na		
UAN (amphipod-start)	.433	**	na	na	na
UAN (amphipod-end)	.320	*	na	na	na
UAN (urchin)	na	na		-.733	***
silver	-.122	ns	.030	ns	-.178
aluminum	.055	ns	-.034	ns	-.550
arsenic	-.001	ns	-.166	ns	-.353
cadmium	.266	ns	-.417	*	-.538
chromium	.170	ns	-.108	ns	-.560
copper	.041	ns	-.061	ns	-.614
iron	.153	ns	.085	ns	-.627
mercury	-.013	ns	-.145	ns	-.611
manganese	.126	ns	.018	ns	-.660
nickel	.077	ns	-.035	ns	-.567
lead	.097	ns	-.136	ns	-.638
selenium	-.121	ns	-.108	ns	-.196
tin	.010	ns	-.490	ns	-.523
zinc	.153	ns	-.181	ns	-.669
percent fines	.158	ns	-.050	ns	-.631
percent toc	.146	ns	.155	ns	-.565
total 5 SEM	.199	ns	-.425	*	-.634
SEM:AVS ratio	.106	ns	-.857	***	-.328
total gc PAHs	.115	ns	.039	ns	-.576
sum 8 LPAHs	.230	ns	.000	ns	-.346
sum 16 HPAHs	.117	ns	-.037	ns	-.631
total PCBs	.077	ns	-.075	ns	-.518
total pesticides	.163	ns	.353	*	-.144
total DDTs	.045	ns	.221	ns	-.137
sum 9 metals/ERMs	.166	ns	-.179	ns	-.665
sum 3 cohgs/ERMs	.084	ns	-.067	ns	-.514
sum 13 pahs/ERMs	.250	ns	-.144	ns	-.560
mean of 25 ERM quotients	.171	ns	-.161	ns	-.700

ns = non-significant ($p>0.05$); * $p<0.05$; ** $p<0.001$; *** $p<0.0001$

UAN = un-ionized ammonia. TOC = total organic carbon.

SEM/AVS = simultaneously extracted metals/acid-volatile sulfides.

PAHs = polynuclear aromatic hydrocarbons; PCBs = polychlorinated biphenyls.

cohgs = chlorinated organic hydrocarbons

ERMs = effects range median values

Table 11. Spearman rank correlations (Rho, corrected for ties) between measures of toxicity and chemical concentrations in sediments from St. Simons Sound (n=20).

Chemical	Amphipod survival		Microbial bioluminescence		Urchin fertilization		Urchin development	
UAN (amphipod-start)	0.450	ns	na		na		na	
UAN (amphipod-end)	0.206	ns	na					
UAN (urchin)				-0.546	*	-0.645	*	
silver	0.424	ns	-0.256	ns	-0.664	*	-0.564	*
aluminum	0.282	ns	-0.736	**	-0.793	**	-0.484	*
arsenic	0.226	ns	-0.679	*	-0.593	*	-0.407	ns
cadmium	-0.088	ns	-0.269	ns	-0.172	ns	-0.140	ns
chromium	0.375	ns	-0.660	*	-0.868	**	-0.592	*
copper	0.165	ns	-0.692	*	-0.815	**	-0.580	*
iron	0.346	ns	-0.733	**	-0.763	**	-0.470	*
mercury	0.300	ns	-0.538	*	-0.665	*	-0.582	*
manganese	0.335	ns	-0.662	*	-0.775	**	-0.586	*
nickel	0.206	ns	-0.769	**	-0.803	**	-0.567	*
lead	0.206	ns	-0.703	*	-0.736	*	-0.440	ns
selenium	0.345	ns	-0.081	ns	-0.402	ns	-0.321	ns
tin	0.417	ns	-0.570	ns	-0.867	*	-0.560	*
zinc	0.286	ns	-0.683	*	-0.778	**	-0.464	*
percent fines	0.328	ns	-0.635	*	-0.815	**	-0.583	*
percent toc	0.152	ns	-0.703	*	-0.777	**	-0.531	*
total 5 SEM	-0.031	ns	-0.836	***	-0.486	*	-0.295	ns
SEM:AVS ratio	-0.123	ns	0.774	**	0.411	ns	0.335	ns
total gc PAHs	0.159	ns	-0.587	*	-0.697	*	-0.497	*
sum 8 LPAHs	0.097	ns	-0.685	*	-0.665	*	-0.466	*
sum 16 HPAHs	0.142	ns	-0.542	*	-0.666	*	-0.520	*
total PCBs	0.423	ns	-0.062	ns	-0.246	ns	-0.135	ns
total pesticides	0.282	ns	0.049	ns	-0.103	ns	0.098	ns
total DDTs	0.361	ns	-0.283	ns	-0.358	ns	-0.016	ns
sum 9 metals/ERMs	0.185	ns	-0.742	*	-0.777	**	-0.510	*
sum 3 coh/ERMs	0.430	ns	-0.078	ns	-0.268	ns	-0.130	ns
sum 13 pahs/ERMs	0.194	ns	-0.601	*	-0.686	*	-0.441	*
mean 25 ERM quotients	0.286	ns	-0.608	*	-0.617	*	-0.396	ns

ns = non-significant ($p>0.05$); * $p<0.05$; ** $p<0.001$; *** $p<0.0001$

UAN = un-ionized ammonia. TOC = total organic carbon.

SEM/AVS = simultaneously extracted metals/acid-volatile sulfides.

PAHs = polynuclear aromatic hydrocarbons; PCBs = polychlorinated biphenyls.

coh = chlorinated organic hydrocarbons

ERMs = effects range median values

Table 12. Spearman rank correlations (Rho, corrected for ties) between measures of toxicity and chemical concentrations in sediments from all South Carolina/Georgia areas (n=147).

Chemical	Amphipod survival	Microbial bio-luminescence		Urchin fertilization	Urchin development
		bioluminescence	urchin		
UAN (amphipod-start)	.510	***	na	na	na
UAN (amphipod-end)	.487	***	na	na	na
UAN (urchin)				-.500	*** -.676 ***
sum 9 metals/ERMs	.238	*	-.356 ***	-.401	*** -.632 ***
sum 3 cohds/ERMs	-.303	**	-.001 ns	.055 ns	-.263 *
sum 13 pahs/ERMs	.060	ns	-.273 **	-.385 ***	-.537 ***
mean of 25 ERM quot.	.130	ns	-.295 **	-.414 ***	-.609 ***

ns = non-significant ($p>0.05$); * $p<0.05$; ** $p<0.001$; *** $p<0.0001$

UAN = un-ionized ammonia. TOC = total organic carbon.

PAHs = polynuclear aromatic hydrocarbons; PCBs = polychlorinated biphenyls.

cohds = chlorinated organic hydrocarbons

ERMs = effects range median values

Table 13. Numbers of samples (out of 140 analyzed) in which ERL and ERM values (from Long et al., 1995) were exceeded.

Chemical	Number of samples exceeding ERL value	Number of samples exceeding ERM value
silver	none	none
arsenic	6 4	none
cadmium	8	none
chromium	9	none
copper	5	none
mercury	2 0	2
nickel	3 3	none
lead	4	none
zinc	3	none
naphthalene	4	none
2-methyl naphthalene	1 0	none
acenaphthylene	1 3	none
acenaphthene	4 4	none
fluorene	3 9	none
phenanthrene	2 1	none
anthracene	2 5	1
fluoranthene	1 1	none
pyrene	1 1	1
benzo(a)anthracene	1 2	none
chrysene	1 2	none
benz(a)pyrene	6	none
dibenzo(a,h)anthracene	6	none
sum LPAHs	8	2
sum HPAHs	3 1	none
total PAHs	1 6	none
p,p'-DDE	1	none
total DDTs	3 2	none
total PCBs	1 6	none

Acknowledgments

Funding for this survey was provided by the Coastal Ocean Program and the National Status and Trends Program of NOAA. The Charleston Harbor Project (Mr. Heyward Robinson) funded the analyses of selected samples from the Charleston Harbor area. The state of Florida Department of Environmental Protection assisted in the collection of samples (Ms. Gail M. Sloane). Dr. Kevin Summers (U.S. EPA, Gulf Breeze, FL) assisted in survey design. All Microtox™ bioassays and all chemical analyses were performed by the National Marine Fisheries Service, Southeast Fisheries Science Center in Charleston (Dr. Geoffrey I. Scott, Dr. Michael Fulton, Dr. John Kucklick, Mr. Brian Thompson). Sea urchin tests and the 1993 amphipod tests were performed by the National Biological Service laboratory in Corpus Christi (Dr. R. Scott Carr, Mr. Duane Chapman, Mr. Jim Biedenbach). Amphipod survival tests were performed in 1994 by Science Applications International Corporation (Dr. K. John Scott, Dr. Glen B. Thursby, Ms. Cornelia Weber). Tests of copepod reproduction were performed by the University of South Carolina in Columbia (Dr. G. Thomas Chandler). Cytochrome P-450 RGS assays were performed by Dr. Jack Anderson, Columbia Analytical Services, Carlsbad, CA. Ms. Jo Linse provided valuable patience, assistance, and hard work in the collection of samples, analyses of data, and preparation of study designs and graphics.

References

- Alexander, C., J. Ertel, R. Lee, B. Loganathan, J. Martin, R. Smith, S. Wakeham, and H. Windom. 1994. Pollution history of the Savannah Estuary. Skidaway Institute of Oceanography. Savannah, GA. 29 pg.
- Allen, H. E., F. Gongmin, and B. Deng. 1993. Analysis of acid volatile sulfide (AVS) and simultaneously-extracted metals (SEM) for estimation of potential toxicity in aquatic systems. Environ. Toxicol. Chem 12: 1441-1453.
- Anderson, J. W., S. S. Rossi, R. H. Tukey, T. Vu, and L. C. Quattrochi. 1995. A biomarker, 450 RGS, for assessing the potential toxicity of organic compounds in environmental samples. Envir. Toxicol. & Chem. 14(7): 1159-1169.
- Anderson, J. W., S. S. Rossi, R. H. Tukey, T. Vu, and L. C. Quattrochi. 1995. A biomarker, P450 RGS, for assessing the induction of potential of environmental samples. Envir. Toxicol. & Chem. 14(7): 1159-1169.
- Anderson, J. W., K. Bothner, T. Vu, and R. H. Tukey. 1996. Using a biomarker (P450 RGS) test method on environmental samples. pp. 277-286. Chapter 15 in: Techniques in Aquatic Toxicology. Editor - G. K. Ostrander. Lewis Publishers, Boca Raton, FL.
- APHA. 1996. P450 reporter gene responses to dioxin-like organics. Method 8070, pp. 24-25. In: Standard Methods for the Examination of Water and Wastewater. 19th edition supplement. American Public Health Association, Washington, D.C.

ASTM. 1990. Standard Guide for Conducting 10-day Static Toxicity Test with Marine and Estuarine Amphipods. Designation E 1367-92. Annual Book of Standards. 11.04. American Society for Testing and Materials. Philadelphia, PA.

ASTM. 1992. Standard Guide for Conducting 10-day Static Toxicity Test with Marine and Estuarine Amphipods. Designation E 1367-92. Annual Book of Standards. 11.04. American Society for Testing and Materials. Philadelphia, PA.

ASTM. 1997. E 1853 standard guide for measuring the presence of planar organic compounds which induce CYP1A, reporter gene test systems. Vol. 1.05. American Society for Testing and Materials. Philadelphia, PA. In press.

Bronstein, S. 1995. Brunswick site's pollution 'off the chart'. The Atlanta Constitution. January 20, 1995.

Carr, R.S. 1993a. Sediment quality assessment survey of the Galveston Bay System. Galveston Bay National Estuary Program Report, GBNEP-30, 101 pp.

Carr, R.S. and D.C. Chapman. 1992 Comparison of solid-phase and pore-water approaches for assessing the quality of marine and estuarine sediments. *Chem. Ecol.* 7:19-30.

CFR. 1991. Definition and procedure for the determination of the method detection limit. Appendix B to Part 136. Code of Federal Register 40, Ch. 1 (7-1-91 Edition).

Carr, R.S., E. R. Long, H. L. Windom, D. C. Chapman, G. Thursby, G. M. Sloane, and D. A. Wolfe. 1996. Sediment quality assessment studies of Tampa Bay, Florida. *Envir. Toxicol. & Chem.* 15(7): 1218-1231.

Chandler, G. T. 1990. Effects of sediment-bound residues of the pyrethroid insecticide fenvalerate on survival and reproduction of meiobenthic copepods. *Mar. Envir. Res.* 29: 65-76.

Chandler, G. T. and G. I. Scott. 1991. Effects of sediment-bound endosulfan on survival, reproduction and larval settlement on meiobenthic polychaetes and copepods. *Environ. Toxicol. and Chem.* 10: 375-382.

Chapman, P. M. 1995. Do sediment toxicity tests require field validation? *Envir. Toxicol. & Chem.* 14 (9): 1451-1453.

Chapman, P. M., R. N. Dexter, and E. R. Long. 1987. Synoptic measures of sediment contamination, toxicity and infaunal community composition (the sediment quality triad) in San Francisco Bay. *Marine Ecology-Progress Series* 37: 75-96.

Davis, K. B. and R. F. Van Dolah. 1992. Characterization of the physical, chemical, and biological conditions and trends in the Charleston Harbor estuary: 1970-1985. Final Report. SC Sea Grant Consortium, Charleston, SC. 123 pg.

Fairey, R., J. Hunt, C. J. Wilson, F. LaCaro, M. Stephenson, M. Puckett and E. R. Long. 1996. Chemistry, toxicity and benthic community conditions in sediments of the San Diego Bay region. Final report from the California State Water Resources Control Board. Sacramento, CA. 169 pp.

Fulton, M. H. 1989. The effects of certain intrinsic and extrinsic variables on the lethal and sublethal toxicity of selected organophosphorus insecticides in the mummichog, *Fundulus heteroclitus* under laboratory and field conditions. Ph.D. Dissertation, University of South Carolina.

Fulton, M. H., G. I. Scott, A. R. Fortner, T. Bidleman, and B. Ngabe. 1993. The effects of urbanization on small high salinity estuaries of the Southeastern United States. *Arch. Envir. Contam. and Toxicol.*

Hamilton, M. A., R. C. Russo, and R. V. Thurston. 1977. Trimmed Spearman-Karber method for estimating median lethal concentrations in toxicity bioassays. *Environ. Sci. Technol.* 11: 714-719.

Hanson, P. J. and D. W. Evans. 1991. Metal contaminant assessment for the Southeast Atlantic and Gulf of Mexico coasts: Results of the National Benthic Surveillance Project over the first four years 1984-1987. NOAA Technical Memorandum NMFS-SEFSC 284. National Marine Fisheries Service, Beaufort, NC. 22 pg.

Heimbuch, D., H. Wilson, J. Seibel, and S. Weisberg. 1995. R-EMAP data analysis approach for estimating the proportion of area that is subnominal. Informal report prepared for V. Serveiss, U. S. EPA, Research Triangle Park, NC. 22 pg.

Holland, A. F., editor. 1990. Near coastal program plan for estuaries. U. S. EPA 600.14-90-033. U. S. EPA Office of Research and Development, ERL-Narragansett, R.I.

Hyland, J. L., T. J. Herrlinger, T. R. Snoots, A. H. Ringwood, R. F. VanDolah, C. T. Hackney, G. A. Nelson, J. S. Rosen, and S. A. Kokkinakis. 1996. Environmental quality of estuaries of the Carolinian Province: 1994. NOAA Tech. Memo. NOS ORCA 97. National Oceanic and Atmospheric Administration, Charleston, S.C. 102 pg.

Johnson, B.T. 1992. An evaluation of genotoxicity assay with liver S9 for activation and luminescent bacteria for detection. *Environ. Toxicol. Chem.* 11:473-480.

Johnson, B.T. 1993. Activated Mutatox® Assay for detection of genotoxic substances. *Environ. Toxicol. Water Qual.* 8:108-113.

Kohn, N. P., J.Q. Word, and D. K. Niyogi. 1994. Acute toxicity of ammonia to four species of marine amphipod. *Mar. Env. Res.* 38 (1994) 1-15.

Krahn, M..M., Wigren, C.A. , Pearce, R.W., Moore, L.K., Boger, R.G., Macleod, W.D. Jr., Chan, S.L., and Brown, D.W. 1988. New HPLC cleanup and revised extraction procedures for organic contaminants. NOAA Technical Memorandum NMFS F/MWC-153:23-47.

Kucklick, J. R. and T. F. Bidleman. 1994. Organic contaminants in Winyah Bay, South Carolina I: Pesticides and polycyclic aromatic hydrocarbons in subsurface and microlayer waters. *Marine Environ. Res.* 37: 63-78.

Lauenstein, G. G., M. R. Harmon, and B. W. Gottholm. 1993. National Status and Trends Program: Monitoring site descriptions (1984-1990) for the National Mussel Watch and Benthic Surveillance Projects. NOAA Technical Memorandum NOS ORCA 70. National Oceanic and Atmospheric Administration, Silver Spring, MD. 358 pg.

Lauenstein, G. G. and A. Y. Cantillo, editors. 1993. Sampling and analytical methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch projects. 1984-1992. NOAA Tech. Memo. NOS ORCA 71. National Oceanic and Atmospheric Administration. Silver Spring, MD.

Long, E. R. and P. M. Chapman. 1985. A sediment quality triad: Measures of sediment contamination, toxicity, and infaunal community composition in Puget Sound. *Mar. Pollu. Bull.* 16(10): 4-5-415.

Long, E. R. and L.G. Morgan. 1990. The potential for biological effects of sediment-sorbed contaminants tested in the National Status and Trends Program. NOAA Technical Memorandum NOS OMA 52. National Oceanic and Atmospheric Administration. Seattle, Washington. 175 pp. and appendices.

Long, E. R. and M. F. Buchman. 1989. An evaluation of candidate measures of biological effects for the National Status and Trends Program. NOAA Tech. Memo. NOS OMA 45. National Oceanic and Atmospheric Administration, Seattle, WA. 106 pg.

Long, E. R. and R. Markel. 1992. An evaluation of the extent and magnitude of biological effects associated with chemical contaminants in San Francisco Bay, California. NOAA Tech. Memo. NOS ORCA 64. National Oceanic and Atmospheric Administration, Seattle, WA. 86 pg.

Long, E. R., D. A. Wolfe, R. S. Carr, K. J. Scott, G. B. Thursby, H. L. Windom, R. Lee, F. D. Calder, G. M. Sloane, and T. Seal. 1994. Magnitude and extent of sediment toxicity in Tampa Bay, Florida. NOAA Tech. Memo. NOS ORCA 78. National Oceanic and Atmospheric Administration, Silver Spring, MD. 138 pg.

Long, E. R., D. A. Wolfe, K. J. Scott, G. B. Thursby, E. A. Stern, C. Peven, and T. Schwartz. 1995a. Magnitude and extent of sediment toxicity in the Hudson-Raritan Estuary. NOAA Tech. Memo. NOS ORCA 88. National Oceanic and Atmospheric Administration, Silver Spring, MD.

Long, E. R., D. D. MacDonald, S. L. Smith, and F. D. Calder. 1995b. Incidence of adverse biological effects within ranges of chemical concentrations in marine and estuarine sediments. *Environ. Mgt.* 19(1): 81-97.

Long, E. R., G. M. Sloane, R. S. Carr, K. J. Scott, G. B. Thursby, and T. Wade. 1996. Sediment toxicity in Boston Harbor: Magnitude, extent and relationships with chemical toxicants. NOAA Tech. Memo. NOS ORCA 96. National Oceanic and Atmospheric Administration, Silver Spring, MD. 133 pg.

Morgan, B.J.T. 1992. Analysis of quantal response data. Chapman and Hall. London, UK.

Moser, B. K. and G. R. Stevens. 1992. Homogeneity of variance in the two-sample means tests. American Statistician 46: 19-21.

National Oceanic and Atmospheric Administration. 1985. National Estuarine Inventory Data Atlas. Volume 1. Strategic Assessment Branch, Ocean Assessment Division.

National Oceanic and Atmospheric Administration. 1989. National Status and Trends Program for Marine Environmental Quality. Progress Report. A Summary of Data on Tissue Contamination from the First Three Years (1986-1988) of the Mussel Watch Project. NOAA technical Memorandum NOS OMA 49. Rockville, MD. 22 pp. & app.

National Oceanic and Atmospheric Administration. 1991. National Status and Trends Program for Marine Environmental Quality. Progress Report. Second Summary of Data on Chemical Contaminants in Sediments from the National Status and Trends Program. NOAA Technical Memorandum NOS OMA 59. Rockville, MD. 29 pp. & app.

National Oceanic and Atmospheric Administration. 1992. The National Status and Trends Program. Marine Environmental Quality. NOAA Special Publication. U. S. Department of Commerce, NOAA/NOS , Rockville, MD. 18 pg.

O'Connor, T. P. and B. Beliaeff. 1995. Recent trends in coastal environmental quality: Results from the Mussel Watch Project. National Status and Trends Program. NOAA, Silver Spring, MD. 40 pg.

O'Connor, T. P. and C. N. Ehler. 1991. Results from the NOAA National Status and Trends Program on distribution and effects of chemical contamination in the coastal and estuarine United States. Envir. Monit. & Assessment 17: 33-49.

O'Connor, T. P. 1991. Concentrations of organic contaminants in mollusks and sediments at NOAA National Status and Trend sites in coastal and estuarine United States. Envir. Health Perspectives 90: 69-73.

Plumb, R.H., Jr. 1981. Procedures for handling and chemical analysis of sediment and water samples. Tech. Rep. EPA CE-81-1. U.S. Army Engineer Waterways Experiment Station, CE, Vicksburg, Mississippi.

Sapudar, R. A., C. J. Wilson, M. L. Reid, E. R. Long, M. Stephenson, M. Puckett, R. Fairey, J. Hunt, B. Anderson, D. Holstad, J. Newman, and S. Birosik. 1994. Sediment chemistry and toxicity in the vicinity of the Los Angeles and Long Beach Harbors. California State Water Resources Control Board, Sacramento, CA. 77 pg.

SAS Institute, Inc. 1992. SAS/STAT User's Guide, Version 6, Fourth Edition, vol. 2. Cary, NC: SAS Institute, Inc. 846 pg.

Schimmel, S. C., B. D. Melzian, D. E. Campbell, C. J. Strobel, S. J. Benyi, J. S. Rosen, and H. W. Buffum. 1994. Statistical summary EMAP-Estuaries Virginian Province - 1991. EPA/620/R/94/005. U. S. Environmental Protection Agency, Narragansett, RI. 77 pp.

Schantz M.M., Benner B.A., Chesler S.N., Koster B.J., Hehn K.E., Stone S.F., Kelly WR, Zeisler R., and Wise S.A. 1990. Preparation and analysis of a marine sediment reference material for the determination of trace organic constituents. *Fresenius Zeitschrift fur Analytische Chemie* 338:501, 514.

Schropp, S.J. and H.L. Windom. 1988. A guide to the Interpretation of Metal Concentrations in Estuarine Sediments. Florida Department of Environmental Regulation, Tallahassee, Fl. 44 pp. & app.

Schropp, S. J., F. G. Lewis, H. L. Windom, J. D. Ryan, F. D. Calder, and L. C. Burney. 1990. Interpretation of metal concentrations in estuarine sediments of Florida using aluminum as a reference element. *Estuaries* 13(3): 227-235.

Scott, G. I. and others. 1990. Agricultural insecticide runoff effects on estuarine organisms: Correlating laboratory and field toxicity tests, ecophysiology bioassays and ecotoxicological biomonitoring. Final report submitted to U. S. EPA, Gulf Breeze Research Laboratory. 281 pg.

Scott, G. I. and others. 1993. Agricultural insecticide runoff effects on estuarine organisms: Correlating laboratory and field toxicity tests, ecophysiology assays and ecotoxicological biomonitoring. Final report submitted to U. S. EPA, Gulf Breeze Research Laboratory. 281 pg.

Sloane, G. M., E. R. Long, R. S. Carr, T. Johnson, J. Biedenbach, K. J. Scott, G. B. Thursby, E. Crecelius, C. Peven, H. L. Windom, R. D. Smith, and B. Loganathon. Magnitude and extent of sediment toxicity in four bays of the Florida panhandle: Pensacola, Choctawhatchee, St. Andrew and Apalachicola. NOAA Technical Memorandum NOS ORCA 117. National Oceanic and Atmospheric Administration, Silver Spring, MD.

Smith, R.G. 1993. Determination of mercury in environmental samples by isotope dilution/ICPMS. *Analytical Chemistry* 65:2485-2489.

South Carolina Sea Grant Consortium. 1992. Characterization of the physical, chemical, and biological conditions and trends in three South Carolina estuaries: 1970-1985. South Carolina Sea Grant Consortium, Charleston, SC.

Strawbridge,S., B. C. Coull, and G. T. Chandler. 1992. Reproductive output of a meiobenthic copepod exposed to sediment-associated fenvalerate. *Arch. Environ. Contam. Toxicol.* 23: 295-300.

Summers, J. K., J. M. Macauley, P. T. Heitmuller, V. D. Engle, A. Matt Adams, and G. T. Brooks. 1993. Statistical Summary: EMAP-Estuaries. Louisianian Province - 1991. U. S. EPA/600/R-93-001. U. S. Environmental Protection Agency, Gulf Breeze, FL. 101 pg.

Swartz, R. C., W. A. DeBen, K. A. Sercu, and J. O. Lamberson. 1982. Sediment toxicity and the distribution of amphipods in Commencement Bay, Washington, USA. *Mar. Pollu. Bull.* 13: 359-364.

Swartz, R. C., F. A. Cole, D. W. Schults, and W. A. DeBen. 1986. Ecological changes on the Palos Verdes shelf near a large sewage outfall: 1980-1983. *Marine Ecology-Progress Series* 31: 1-13.

Swartz, R. C., F. A. Cole, J. O. Lamberson, S. P. Ferraro, D. W. Schults, W. A. DeBen, H. Lee II, and R. J. Ozretich. 1994. Sediment toxicity, contamination and amphipod abundance at a DDT-and dieldrin-contaminated site in San Francisco Bay. *Envir. Toxicol. & Chem.* 13(6): 949-962.

U. S. EPA and U. S. ACOE. 1991. Evaluation of dredged material proposed for ocean disposal. Testing Manual. U. S. Environmental Protection Agency and Army Corps of Engineers. EPA-503/8-91-001. Washington, DC.

U.S. EPA 1986. Recommended protocols for conducting laboratory bioassays on Puget Sound sediments. 52 pp. Prepared for U.S. Environmental Protection Agency, Region 10, Puget Sound Estuary Program, Seattle, WA.

U.S. EPA 1990. Recommended protocols for conducting laboratory bioassays on Puget Sound sediments. Final Report. 79 pp. Prepared for U.S. Environmental Protection Agency, Region 10, Puget Sound Estuary Program, Seattle, WA.

U. S. EPA. 1994. Recommended guidelines for conducting laboratory bioassays on Puget Sound sediments. 81 pp. Prepared for U.S. Environmental Protection Agency, Region 10, Puget Sound Estuary Program, Seattle, WA.

U. S. EPA. 1994. Sediment quality criteria. *Federal Register* 59(11): 2652-2656.

Windom, H. L., S. J. Schropp, F. D. Calder, J. D. Ryan, R. G. Smith,Jr., L. C. Burney, F. G. Lewis, and C. H. Rawlinson. 1989. Natural trace metal concentrations in estuarine and coastal marine sediments of the Southeastern United States. *Envir. Sci. & Technol.* 23: 314-320.

Winger, P. V., P. J. Lasier, and H. Geitner. 1993. Toxicity of sediments and pore water from Brunswick Estuary, Georgia. *Arch. Environ. Contam. and Toxicol.* 25: 371-376.

Wise, S.A., B.A., Benner, G.D. Byrd, S.N. Chesler, R.E. Rebbert, M.M. Schantz. 1988. Determination of polycyclic aromatic hydrocarbons in a coal tar standard reference material. *Analytical Chemistry* 60:887, 894.

Wolfe, D. A. 1992. Selection of bioindicators of pollution for marine monitoring programmes. *Chemistry and Ecology* 6: 149-167.

Wolfe, D. A., E. R. Long, and A. Robertson. 1993. The NS&T bioeffects surveys: Design strategies and preliminary results. In: *Proceedings, Coastal Zone '93, the 8th Symposium on Coastal and Ocean Management* Vol. 1: 298-312. O. T. Magooon, W. S. Wilson, H. Converse and L. T. Tobin (editors). New York. American Society of Civil Engineers.

Wolfe, D. A., S. B. Bricker, E. R. Long, K. J. Scott, and G. B. Thursby. 1994. Biological effects of toxic contaminants in sediments from Long Island Sound and environs. NOAA Tech. Memo. NOS ORCA 80. National Oceanic and Atmospheric Administration. Silver Springs, MD.

List of Appendices

A1. Field notes from Charleston Harbor and vicinity	160
A2. Field notes from the upper Savannah River	163
A3. Field notes from the lower Savannah River	164
A4. Field notes from the St. Simons Sound	169
B1. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay	173
B2. Toxicity and chemistry data from upper and lower Savannah River sediment.....	226
B3. Toxicity and chemistry data from St. Simons Sound	272
C. Method detection limits (MDLs) for organics and trace metals	288

APPENDIX A1. Field notes from Charleston Harbor and vicinity.

Strata No.	Station No.	Location	Station	Date	Latitude	Longitude	Depth m
A -1	1	upper Leadenwah-Eastern Branch		5/1 3 /93	32°38.96 'N	80°11.17 'W	0.3
A -1	2	upper Leadenwah-Eastern Branch		5/1 3 /93	32°38.98 'N	80°10.95 'W	0.6
A -1	8	upper Leadenwah Creek off Maybank Highway by Treatment site)		5/1 4 /93	32°39.28 'N	80°10.40 'W	0.6
A -2	1	Leadenwah Creek, Middle Creek upper tidal zone below Spartina		5/1 3 /93	32°38.18 'N	80°12.66 'W	1.2
A -2	3	Eastern Leadenwah Creek-middle junction of Eastern and Mid Branch Leadenwah Creek		5/1 3 /93	32°37.82 'N	80°12.54 'W	0.6
A -2	7	lower Leadenwah by Eastern Bank		5/1 2 /93	32°38.93 'N	80°12.15 'W	0.6
A -3	1	lower Leadenwah Creek		5/1 3 /93	32°37.02 'N	80°13.52 'W	0.6
A -3	2	lower Leadenwah-Main Creek by community dock		5/1 3 /93	32°37.02 'N	80°13.88 'W	0.9
A -3	3	Winyah Bay; downstream of Steel Mill, upper Georgetown Harbor		5/1 3 /93	32°37.32 'N	80°13.25 'W	0.9
B -1	1	Winyah Bay; upper Georgetown Harbor off Front Street by Steel Mill		6/1 0 /93	32°22.01 'N	79°17.20 'W	3.6
B -1	2	Winyah Bay; upper Georgetown Harbor below Steel Mill		6/1 0 /93	32°21.88 'N	79°17.00 'W	2.3
B -1	3	Winyah Bay; upper Georgetown Harbor beside Paper Mill		6/1 0 /93	32°21.99 'N	79°17.11 'W	3.0
B -2	1	Winyah Bay; Turning Basin, downstream from paper mill		6/1 0 /93	32°17.53 'N	79°17.44 'W	6.4
B -3	1	Winyah Bay; Sampit Point Channel below Paper Mill and Steel Mill		6/1 1 /93	32°21.33 'N	79°16.89 'W	3.6
B -4	1	SE of Marina in Winyah Bay		6/1 1 /93	32°20.40 'N	79°16.57 'W	1.2
B -5	1	Winyah Bay; Upper Rabbit Island Channel below Paper Mill)		6/1 1 /93	33°19.84 'N	79°16.83 'W	3.6
B -6	1	Upper Rabbit Isle Channel-Winyah Bay		6/1 1 /93	33°20.40 'N	79°16.58 'W	4.5
B6	2	Lower Rabbit Isle Channel-Winyah Bay		6/1 1 /93	32°19.17 'N	79°16.96 'W	4.5
B -7	1	Charleston Harbor (Off Mt. Pleasant near sea buoy)		5/1 2 /93	32°46.23 'N	79°53.07 'W	9.7
C -1	1	Charleston Harbor (Off Ft. Sumter)		5/1 2 /93	32°45.33 'N	79°51.27 'W	13.0
C -2	1	Charleston Harbor	nd	nd	nd	nd	2.4
C -2	3	Charleston Harbor (west of Ft. Sumter at third range antenna)		5/1 2 /93	32°45.16 'N	79°52.87 'W	1.7
C -2	4	Charleston Harbor (1/4 mile east of Castle Pinckney)		5/1 2 /93	32°46.17 'N	79°54.58 'W	5.5
C -3	1	Charleston Harbor (1/2 mile east of Castle Pinckney)		5/1 2 /93	32°45.79 'N	79°54.21 'W	6.1
C -4	1	Charleston Harbor (Coast Guard Dock Ashley River 100 m. off)		5/1 2 /93	32°46.35 'N	79°56.70 'W	9.7
D -1	1	Charleston Harbor (City Marina Ashley River)		5/1 2 /93	32°46.64 'N	79°57.10 'W	9.7
D -1	2	Charleston Harbor		5/1 4 /93	32°46.79 'N	79°57.45 'W	9.7
D -1	3	Charleston Harbor (West of Highway 17, Ashley River)		5/1 4 /93	32°47.15 'N	79°57.85 'W	6.4
D -1	4	Charleston Harbor (East of Highway 17 bridge near Ashley Marina)		5/1 4 /93	32°46.86 'N	79°57.61 'W	6.7
D -2	1	Charleston Harbor (Ashley River)		5/1 3 /93	32°48.24 'N	79°58.33 'W	6.4

APPENDIX A1 continued.

D - 2	2	Charleston Harbor (Ashley River)	5/1 3/93	32°48.14 'N	79°58.27 'W	5.5
D - 2	3	Charleston Harbor (Ashley River across from The Citadel)	5/1 3/93	32°47.91 'N	79°58.37 'W	5.5
D - 3	1	Charleston Harbor (Across Ashley River from Koppers Creosote Plant)	5/1 3/93	32°49.30 'N	79°57.98 'W	2.1
D - 3	2	Charleston Harbor (Ashley River)	5/1 3/93	32°48.43 'N	79°58.20 'W	5.5
D - 3	3	Charleston Harbor (Ashley River)	5/1 3/93	32°48.43 'N	79°58.20 'W	5.5
D - 4	1	Charleston Harbor (Duck Island Reach - Ashley River)	5/1 3/93	32°50.17 'N	79°58.80 'W	4.6
D - 4	2	Charleston Harbor (Ashley River, Duck Island Reach)	5/1 3/93	32°49.51 'N	79°57.95 'W	6.4
D - 4	4	Charleston Harbor (Ashley River, Duck Island Reach)	5/1 3/93	32°49.88 'N	79°58.14 'W	5.8
E - 1	1	Charleston Harbor (1/4 mile SW of Patriots Point)	5/1 4/93	32°47.88 'N	79°54.81 'W	7.6
E - 1	2	Charleston Harbor (300 yards south of Patriots Point)	5/1 4/93	32°47.04 'N	79°54.54 'W	5.5
E - 1	3	Charleston Harbor (Horseroach Channel SW off Yorktown/Patriots Pt)	5/1 4/93	32°47.21 'N	79°54.83 'W	8.5
E - 2	1	Charleston Harbor (1/2 mi. west of Cooper River Bridge)	5/1 4/93	32°48.93 'N	79°54.81 'W	10.9
E - 2	2	Charleston Harbor (Daniel Island, SE tip of peninsula by red buoy)	5/1 4/93	32°48.94 'N	79°55.09 'W	13.9
E - 3	1	Charleston Harbor (1/2 miles west of Cooper River Bridge)	5/1 4/93	32°48.64 'N	79°55.80 'W	13.3
E - 3	2	Charleston Harbor (Town Creek Reach)	5/1 4/93	32°48.25 'N	79°55.83 'W	6.4
F - 1	1	Charleston Harbor (Lower Wando River)	6/1 0/93	32°49.33 'N	79°54.25 'W	9.1
F - 1	2	Charleston Harbor (Lower Wando River)	6/1 0/93	32°50.01 'N	79°53.68 'W	13.6
F - 1	3	Charleston Harbor (Lower Wando River)	6/1 0/93	32°50.81 'N	79°53.77 'W	5.5
F - 2	1	Charleston Harbor (Wando River, upstream of Mark Clark Bridge)	6/9/93	32°52.03 'N	79°53.35 'W	4.5
F - 2	2	Charleston Harbor (Wando River, upstream of Mark Clark Bridge)	6/9/93	32°51.62 'N	79°53.80 'W	7.9
F - 2	3	Charleston Harbor (Wando River)	6/9/93	nd	nd	5.5
F - 2	4	Charleston Harbor (Wando River, upstream of Mark Clark Bridge)	6/9/93	32°52.36 'N	79°52.13 'W	3.9
G - 1	1	Charleston Harbor (Main Channel, Shipyard Creek, Copper River)	6/9/93	32°49.80 'N	79°56.42 'W	11.2
G - 2	1	Charleston Harbor (Shipyard Creek off of a dry dock Cooper River)	6/9/93	32°50.04 'N	79°56.53 'W	11.5
H - 1	1	Charleston Harbor (Cooper River)	6/8/93	32°49.76 'N	79°55.80 'W	13.6
H - 1	1	Charleston Harbor (Myers Bend Reach, Cooper River)	6/9/93	32°49.79 'N	79°55.77 'W	13.6
H - 1	2	Charleston Harbor (Myers Beach Reach, Cooper River)	6/9/93	nd	nd	nd
H - 1	3	Charleston Harbor (Myers Beach Reach, Cooper River)	6/9/93	nd	nd	nd
H - 1	4	Charleston Harbor (Myers Beach Reach, Cooper River)	6/9/93	32°49.47 'N	79°55.70 'W	13.3
H - 1	5	Charleston Harbor (Myers Beach Reach, Cooper River)	6/9/93	32°49.75 'N	79°56.07 'W	13.6
H - 2	2	Charleston Harbor (Daniel Island Reach, Cooper River)	6/8/93	32°50.24 'N	79°55.94 'W	5.5
H - 2	3	Charleston Harbor (Daniel Island Reach, Cooper River)	6/8/93	32°50.38 'N	79°56.02 'W	3.6

APPENDIX A1 continued.

APPENDIX A1 continued.											
H - 2	6	Charleston Harbor (Daniel Island Reach, Cooper River)		6/ 8/ 93	32°50.84 'N	79°55.60 'W	4.5				
H - 3	1	Charleston Harbor (Clouter Creek, next to ship docks, Cooper River)		6/ 8/ 93	32°51.37 'N	79°56.92 'W	13.6				
H - 3	3	Charleston Harbor (Clouter Creek Reach, Cooper River)		6/ 8/ 93	32°51.122 'N	79°56.40 'W	12.7				
H - 3	5	Charleston Harbor (Clouter Creek Reach)		6/ 8/ 93	32°51.12 'N	79°56.60 'W	13.6				
H - 4	2	Charleston Harbor (Navy Yard Reach, Cooper River)		6/ 7/ 93	32°52.09 'N	79°57.63 'W	4.8				
H - 4	3	Charleston Harbor (Navy Yard Reach just off docks, Cooper River)		6/ 7/ 93	32°51.51 'N	79°57.33 'W	13.6				
H - 4	5	Charleston Harbor (Navy Yard Reach, middle of pier area, Cooper River)		6/ 7/ 93	32°51.88 'N	79°57.76 'W	11.8				
H - 5	1	Charleston Harbor (East of Mark Clark Bridge, Cooper River)		6/ 7/ 93	32°53.14 'N	79°57.70 'W	4.8				
H - 5	2	Charleston Harbor (SE of Hess Terminal, Cooper River)		6/ 7/ 93	32°52.32 'N	79°57.98 'W	4.2				
H - 5	3	Charleston Harbor (Cooper River)		6/ 7/ 93	nd	nd	13.6				
H - 5	4	Charleston Harbor (East channel near dredge, Cooper River)		6/ 7/ 93	32°52.97 'N	79°57.84 'W	6.1				
H - 5	5	Charleston Harbor (Cooper River)		6/ 7/ 93	nd	nd	nd				
H - 5	8	Charleston Harbor (opposite side of Noisette Creek, Cooper River)		6/ 7/ 93	32°52.41 'N	79°57.77 'W	5.5				
H - 6	1	Charleston Harbor (Filbin Creek Reach, Cooper River)		6/ 7/ 93	32°54.11 'N	79°57.37 'W	8.5				
H - 6	2	Charleston Harbor (Filbin Creek, Cooper River)		6/ 7/ 93	32°53.98 'N	79°57.58 'W	14.2				
H - 6	3	Charleston Harbor (Filbin Creek Reach, Cooper River)		6/ 7/ 93	32°53.75 'N	79°57.48 'W	5.5				
H - 7	1	Charleston Harbor (Ordinance Reach, Cooper River)		6/ 7/ 93	32°54.36 'N	79°57.09 'W	14.2				
H - 8	1	Charleston Harbor (Drydock reach, Cooper River)		6/ 7/ 93	32°55.05 'N	79°55.68 'W	13.0				
H - 5	6			6/ 7/ 93	nd	nd	nd				
H - 5	7			6/ 7/ 93	nd	nd	nd				
Reference	Ni-Oyster Landing	Charleston Harbor (Baruch-Oyster Landing)		6/10/93	nd	nd	0.3				
Reference	Ni-Oyster Landing	Charleston Harbor (North Inlet-Winyah-Oyster Landing by oyster cages)		5/14/93	nd	nd	0.3				
CHP-1	CHP-1	Charleston Harbor (Shipyard Creek block G-2, upper most end)		6/9/93	32°50.28 'N	79°56.80 'W	10.6				
CHP-2	2	Charleston Harbor (Romney Landfill)		5/14/93	32°48.23 'N	79°55.91 'W	7.9				
CHP	3 - Aquarium site	Charleston Harbor (Charleston Aquarium site)		5/14/93	32°47.35 'N	79°55.45 'W	3.3				
CHP-4	Plum Island Outfall	Charleston Harbor (Plum Island Outfall 200 ft NW of outfall)		5/12/93	32°46.06 'N	79°56.31 'W	7.3				
CHP-5	5	Charleston Harbor (Brittlebank Park, Ashley River)		5/13/93	32°47.17 'N	79°57.72 'W	6.4				
CHP-6	6	Charleston Harbor (Koopers Creasote Plant Ashley River, 150 m SE of p		5/13/93	32°49.14 'N	79°57.85 'W	9.7				
CHP-7	Dolphin Cove Mar.	Charleston Harbor (150 m west of Dolphin Cove Marina, Ashley River)		5/13/93	32°49.85 'N	79°57.98 'W	7.6				
CHP-8	Brickyard Creek	Charleston Harbor (Brickyard Creek-Ashley River)		5/13/93	32°49.99 'N	80°00.04 'W	2.7				
CHP-9	CHP-9	Charleston Harbor (mouth of Wallace Creek, base of Spartina)		6/8/93	nd	nd	0.0				
CHP-10	CHP-10	Charleston Harbor (Detyens Shipyard, 100 yards off Wando River)		6/9/93	32°55.67 'N	79°49.85 'W	13.6				
CHP-11	CHP-11	Charleston Harbor (Isle of Palms Connector)		6/11/93	nd	nd	1.2				

Appendix A2. Field notes from the upper Savannah River.

Strata No	Station No	Station	Location	Date	Time	Latitude	Longitude	Depth m
(US)A1	1	upper Savannah River		7/24/94	10:40 AM	31°12.86 'N	81°08.95 'W	1.5
(US)A1	2	upper Savannah River (south bank)		7/24/94	11:53 AM	32°12.54 'N	81°09.00 'W	2.5
(US)A1	3	upper Savannah River (south bank, Ursia Island)		7/24/94	12:50 PM	32°11.95 'N	81°09.32 'W	2.4
(US)A2	2	upper Savannah River		7/23/94	11:30 AM	32°11.12 'N	81°09.39 'W	3.3
(US)A2	3	upper Savannah River (West of Confluence of Steamboat River)		7/23/94	12:39 PM	32°10.77 'N	81°09.29 'W	nd
(US)A2	4	upper Savannah River		7/23/94	8:47 AM	32°11.31 'N	81°09.10 'W	nd
(US)B1	1	upper Savannah River (Port Wentworth turning basin)		7/25/94	12:10 PM	32°09.17 'N	81°08.96 'W	1.9
(US)B1	2	upper Savannah River (Port Wentworth channel)		7/25/94	9:39 AM	32°09.47 'N	81°09.22 'W	1.3
(US)B1	5	upper Savannah River (Port Wentworth channel)		7/25/94	11:00 AM	32°09.12 'N	81°08.76 'W	8.3
(US)B2	1	upper Savannah River (Whiteball channel)		7/21/94	15:02 PM	32°08.31 'N	81°08.51 'W	13.3
(US)B2	3	upper Savannah River		7/22/94	10:57 AM	32°08.41 'N	81°08.47 'W	13.6
(US)B2	4	upper Savannah River (Whiteball channel)		7/22/94	12:00 PM	32°08.65 'N	81°08.50 'W	10.9
(US)B3	3	upper Savannah River (South of Hutchinson Island, Kings Island Channel)		7/25/94	13:08 PM	32°06.93 'N	81°07.65 'W	14.8
(US)B3	5	upper Savannah River (South Kings Island channel, mouth of Dundee channel)		7/26/94	7:30 AM	32°06.96 'N	81°07.64 'W	1.3
(US)B3	6	upper Savannah River (Kings Island channel)		7/26/94	8:10 AM	32°06.57 'N	81°07.38 'W	13.3
(US)B4	4	upper Savannah River (Marsh Island channel east)		7/26/94	8:55 AM	32°05.70 'N	81°06.35 'W	13.9
(US)B4	5	upper Savannah River (Marsh Island channel east)		7/26/94	10:20 AM	32°06.24 'N	81°06.98 'W	14.2
(US)B4	6	upper Savannah River (between Kings and Marsh Islands in front of pulp mill)		7/26/94	11:10 AM	32°06.35 'N	81°07.19 'W	15.1
(US)B5	1	upper Savannah River (east of suspension bridge)		7/19/94	13:05 PM	32°05.20 'N	81°05.82 'W	1.2
(US)B5	2	upper Savannah River (N.W. of suspension bridge)		7/19/94	15:20 PM	32°05.42 'N	81°06.11 'W	14.2
(US)B5	3	upper Savannah River (S.W. of suspension bridge)		7/20/94	8:17 AM	32°05.58 'N	81°06.16 'W	14.5
(US)B6	1	upper Savannah River (front channel)		7/23/94	14:30 PM	32°04.79 'N	81°04.18 'W	12.1
(US)B6	2	upper Savannah River (front channel)		7/23/94	15:15 PM	32°04.85 'N	81°04.96 'W	13.6
(US)B6	3	upper Savannah River (front channel)		7/23/94	16:01 PM	32°04.74 'N	81°04.38 'W	nd
(US)B7	1	upper Savannah River (wrecks channel)		7/21/94	8:45 AM	32°04.80 'N	81°03.71 'W	17.6
(US)B7	2	upper Savannah River (wrecks channel)		7/24/94	16:24 PM	32°04.86 'N	81°02.88 'W	10.3
(US)B7	3	upper Savannah River (wrecks channel)		7/24/94	17:23 PM	32°04.82 'N	81°03.58 'W	13.3
(US)B8	1	upper Savannah River (Kings Island turning basin)		7/21/94	10:15 AM	32°08.00 'N	81°08.21 'W	12.1
(US)B8	2	upper Savannah River (Kings Island turning basin)		7/21/94	11:00 AM	nd	nd	nd
(US)B8	3	upper Savannah River (Kings Island turning basin)		7/22/94	14:12 PM	32°08.17 'N	81°08.42 'W	1.2
(US)B9	1	upper Savannah River (Finger canal, mouth of Dundee Canal)		7/20/94	11:50 AM	32°06.75 'N	81°07.38 'W	10.6
(US)B9	2	upper Savannah River (Finger canal, mouth of Dundee Canal)		7/19/94	10:05 AM	32°06.74 'N	81°07.78 'W	nd
(US)G	1	upper Savannah River (ocean terminal)		7/19/94	11:35 AM	32°05.51 'N	81°06.28 'W	4.6
(US)G	2	upper Savannah River		7/19/94	12:10 PM	32°05.50 'N	81°06.25 'W	2
(US)H	1	upper Savannah River (Stevens terminal)		7/19/94	9:07 AM	32°05.25 'N	81°05.37 'W	4.3
(US)H	2	upper Savannah River (Stevens terminal)		7/19/94	9:52 AM	32°05.02 'N	81°05.34 'W	5

Appendix A3. Field notes from the lower Savannah River.

Strata No.	Station No.	Station	Location	Date	Time	Latitude	Longitude	Depth m
(S)E1	1-1	Savannah River	(south channel)	6/25/94	16:15 PM	32°05.34 'N	81°00.80 'W	2.0
(S)E1	2-1	Savannah River	(south channel)	6/25/94	17:00 PM	32°04.76 'N	80°59.94 'W	1.6
(S)E1	3-1	Savannah River	(south channel)	6/25/94	18:02 PM	32°03.86 'N	80°58.15 'W	3.1
(S)C	1	Back River		6/26/94	11:30 AM	32°05.77 'N	81°04.23 'W	3.6
(S)C	2	Back River		6/26/94	12:15 PM	32°05.39 'N	81°03.59 'W	3.3
(S)C	3	Back River		6/26/94	13:04 PM	32°05.44 'N	81°04.07 'W	2.3
(S)E2	1	Savannah River	(south channel)	6/26/94	14:30 PM	32°03.74 'N	80°57.89 'W	0.9
(S)E2	2	Savannah River	(south channel)	6/26/94	15:23 PM	32°02.39 'N	80°56.44 'W	4.0
(S)E2	3	Savannah River	(south channel)	6/26/94	16:25 PM	32°01.88 'N	80°55.44 'W	3.7
(S)D1	1-1	Savannah River	(lower channel)	6/27/94	9:35 PM	32°04.89 'N	81°02.31 'W	8.4
(S)D1	2-2	Savannah River	(lower channel)	6/27/94	11:05 PM	32°05.37 'N	81°01.63 'W	6.5
(S)D1	3-1	Savannah River	(lower channel)	6/27/94	12:10 PM	32°05.91 'N	81°00.30 'W	2.9
(S)D2	3-1	Savannah River	(lower channel)	6/27/94	17:15 PM	32°03.72 'N	80°57.00 'W	4.0
(S)D2	2-2	Savannah River	(lower channel)	6/27/94	18:20 PM	32°04.59 'N	80°58.52 'W	5.4
(S)D2	1-1	Savannah River	(lower channel)	6/28/94	19:20 PM	32°04.48 'N	80°58.83 'W	3.7
(S)E3	2	Savannah River	(south channel)	6/28/94	10:05 AM	32°01.25 'N	80°53.74 'W	3.8
(S)E3	5	Savannah River	(south channel)	6/28/94	10:50 AM	32°01.57 'N	80°54.09 'W	4.8
(S)D3	1-4	Savannah River	(lower channel)	6/28/94	12:28 PM	32°03.09 'N	80°56.44 'W	2.9
(S)D3	2-1	Savannah River	(lower channel)	6/28/94	13:29 PM	32°02.51 'N	80°55.32 'W	2.5
(S)D3	3-2	Savannah River	(lower channel)	6/28/94	14:49 PM	32°02.12 'N	80°54.96 'W	3.5
(S)E3	9	Savannah River	(south channel)	6/28/94	15:50 PM	32°01.13 'N	80°53.18 'W	4.5

Appendix A3. Field notes from the lower Savannah River.

Strata	No.	Station No.	Air Temp. °C	Surface Temp. °C	Bottom Temp. °C	Surface ppt	Salinity ppt	Bottom Salinity ppt
(S)E1	1-1	n d	28.0	28.0	28.0	6.5	7.0	
(S)E1	2-1	n d	28.0	27.0	27.0	8.0	8.5	
(S)E1	3-1	n d	28.0	28.5	28.5	12.0	14.2	
(S)C	1	25.0	27.5	28.0	28.0	4.2	8.8	
(S)C	2	n d	28.0	28.3	28.3	3.9	6.6	
(S)C	3	n d	28.5	28.5	28.5	5.9	8.7	
(S)E2	1	n d	27.8	28.2	28.2	12.6	12.3	
(S)E2	2	n d	28.5	25.0	25.0	14.0	16.0	
(S)E2	3	n d	26.0	24.9	24.9	15.5	16.1	
(S)D1	1-1	26.5	27.9	28.0	28.0	5.0	14.0	
(S)D1	2-2	27.0	27.5	27.5	27.5	8.6	12.3	
(S)D1	3-1	27.8	27.9	27.4	27.4	11.0	15.0	
(S)D2	3-1	26.8	25.5	25.2	25.2	11.8	14.0	
(S)D2	2-2	22.5	27.0	27.5	27.5	8.5	10.7	
(S)D2	1-1	n d	26.0	25.1	25.1	7.3	7.6	
(S)E3	2	22.0	23.0	23.2	23.2	19.8	24.3	
(S)E3	5	n d	23.5	26.9	26.9	21.1	22.2	
(S)D3	1-4	n d	27.0	27.4	27.4	23.9	25.1	
(S)D3	2-1	n d	27.8	27.5	27.5	22.5	25.4	
(S)D3	3-2	27.5	28.0	27.5	27.5	26.0	26.0	
(S)E3	9	26.5	27.5	27.5	27.5	16.5	16.5	

Appendix A3. Field notes from the lower Savannah River.

Strata No.	Station No.	Surface D.O. mg / L	Bottom D.O. mg / L	Conductivity micro moles	Conductivity micro moles	Bottom moles
(S)E1	1-1	4.6	4.6	12,000	12,000	12,500
(S)E1	2-1	4.7	4.8	14,100	14,100	15,100
(S)E1	3-1	5.3	6.5	25,000	25,000	21,100
(S)C	1	5.1	3.5	7,900	7,900	6,100
(S)C	2	5.5	3.9	7,300	7,300	11,900
(S)C	3	4.8	3.75	10,800	10,800	15,900
(S)E2	1	5.1	5.1	21,800	21,800	21,600
(S)E2	2	5.3	4.85	24,500	24,500	26,200
(S)E2	3	5.3	5.1	25,800	25,800	25,800
(S)D1	1-1	4.5	3.8	8,400	8,400	25,000
(S)D1	2-2	4.9	4.6	15,400	15,400	21,880
(S)D1	3-1	4.8	4.7	19,000	19,000	26,000
(S)D2	3-1	5.4	5.2	19,200	19,200	23,300
(S)D2	2-2	5.0	4.5	15,300	15,300	18,500
(S)D2	1-1	4.7	4.85	12,500	12,500	13,000
(S)E3	2	6.2	5.6	30,500	30,500	37,000
(S)E3	5	5.9	5.5	32,700	32,700	36,500
(S)D3	1-4	5.5	5.5	49,700	49,700	42,000
(S)D3	2-1	6.1	5.4	38,200	38,200	42,300
(S)D3	3-2	6.15	6.4	43,500	43,500	43,500
(S)E3	9	6.45	6.5	28,500	28,500	29,000

Appendix A3. Field notes from the lower Savannah River.

Strata No.	Station No.	Sediment	Description
(S)E1	1-1	2 cm dark brown muddy sand over black clay, slight petroleum sheen.	
(S)E1	2-1	2 cm brown muddy sand overlying black clay layer.	
(S)E1	3-1	1st grab: muddy sand, shell debris; 2nd grab: sandy grey clay marl, lots of clay, mud crab; note: it took 6 grabs to get sample.	
(S)C	1	1/2 cm thin brown film over black silty clay, "silty mud", slight H2S odor; note: sampling site is next to spoil island.	
(S)C	2	1/2 cm light brown silty clay overlying black silty clay; note about sampling site: anchorage area, barges, impounded waters, dam; photos: roll E: 1-3.	
(S)C	3	Light brown medium to course sand; spoil area adjacent; boat anchorage; channel runs 45 ft. deep with "shoulders" starting approximately 15 ft.; photos: roll E: 4-6.	
(S)E2	1	Brown muddy sand, medium-course grain size, polychaete tubes, shell debris, spartina both sides, south of Bird Island; photos: roll E: 7-8.	
(S)E2	2	2 mm light brown sandy clay over grey clay (curling up the sides of grab), Highway 80 nearby, spartina both banks; photos: roll E: 9-10.	
(S)E2	3	1 cm light brown sandy clay with some shell debris over black sandy clay, no odor, spartina marsh on both sides, Highway 80 to the south; photos: roll E: 12-13.	
(S)D1	1-1	2 cm soft olivine silt over black silty clay with some gravel, slight petroleum sheen; 3rd grab: sandy clay over black; 4th grab: same as 3rd; station at confluence of Back River and Savannah River, ship traffic, petroleum factory, spartina marsh to the south, it took 9 drops to get sample; photos: roll E: 14-20.	
(S)D1	2-2	Thin layer of brown coarse sand (sand appears to be in furrows) over firm grey clay marl, spoil island to northwest, refinery to south; photos: roll E: 24-26.	
(S)D1	3-1	Light brown-olivine soft sandy-silty clay, petroleum sheen, gravel in grab, spoil islands on both sides, note: station moved closer to shoreline, have to get river stations on slack tides, Elba Island is a spoil island; photos: roll F: 1-3.	
(S)D2	3-1	Brown muddy sand wi some clay, shell hash, clam (mulinia); 2nd grab: more clay, forest/marsh on both banks, S+E of Intracoastal; photos: roll F: 4-6.	
(S)D2	2-2	1 mm brown sand layer over grey clay, some shell hash; 2nd grab: 5 mm brown sand over grey clay; 3rd grab: 4-5 mm sand over grey clay; went to station 2-2 because slope of channel was too steep; photos: roll F: 7-10.	

Appendix A3. Field notes from the lower Savannah River.			
Strata No.	Station No.	Sediment	Description
(S)D2	1-1	Grey clay marl on surface, thin layer brown sand over grey clay; 2nd grab: marl/slate;	numerous drops - moved station from 45' to 15' along shore; photos: roll F: 12-13.
(S)E3	2	Brown sand with some clay, silt mixed in with sand, shell hash; 2nd grab: more sand component with clay inclusions; spartina both sides; bridge to west; photos: roll F: 14-18.	Olivine/brown silt > 1 mm over dark charcoal clay (pluff mud), shell hash on top, some wood chips, fecal pellets on surface, worm tubes, polychaete burrows; spartina on both sides, bridge to east; photos: roll F: 19-21.
(S)D3	1-4	2 mm brown silt over sand, highly bedded, clay inclusions, worms poly or oligochaetes; 1-1: shells, coarse gravel, coarse sand - rejected; 1-2: same; 1-3: same local; photos: roll F: 22-24.	Thin brown silt layer over coarse sand, shell and rock with clay inclusions below "muddy sand", vegetation on both banks, sheen/specks visible when homogenizing; photos: roll G: 5-8.
(S)D3	2-1	Thin brown silt over dark charcoal silt, no noticeable odor, some plant debris, gray clay inclusions, worm tubes and hogchoker observed, vegetation on both banks; 3-1: dredged - rock only; photos: roll G: 9-13.	Pluff mud with thin brown silt layer on top, some worm tubes, some plant material, lots of biological activity, vegetation on both banks; photos: roll G: 15-19.
(S)E3	9		

APPENDIX A4. Field notes from St. Simons Sound.

Strata	No.	Station No.	Station Location	Date	Time	Latitude	Longitude	Depth m
(B)A1	1	Turtle River	6/22/94	9:11 AM	31°13.08 'N	81°33.64 'W	8.5	
(B)A2	2	Turtle River	6/22/94	10:41 AM	31°12.00 'N	81°33.26 'W	5.0	
(B)A3	1	Turtle River	6/22/94	11:46 AM	31°11.66 'N	81°32.21 'W	3.0	
(B)P	1	Purvis Creek	6/22/94	12:40 PM	31°11.19 'N	81°31.02 'W	7.0	
(B)D1	1	Turtle River	6/22/94	15:38 PM	31°10.25 'N	81°31.60 'W	7.5	
(B)D2	1	Turtle River	6/22/94	16:34 PM	31°08.90 'N	81°31.87 'W	6.2	
(B)E2	2	St. Simons Sound	6/23/94	8:40 AM	31°05.76 'N	81°26.66 'W	7.0	
(B)E1	1	St. Simons Sound	6/23/94	9:30 AM	31°06.83 'N	81°29.41 'W	3.5	
(B)D3	2	Turtle River	6/23/94	11:10 AM	31°07.08 'N	81°30.25 'W	3.5	
(B)B1	1	Academy Creek	6/23/94	12:30 PM	31°09.48 'N	81°29.97 'W	5.0	
(B)B2	1	Academy Creek	6/23/94	13:05 PM	31°09.14 'N	81°29.98 'W	4.5	
(B)C1	1	East River	6/23/94	13:47 PM	31°08.83 'N	81°29.94 'W	8.5	
(B)C2	1	East River	6/23/94	14:26 PM	31°07.90 'N	81°29.69 'W	6.5	
(B)C3	1	East River	6/23/94	15:27 PM	31°07.81 'N	81°29.66 'W	8.0	
(B)E3	1	St. Simons Sound	6/24/94	9:00 AM	31°06.97 'N	81°25.80 'W	3.0	
(B)G3	1	St. Simons Sound	6/24/94	9:55 AM	31°08.11 'N	81°25.18 'W	6.8	
(B)G2	1	St. Simons Sound	6/24/94	11:10 AM	31°08.11 'N	81°25.18 'W	5.1	
(B)G1	1	Back River	6/24/94	12:10 PM	31°08.96 'N	81°26.66 'W	6.5	
(B)F	1-1	Terry Creek	6/24/94	13:00 PM	31°09.90 'N	81°27.64 'W	3.5	
(B)F	2-1	Dupree Creek	6/24/94	13:40 PM	31°09.96 'N	81°28.28 'W	1.1	

APPENDIX A4. Field notes from St. Simons Sound.

Strata	No.	Station	No.	Air Temp.	Surface Temp.	Bottom Temp.	°C	°C	Surface Salinity	Bottom Salinity	pp t	pp t
(B)A		1-1		25.5	28.5		28.5		24.5		23.0	
(B)A		2-2		29.0	29.0		29.0		22.0		22.0	
(B)A		3-1		28.0	29.5		29.0		21.5		21.5	
(B)P		1		31.0	30.5		29.5		20.5		20.5	
(B)D		1-1		31.0	30.0		29.5		20.5		21.5	
(B)D		2-1		nd	29.0		26.0		23.5		25.0	
(B)E		2-2		nd	29.5		29.5		28.5		28.5	
(B)E		1-1		nd	29.5		29.0		26.5		26.5	
(B)D		3-2		nd	29.0		29.0		25.5		25.5	
(B)B		1-1		30.5	30.5		29.5		25.0		25.5	
(B)B		2-1		30.0	30.0		27.5		25.0		27.0	
(B)C		1-1		30.0	29.5		29.0		26.0		28.5	
(B)C		2-1		30.0	30.0		29.0		25.0		26.0	
(B)C		3-1		30.5	30.0		29.0		25.0		23.5	
(B)E		3-1		nd	32.0		31.5		28.5		28.5	
(B)G		3-1		nd	30.5		31.0		29.5		29.5	
(B)G		2-1		nd	29.0		29.5		30.0		29.5	
(B)G		1-1		nd	30.0		30.0		27.0		24.5	
(B)F		1-1		nd	31.0		30.0		21.0		24.3	
(B)F		2-1		nd	32.0		30.0		17.2		21.5	

APPENDIX A4. Field notes from St. Simons Sound.

Strata No.	Station No.	Surface D.O. mg/L	Bottom D.O. mg/L	Conductivity		Conductivity	
				micro	moles	micro	moles
(B)A	1-1	2.8	2.25	38,000	38,000	38,000	38,000
(B)A	2-2	2.6	2.55	38,000	38,000	38,000	38,000
(B)A	3-1	27.0	2.6	37,000	37,000	37,000	37,000
(B)P	1	3.55	1.9	36,500	36,500	36,500	36,500
(B)D	1-1	3.65	2.95	37,000	37,000	37,000	37,000
(B)D	2-1	2.8	2.7	40,500	40,500	40,500	40,500
(B)E	2-2	6.1	5.9	48,500	48,500	48,500	48,500
(B)E	1-1	4.6	4.4	46,000	46,000	45,000	45,000
(B)D	3-2	4.2	4.2	44,500	44,500	44,500	44,500
(B)B	1-1	12.5	0.4	44,000	44,000	44,000	44,000
(B)B	2-1	8.5	0.9	43,500	43,500	44,000	44,000
(B)C	1-1	6.8	3.1	43,500	43,500	46,500	46,500
(B)C	2-1	5.3	3.7	43,500	43,500	43,500	43,500
(B)C	3-1	5.6	3.0	44,000	44,000	39,500	39,500
(B)E	3-1	5.2	5.2	49,500	49,500	49,500	49,500
(B)G	3-1	5.6	5.4	>50,000	>50,000	>50,000	>50,000
(B)G	2-1	5.4	5.2	49,900	49,900	>50,000	>50,000
(B)G	1-1	4.8	5.0	47,500	47,500	42,500	42,500
(B)F	1-1	4.4	3.6	38,100	38,100	42,800	42,800
(B)F	2-1	4.8	4.1	31,500	31,500	36,500	36,500

APPENDIX A4. Field notes from St. Simons Sound.

Strata No.	Station No.	Sediment	Description
(B)A	1-1	Gravely-shell hash-dark gray mud, crustaceans, bivalves, tubeworms, amphipods, lots of small mussels, "healthy substrate"; photos: roll A: 2-3.	
(B)A	2-2	Light brown sandy silt overlying a grey clay; A2-1 two grabs were attempted - both were wash outs; In A2-2 we had to move the site out of the channel which had been dredged, closer to shore to obtain undredged material; photos: roll A: 4-8.	
(B)A	3-1	1st grab: 1 cm light brown sandy mud overlying black anoxic sandy mud, H2S odor, plant debris (spartina); 2nd grab: lots of wood debris; 3rd grab: similar to 1st grab; photos: roll A: 9-16.	
(B)P	1	1st grab: clay marl; 2nd grab: light tan muddy sand; 3rd grab: clay balls; 4th grab: clay marl and sand, plant debris, there is a sheen in sample; noted: there is a visible sheen on the exposed intertidal area; note: visible oysters at site; wood storks (endangered) and loggerhead sea turtle at station; Coastguard: LORAN is down; lengthy problems with winch at site - 2.5 hours to find/fix problem; photos: roll A: 17-19.	
(B)D	1-1	Brown muddy sand with black muddy sand inclusions.	
(B)D	2-1	Muddy sand, dendritic sponge, callinects crab, lots of shell hash, amphipods, quahog clam, fish, box crab, shrimp; note: it took 12 drops to get this sample! photos: roll A: 24, roll B: 1-3.	
(B)E	2-2	Slightly sandy silty clay, 1 cm olivine over dark grey layer, some shell hash; note: E 2-1 contained lots of shell hash with little mud - rejected; LORAN still down; photos: roll B: 4-7.	
(B)E	1-1	1st grab: clay, marly clay; 2nd grab: silty clay, oyster shell, H2S odor; 3rd grab: fine olivine silty clay over deep black silty clay, shell debris.	
(B)D	3-2	Muddy sand with some clay, 1 cm light brown overlying dark grey-black layer; LORAN still down; photos: roll B: 11-16.	
(B)G	1-1	2 cm brown runny "pluff" mud overlying solid jet-black gooey clay, organically enriched, note: a tad bit fresher on bottom; photos: roll D: 7-10 (prints).	
(B)F	1-1	2 cm brown "pluff" mud overlying jet-black clay; photos: roll D: 11-14 (prints).	
(B)F	2-1	Very thin brown floc layer overlying black anoxic silty mud containing clay balls, woody debris and plant debris, VERY STRONG petroleum/solvent odor, organic sheen in sample; last grab: sheen and turpentine odor; note: oysters at site.	

Appendix B1. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Percent amphipod survival	Amph Survival as percent of control	Significance (% of sample)	Microtox EC50 (mg/ml)	Microtox EC50 %Ref.(177 & 227)^	Significance
93-146	C1-1	Ashley River	86	101.18	ns	101.00	9.80	519.25
93-147	C2-4	Ashley River	84	98.82	ns	30.27	2.92	155.60
93-148	C3-1	Ashley River	86	101.18	ns	90.97	8.46	467.71
93-149	C4-1	Ashley River	93	109.41	ns	101.00	9.43	519.25
93-150	CHP4	Plum Isl. Outfall	83	97.65	ns	12.45	1.16	64.02
93-151	C5-1	Ashley River	87	102.35	ns	62.56	5.82	321.63
93-152	D1-1	Ashley River	91	107.06	ns	6.63	0.63	34.10
93-153	A2-3	Leadenwah Creek	87	102.35	ns	29.94	2.69	153.91
93-154	A3-1	Leadenwah Creek	86	101.18	ns	11.75	1.10	60.39
93-155	A3-3	Leadenwah Creek	86	101.18	ns	32.93	3.12	169.30
93-156	A2-7	Leadenwah Creek	88	103.53	ns	19.76	1.39	101.57
93-157	A2-1	Leadenwah Creek	87	102.35	ns	21.92	1.58	112.69
93-158	A1-1	Leadenwah Creek	90	105.88	ns	0.20	0.01	1.05
93-159	A1-2	Leadenwah Creek	91	107.06	ns	65.37	4.17	336.08
93-160	CHP5	Brittle Bank Park	85	100.00	ns	56.19	3.87	288.86
93-161	CHP8	Brickyard Creek	87	102.35	ns	40.34	2.58	207.37
93-162	D4-1	Ashley River	84	98.82	ns	101.00	6.79	519.25
93-163	D4-2	Ashley River	89	104.71	ns	17.45	1.16	89.70
93-164	D4-4	Ashley River	87	102.35	ns	8.91	0.65	45.79
93-165	D2-1	Ashley River	88	103.53	ns	28.99	2.03	149.04
93-166	D2-3	Ashley River	77	90.59	ns	34.94	2.31	179.63
93-167	D2-2	Ashley River	90	105.88	ns	8.94	0.58	45.97
93-168	CHP6	Koppers	82	96.47	ns	71.13	4.91	365.69
93-169	D3-3	Ashley River	88	103.53	ns	36.94	2.71	189.92
93-170	D3-1	Ashley River	81	95.29	ns	0.01	0.0004	0.04
93-171	CHP7	Dolphin Cv Marina	85	100.00	ns	0.33	0.03	1.72
93-172	D3-2	Ashley River	91	107.06	ns	7.62	0.49	39.18
93-173	E1-1	Cooper River	81	95.29	ns	47.50	3.28	244.20
93-174	E2-2	Cooper River	84	98.82	ns	11.32	0.81	58.21
93-175	E2-1	Cooper River	85	100.00	ns	0.91	0/06	4.69
93-176	A1-8	Leadenwah Creek	82	96.47	ns	20.48	1.42	105.29
93-177	Ref. No.	North Inlet	90	105.88	ns	19.45	1.46	100.00
93-178	D1-3	Ashley River	85	100.00	ns	40.94	2.93	210.46
93-179	D1-4	Ashley River	79	92.94	ns	101.00	7.45	519.25
93-180	CHP3	Aquarium Site	84	98.82	ns	0.33	0.02	1.70
93-181	E1-3	Cooper River	90	105.88	ns	101.00	7.49	519.25
93-182	E1-2	Cooper River	85	100.00	ns	92.27	6.71	474.36
93-183	E3-1	Cooper River	90	105.88	ns	101.00	7.24	519.25
93-184	E3-3	Cooper River	81	95.29	ns	12.66	0.89	65.10
93-185	CHP2	Romney St.	90	105.88	ns	101.00	7.48	519.25
93-186	A3-2	Leadenwah Creek	87	102.35	ns	101.00	6.60	519.25
93-196	H4-2	Cooper River	91	103.41	ns	44.69	3.03	101.72

Appendix B1. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Percent amphipod survival	Amph Survival as percent of control	Significance (% of sample)	Microtox EC50 (mg/ml)	Microtox EC50 %Ref.(177 & 227) ^a	Significance
93-197	H4-3	Cooper River	91	103.41	ns	101.00	7.22	229.90
93-198	H4-5	Cooper River	94	106.82	ns	60.72	4.14	138.22
93-199	H5-2	Cooper River	93	105.68	ns	59.03	3.84	134.37
93-200	H5-4	Cooper River	90	102.27	ns	3.42	0.24	*
93-201	H5-8	Cooper River	96	109.09	ns	20.74	1.58	47.21
93-202	H6-1	Cooper River	92	104.55	ns	6.10	0.45	*
93-203	H6-2	Cooper River	88	100.00	ns	4.27	0.31	13.89
93-204	H6-3	Cooper River	93	105.68	ns	101.00	7.28	*
93-205	H7-1	Cooper River	88	100.00	ns	3.06	0.22	229.90
93-206	H8-1	Cooper River	93	105.68	ns	4.68	0.33	6.97
93-207	H2-2	Cooper River	86	97.73	ns	101.00	7.55	**
93-208	H2-3	Cooper River	88	100.00	ns	8.37	0.62	10.65
93-209	H2-6	Cooper River	86	97.73	ns	2.23	0.17	*
93-210	CHP9	Wallace Channel	91	103.41	ns	14.45	1.01	5.08
93-211	H3-1	Cooper River	91	103.41	ns	101.00	7.12	32.90
93-212	H3-3	Cooper River	89	101.14	ns	6.30	0.47	*
93-213	H3-5	Cooper River	93	105.68	ns	12.73	0.89	229.90
93-214	F1-1	Wando River	93	105.68	ns	71.30	5.29	14.33
93-215	F1-2	Wando River	97.5	110.80	ns	2.63	0.18	28.99
93-216	F1-3	Wando River	93	105.68	ns	101.00	7.29	*
93-217	H1-1	Cooper River	92	104.55	ns	0.52	0.04	1.18
93-218	H1-4	Cooper River	87	98.86	ns	80.18	5.80	182.51
93-219	H1-5	Cooper River	86	97.73	ns	12.08	0.83	*
93-220	F2-1	Wando River	87	98.86	ns	61.16	4.38	27.50
93-221	F2-2	Wando River	90	102.27	ns	101.00	7.53	139.21
93-222	F2-4	Wando River	91	103.41	ns	53.71	3.63	229.90
93-223	G1-1	Cooper River	91	103.41	ns	4.10	0.28	122.27
93-224	G2-1	Cooper River	91	103.41	ns	11.37	0.78	9.33
93-225	CHP1	Shipyard Creek	91	103.41	ns	50.39	3.76	25.89
								114.71

Appendix B1. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Percent amphipod survival	Amph Survival as percent of control	Significance (% of sample)	Microtox EC50 (mg/ml)	Microtox EC50 %Ref.(177 & 227) [^]	Significance
93-226	CHP10	Detyen's Shipyard	95	107.95	ns	0.03	0.002	0.06
93-227	Ref. No.	North Inlet	84	95.45	ns	43.93	3.01	100.00
93-228	B1-1	Winyah Bay	92	104.55	ns	4.06	0.29	9.25
93-229	B3-1	Winyah Bay	93	105.68	ns	101.00	7.14	229.90
93-230	B1-3	Winyah Bay	84	95.45	ns	0.11	0.01	0.25
93-231	B1-2	Winyah Bay	94	106.82	ns	0.00	0.00001	0.00
93-232	CHP11	I of P Connector	88	100.00	ns	45.50	3.45	103.57
93-233	B7-1	Winyah Bay	92	104.55	ns	6.31	0.49	14.37
93-234	B6-2	Winyah Bay	93	105.68	ns	101.00	7.03	229.90
93-235	B5-1	Winyah Bay	88	100.00	ns	0.73	0.05	1.66
93-236	B4-1	Winyah Bay	94	106.82	ns	0.00	0.0001	0.00
93-237	B2-1	Winyah Bay	91	103.41	ns	101.00	7.66	229.90
brez	ref1		85					ns
brez	ref1		88					
star	ref2		89					
star	ref2		88					
USC lab ctrl batch 2								
USC lab ctrl batch 3								
NIOL Ref 1 batch 1								
NIOL Ref 1 batch 2								
NIOL Ref 1 batch 3								
ns	non-significant							
[^]	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.						
+	statistically significant for Dunnett's t-test at 0.05							
++	statistically significant for Dunnett's t-test at 0.01							
*	statistically significant for Mann-Whitney and Dunnett's							
**	statistically significant for Mann-Whitney, Dunnets, and Distribution-free							
***	statistically significant for Mann-Whitney, Dunnets, and Distribution-free							
Duplicates		NAPHTHALENE /ETHYLNAPHTHALENE /ETHYLNAPHTHALENE						
		GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	NAPHTHYL
93-165	D2-1	Ashley River	39.5	25.95	11.3	13.40	7.84	25.9
93-165	D2-1	Ashley River	53.9	67.43	12.9	21.35	23.88	87.4
93-166	D2-3	Ashley River	28.6	16.48	8.0	7.71	5.21	38.3
93-166	D2-3	Ashley River	26.3	16.77	9.0	7.94	4.27	53.8
93-167	D2-2	Ashley River	69.6	36.88	15.4	16.50	9.50	57.4
93-167	D2-2	Ashley River	598.2	159.64	86.3	99.60	47.70	149.9

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	% urchdev in 50% porewater	Significance in 25% porewater	% urchdev in 25% porewater	Significance	UAN, ug/l amphipod start	UAN, ug/l amphipod end	UAN, ug/l amph ave.	Pwater UAN ug/l
93-146	C1-1	Ashley River				17.84	0.81	9.32	111.27	
93-147	C2-4	Ashley River				35.70	266.12	150.91	38.12	
93-148	C3-1	Ashley River				16.71	2.59	9.65	58.88	
93-149	C4-1	Ashley River				7.41	1.31	4.36	33.81	
93-150	CHP4	Plum Isl. Outfall				7.39	1.01	4.20	65.73	
93-151	C5-1	Ashley River				14.42	1.04	7.73	52.93	
93-152	D1-1	Ashley River				9.88	0.59	5.24	49.44	
93-153	A2-3	Leadenwah Creek				3.80	15.35	9.58	4.66	
93-154	A3-1	Leadenwah Creek				0.42	25.43	12.93	1.76	
93-155	A3-3	Leadenwah Creek				2.54	1.37	1.95	9.86	
93-156	A2-7	Leadenwah Creek				1.35	1.68	1.52	15.80	
93-157	A2-1	Leadenwah Creek				0.44	3.84	2.14	5.72	
93-158	A1-1	Leadenwah Creek				0.40	0.81	0.60	10.77	
93-159	A1-2	Leadenwah Creek				1.76	1.34	1.55	23.79	
93-160	CHP5	Brittle Bank Park				6.81	1.25	4.03	65.45	
93-161	CHP8	Brickyard Creek				6.10	0.46	3.28	71.69	
93-162	D4-1	Ashley River				21.20	0.77	10.99	106.94	
93-163	D4-2	Ashley River				13.56	0.59	7.08	21.20	
93-164	D4-4	Ashley River				2.59	0.51	1.55	23.60	
93-165	D2-1	Ashley River				29.01	0.60	14.80	29.48	
93-166	D2-3	Ashley River				7.69	0.59	4.14	17.57	
93-167	D2-2	Ashley River				4.89	0.56	2.72	13.72	
93-168	CHP6	Koppers				9.41	0.54	4.97	47.85	
93-169	D3-3	Ashley River				24.93	0.56	12.75	22.80	
93-170	D3-1	Ashley River				10.26	0.53	5.39	47.95	
93-171	CHP7	Dolphin Cv Marina				24.07	0.59	12.33	24.62	
93-172	D3-2	Ashley River				20.62	0.70	10.66	41.56	
93-173	E1-1	Cooper River				11.42	0.63	6.02	39.68	
93-174	E2-2	Cooper River				6.60	0.52	3.56	52.79	
93-175	E2-1	Cooper River				8.41	0.54	4.48	29.32	
93-176	A1-8	Leadenwah Creek				14.94	0.66	7.80	19.04	
93-177	Ref. No.	North Inlet				2.94	0.68	1.81	1.19	
93-178	D1-3	Ashley River				8.35	0.58	4.47	34.47	
93-179	D1-4	Ashley River				11.37	0.59	5.98	44.16	
93-180	CHP3	Aquarium Site				18.15	0.55	9.35	32.03	
93-181	E1-3	Cooper River				18.15	0.61	9.38	37.74	
93-182	E1-2	Cooper River				2.61	0.59	1.60	35.76	
93-183	E3-1	Cooper River				12.20	0.47	6.34	53.38	
93-184	E3-3	Cooper River				9.85	0.53	5.19	28.80	
93-185	CHP2	Romney St.				41.31	0.63	20.97	106.14	
93-186	A3-2	Leadenwah Creek				5.87	0.55	3.21	10.51	
93-196	H4-2	Cooper River				1.50	0.39	0.95	26.55	

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	% urch fert. in 100% porewater	Significance	% urch fert. in 50% porewater	Significance	% urch fert. in 25% porewater	Significance	% urchdevt. in 100% porewater	Significance
93-97	H4-3	Cooper River	94	ns	95.0	ns	94.8	ns	ns	ns
93-98	H4-5	Cooper River	91	+	96.2	ns	96.6	ns	ns	ns
93-99	H5-2	Cooper River	92.8	ns	97.0	ns	92.6	++		
93-200	H5-4	Cooper River	90.2	+	95.2	ns	94.4	ns		
93-201	H5-8	Cooper River	96.6	ns	95.8	ns	97.6	ns		
93-202	H6-1	Cooper River	87.6	++	96.8	ns	93.8	+		
93-203	H6-2	Cooper River	84.8	++	96.4	ns	98.0	ns		
93-204	H6-3	Cooper River	96.2	ns	97.6	ns	96.4	ns		
93-205	H7-1	Cooper River	97	ns	99.0	ns	98.6	ns		
93-206	H8-1	Cooper River	85.2	++	97.4	ns	98.4	ns		
93-207	H2-2	Cooper River	91.8	ns	96.6	ns	95.2	ns		
93-208	H2-3	Cooper River	98	ns	98.4	ns	98.8	ns		
93-209	H2-6	Cooper River	98.2	ns	98.0	ns	96.0	ns		
93-210	CHP9	Wallace Channel	91.4	+	95.4	ns	97.2	ns		
93-211	H3-1	Cooper River	97.4	ns	98.2	ns	98.8	ns		
93-212	H3-3	Cooper River	94.4	ns	97.8	ns	97.6	ns		
93-213	H3-5	Cooper River	82.8	++	97.2	ns	98.0	ns		
93-214	F1-1	Wando River	82.2	++	93.4	ns	86.8	++		
93-215	F1-2	Wando River	88.4	++	96.4	ns	95.0	ns		
93-216	F1-3	Wando River	94.4	ns	95.6	ns	94.8	ns		
93-217	H1-1	Cooper River	19.4	++	80.4	++	83.8	++		
93-218	H1-4	Cooper River	78.2	++	93.2	ns	94.4	++		
93-219	H1-5	Cooper River	70.0	++	92.4	+	95.6	ns		
93-220	F2-1	Wando River	90.6	+	92.0	ns	84.6	++		
93-221	F2-2	Wando River	87.2	++	95.0	ns	93.2	++		
93-222	F2-4	Wando River	94.0	ns	97.4	ns	94.8	ns		
93-223	G1-1	Cooper River	58.8	++	94.0	ns	93.6	+		
93-224	G2-1	Cooper River	79.6	++	91.4	+	94.2	ns		
93-225	CHP1	Shipyard Creek	68.8	++	91.2	++	94.6	ns		

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	% urch fert. in 100% porewater	Significance	% urch fert. in 50% porewater	Significance	% urch fert. in 25% porewater	Significance	% urchdevt. in 100% porewater	Significance
93-226	CHP10	Detyen's	89.2	++	95.2	ns	93.4	+		
93-227	Ref. No.	North Inlet	97.6	ns	97.2	ns	93.2	+		
93-228	B1-1	Winyah Bay	88.4	++	96.2	ns	95.8			
93-229	B3-1	Winyah Bay	98.4	ns	98.4	ns	97.4	ns		
93-230	B1-3	Winyah Bay	74.8	++	95.4	ns	92.8	+		
93-231	B1-2	Winyah Bay	50.2	++	89.0	++	96.4	ns		
93-232	CHP11	I of P Connector	87.6	++	91.6	++	84.3	++		
93-233	B7-1	Winyah Bay	89.8	++	93.4	ns	86.6	++		
93-234	B6-2	Winyah Bay	88.4	++	96.6	ns	95.2	ns		
93-235	B5-1	Winyah Bay	5	++	57.3	++	56.6	++		
93-236	B4-1	Winyah Bay	2.8	++	23.0	++	63.6	++		
93-237	B2-1	Winyah Bay	86	++	95.8	ns	97.0	ns		
brez	ref1		97.6		98.2		98.8			
brez	ref1									
slar	ref2		97.4		99.2		98.8			
slar	ref2									
USC lab ct batch 2										
USC lab ct batch 3										
NIOL Ref 1 batch 1										
NIOL Ref 1 batch 2										
NIOL Ref 1 batch 3										
ns	non-significant									
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.								
+		statistically significant for Dunnell's t-test at 0.05								
++		statistically significant for Dunnell's t-test at 0.01								
*		statistically significant for Mann-Whitney								
**		statistically significant for Mann-Whitney and Dunnett's								
***		statistically significant for Mann-Whitney, Dunnett's, and Distribution-free								
Duplicates										
93-165	D2-1	Ashley River	3,5-trime nap GC, ug/kg 42.6	GC, ug/kg 3.9	FLUORENE ANTHRACENE GC, ug/kg 59.5	FLUORANTHENE GC, ug/kg 273.4	THIOPHENANTHRENACENE GC, ug/kg 44.8	FLUORANTHENE GC, ug/kg 2705.2	PYRENE GC, ug/kg 1195.2	
93-165	D2-1	Ashley River	32.0	<1	152.6	554.3	34.8	1156.1	895.7	758.3
93-166	D2-3	Ashley River	38.9	2.9	22.1	139.3	28.9	294.3	459.3	431.0
93-166	D2-3	Ashley River	44.1	<1	24.6	155.7	10.4	365.3	536.0	607.3
93-167	D2-2	Ashley River	55.5	4.4	35.7	163.9	29.2	665.1	1021.2	961.4
93-167	D2-2	Ashley River	135	18.4	82.4	289.7	90.7	289	666.7	1011.7

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	% urch fert. in 100% porewater	Significance	% urch fert. in 50% porewater	Significance	% urch fert. in 25% porewater	Significance	% urchdevpt. in 100% porewater	Significance
93-146	C1-1	Ashley River	96.2	ns	97.8	ns	92.8	ns	ns	
93-147	C2-4	Ashley River	92.6	ns	95.6	ns	84.4	++		
93-148	C3-1	Ashley River	35.8	++	74.4	++	56.6			
93-149	C4-1	Ashley River	41	++	63.2	++	67.4	++		
93-150	CHP4	Plum Isl. Outfall	66.6	++	71.4	++	68.8	++		
93-151	C5-1	Ashley River	31	++	75.4	++	75.2	++		
93-152	D1-1	Ashley River	98.2	ns	99.4	ns	97.8	ns		
93-153	A2-3	Leadenwah	99.6	ns	99.6	ns	98.8	ns		
93-154	A3-1	Leadenwah Creek	97.8	ns	99.6	ns	99.8	ns		
93-155	A3-3	Leadenwah Creek	96	ns	99.4	ns	99.2	ns		
93-156	A2-7	Leadenwah Creek	99.2	ns	99	ns	99.4	ns		
93-157	A2-1	Leadenwah Creek	99.4	ns	100	ns	99.6	ns		
93-158	A1-1	Leadenwah	96	ns	97	ns	87.8	++		
93-159	A1-2	Leadenwah Creek	97.4	ns	97.8	ns	92.2	+		
93-160	CHP5	Brittle Bank Park	87.6	++	92.4	++	91	++		
93-161	CHP8	Brickyard Creek	92.2	ns	93	++	90.8	++		
93-162	D4-1	Ashley River	94.2	ns	98	ns	95.4	ns		
93-163	D4-2	Ashley River	81	++	92.6	++	87	++		
93-164	D4-4	Ashley River	23	++	58.8	++	40.8	++		
93-165	D2-1	Ashley River	69	++	94	+	84.4	++		
93-166	D2-3	Ashley River	51	++	72.6	++	47	++		
93-167	D2-2	Ashley River	29.6	++	55.8	++	54	++		
93-168	CHP6	Koppers	42.4	++	66.4	++	56.8	++		
93-169	D3-3	Ashley River	72.2	++	83.2	++	69	++		
93-170	D3-1	Ashley River	12	++	89.2	++	95.8	ns		
93-171	CHP7	Dolphin Cv Marina	5	++	70.6	++	89.8	++		
93-172	D3-2	Ashley River	85.2	++	96.4	ns	96.6	ns		
93-173	E1-1	Cooper River	95	ns	96.2	ns	95.2	ns		
93-174	E2-2	Cooper River	92.2	ns	94.6	+	94	ns		
93-175	E2-1	Cooper River	96.4	ns	94	+	95	ns		
93-176	A1-8	Leadenwah	97.4	ns	98.4	ns	98.6	ns		
93-177	Ref. No.	North Inlet	99.8	ns	99.8	ns	99.2	ns		
93-178	D1-3	Ashley River	94.4	ns	97.6	ns	96.4	ns		
93-179	D1-4	Ashley River	97.6	ns	99.6	ns	98.6	ns		
93-180	CHP3	Aquarium Site	91.2	ns	98.6	ns	97.6	ns		
93-181	E1-3	Cooper River	99	ns	99.6	ns	98.4	ns		
93-182	E1-2	Cooper River	98.4	ns	99	ns	99	ns		
93-183	E3-1	Cooper River	99.8	ns	99.6	ns	99.4	ns		
93-184	E3-3	Cooper River	98.4	ns	98.8	ns	97.6	ns		
93-185	CHP2	Romney St.	99.8	ns	98.6	ns	99.4	ns		
93-186	A3-2	Leadenwah	99	ns	99.4	ns	99.2	ns		
93-196	H4-2	Cooper River	96.8	ns	97.2	ns	95.8	ns		

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	% urchdev in 50% porewater	Significance in 25% porewater	% urchdev in 25% porewater	Significance	UAN, ug/l amphipod start	UAN, ug/l amphipod end	UAN, ug/l amph ave.	Pwater UAN ug/l
93-197	H4-3	Cooper River					2.09	0.48	1.29	14.29
93-198	H4-5	Cooper River					26.85	0.84	13.85	68.88
93-199	H5-2	Cooper River					2.03	0.46	1.24	49.15
93-200	H5-4	Cooper River					3.83	0.66	2.24	38.74
93-201	H5-8	Cooper River					3.16	0.49	1.83	71.31
93-202	H6-1	Cooper River					2.30	0.40	1.35	29.99
93-203	H6-2	Cooper River					17.80	0.54	9.17	113.45
93-204	H6-3	Cooper River					1.55	0.49	1.02	15.45
93-205	H7-1	Cooper River					17.26	0.52	8.89	95.61
93-206	H8-1	Cooper River					29.84	0.49	15.17	122.91
93-207	H2-2	Cooper River					1.12	0.54	0.83	47.81
93-208	H2-3	Cooper River					5.64	0.52	3.08	118.39
93-209	H2-6	Cooper River					2.61	0.42	1.52	30.06
93-210	CHP9	Wallace Channel					19.47	0.66	10.07	6.36
93-211	H3-1	Cooper River					10.32	0.58	5.45	103.66
93-212	H3-3	Cooper River					18.22	0.60	9.41	98.71
93-213	H3-5	Cooper River					26.56	0.66	13.61	134.12
93-214	F1-1	Wando River					6.31	0.61	3.46	66.02
93-215	F1-2	Wando River					24.76	0.73	12.75	74.65
93-216	F1-3	Wando River					0.97	0.55	0.76	39.94
93-217	H1-1	Cooper River					8.91	0.61	4.76	249.01
93-218	H1-4	Cooper River					19.64	0.55	10.10	106.90
93-219	H1-5	Cooper River					34.69	0.72	17.70	87.48
93-220	F2-1	Wando River					5.21	0.41	2.81	112.17
93-221	F2-2	Wando River					7.86	0.48	4.17	80.53
93-222	F2-4	Wando River					12.14	0.54	6.34	51.49
93-223	G1-1	Cooper River					31.22	0.66	15.94	61.09
93-224	G2-1	Cooper River					29.56	0.82	15.19	86.58
93-225	CHP1	Shipyard Creek					46.54	0.60	23.57	86.47

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	% urchdev in 50% porewater	Significance in 25% porewater	% urchdev in 25% porewater	Significance	UAN, ug/l amphipod start	UAN, ug/l amphipod end	UAN, ug/l amph ave.	Pwater UAN ug/l
93-226	CHP10	Detyen's Shipyard					10.94	0.74	5.84	70.75
93-227	Ref. No.	North Inlet					5.43	5.07	5.25	88.76
93-228	B1-1	Winyah Bay					14.38	0.66	7.52	54.74
93-229	B3-1	Winyah Bay					19.37	0.48	9.93	175.67
93-230	B1-3	Winyah Bay					26.58	0.37	13.48	54.26
93-231	B1-2	Winyah Bay					29.28	0.34	14.81	68.41
93-232	CHP11	I of P Connector					5.43	0.40	2.92	18.71
93-233	B7-1	Winyah Bay					16.06	0.34	8.20	32.65
93-234	B6-2	Winyah Bay					15.78	0.35	8.06	22.96
93-235	B5-1	Winyah Bay					3.88	3.39	3.63	7.91
93-236	B4-1	Winyah Bay					11.94	3.27	7.60	10.10
93-237	B2-1	Winyah Bay					22.69	0.44	11.57	72.32
brez	ref1						13.63	0.48	7.05	
slar	ref1						9.56	0.54	0.27	
slar	ref2						58.62	34.09		
slar	ref2						3.01	1.51		
USC lab	ctl batch 2									
USC lab	ctl batch 3									
NIOL Ref 1	batch 1									
NIOL Ref 1	batch 2									
NIOL Ref 1	batch 3									
ns	non-significant									
Λ	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.								
+	statistically significant for Dunnett's t-test at 0.05									
++	statistically significant for Dunnett's t-test at 0.01									
*	statistically significant for Mann-Whitney									
**	statistically significant for Mann-Whitney and Dunnets									
***	statistically significant for Mann-Whitney, Dunnets, and Distribution-free									
Duplicates		BENZ(A)ANTHRACENE	GC, ug/kg	GC, ug/kg	BENZ(O)FLUORANTHENE	GC, ug/kg	BENZ(O)PYRENENE	GC, ug/kg	PERYLENE	GC, ug/kg
93-165	D2-1	Ashley River	407.8	646.2	535.0	223.5	307.98	405.36	128.2	122.2
93-165	D2-1	Ashley River	447.5	546.7	621.2	214.1	280.95	450.64		
93-166	D2-3	Ashley River	221.9	336.4	417.7	162.3	215.57	305.11	80.0	74.7
93-166	D2-3	Ashley River	355.3	359.6	488.9	146.8	201.86	302.05		
93-167	D2-2	Ashley River	754.1	1305.2	1024.1	465.3	488.40	695.00	187.3	197.2
93-167	D2-2	Ashley River	540.2	995.4	1317.7	381.3	562.00	726.80		

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFs Lab. No.	STATION No.	Location	P-450 RGS P-450 Assay CAS Lab.Id.	P-450 RGS Average fold Induction (10ul)	P-450 RGS (a) P Equiv (ug/g)	USC batch no.	copepodite production	Signif * naupliar production	Signif * total production	Signif *
93-46	C1-1	Ashley River								
	C2-4	Ashley River								
93-47	C3-1	Ashley River								
93-48	C4-1	Ashley River								
93-49	CHP4	Plum Isl. Outfall								
93-50	C5-1	Ashley River								
93-51	D1-1	Ashley River								
93-52	A2-3	Leadenwah Creek								
93-53	A3-1	Leadenwah Creek								
93-54	A3-3	Leadenwah Creek								
93-55	A3-3	Leadenwah Creek								
93-56	A2-7	Leadenwah Creek								
93-57	A2-1	Leadenwah Creek								
93-58	A1-1	Leadenwah Creek								
93-59	A1-2	Leadenwah Creek								
93-60	CHP5	Brittle Bank Park								
93-61	CHP8	Brickyard Creek								
93-62	D4-1	Ashley River								
93-63	D4-2	Ashley River								
93-64	D4-4	Ashley River								
93-65	D2-1	Ashley River								
93-66	D2-3	Ashley River								
93-67	D2-2	Ashley River								
93-68	CHP6	Koppers Creosote								
93-69	D3-3	Ashley River								
93-70	D3-1	Ashley River								
93-71	CHP7	Dolphin Cv Marina								
93-72	D3-2	Ashley River								
93-73	E1-1	Cooper River								
93-74	E2-2	Cooper River								
93-75	E2-1	Cooper River								
93-76	A1-8	Leadenwah Creek								
93-77	Ref. No.	North Inlet								
93-78	D1-3	Ashley River								
93-79	D1-4	Ashley River								
93-80	CHP3	Aquarium Site								
93-81	E1-3	Cooper River								
93-82	E1-2	Cooper River								
93-83	E3-1	Cooper River								
93-84	E3-3	Cooper River								
93-85	CHP2	Romney St.								
93-86	A3-2	Leadenwah Creek								
93-96	H4-2	Cooper River								

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	P-450 RGS Assay CAS Lab.Id.	Average fold Induction (10ul)	P-450 RGS (a) P Equiv (ug/g)	P-450 RGS batch no. production	USC copepodite Signif * naupliar production	Signif * total production	Signif *
93-226	CHP10	Detyen's Shipyard							
93-227	Ref. No.	North Inlet							
93-228	B1-1	Winyah Bay		21	12.14	18.8			
93-229	B3-1	Winyah Bay		25	4.19	4.22	2	2.00	5.33
93-230	B1-3	Winyah Bay		23	24.71	37.39			95.33
93-231	B1-2	Winyah Bay		22	18.43	25.57			
93-232	CHP11	I of P Connector							
93-233	B7-1	Winyah Bay		29	26.62	37.4			
93-234	B6-2	Winyah Bay		28	4.76	4.57			
93-235	B5-1	Winyah Bay		27	4.29	1.76			
93-236	B4-1	Winyah Bay		26	5.10	4.59			
93-237	B2-1	Winyah Bay		24	9.81	11.97			
bref	ref1								
star	ref2								
star	ref2								
USC lab ct1	batch 2								
USC lab ct1	batch 3								
NIOL Ref 1	batch 1								
NIOL Ref 1	batch 2								
NIOL Ref 1	batch 3								
ns	non-significant								
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177.	NMFS 93-196 to 93-237 compared to reference 227.						
+		statistically significant for Dunnett's t-test at 0.05							
++		statistically significant for Dunnett's t-test at 0.01							
*		statistically significant for Mann-Whitney							
**		statistically significant for Mann-Whitney and Dunnett's							
***		statistically significant for Mann-Whitney, Dunnett's, and Distribution-free							
Duplicates		INDENO(1,2,3-C,D)PYRENE		DIBENZ(A,H)ANTH(1)PERYLENE		10PAH			
93-165	D2-1	Ashley River	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg ;C, ug/kg			
93-165	D2-1	Ashley River		212.4	56.0	176.6	8441.4		
93-166	D2-3	Ashley River		288.5	52.0	220.0	7094.4		
93-166	D2-3	Ashley River		181.0	33.6	136.1	3610.8		
93-167	D2-2	Ashley River		182.1	29.6	116.7	4119.3		
93-167	D2-2	Ashley River		340.2	65.9	272.5	8739.8		
				488.8	107.4	377.8	9419.5		

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. STATION No.	STATION No.	Location	clutch size	Signif *	Ag (µg/g)	Al (%)	As (µg/g)	Cd (µg/g)	Cr (µg/g)	Cu (µg/g)	Fe (%)	Hg (µg/g)	Mn (µg/g)	
93-146	C1-1	Ashley River	<	0.12 < 0.88	3.6	0.11	7.6	1.2	0.4	< 0.01	0.4	< 0.01	64	
93-147	C2-4	Ashley River	<	0.12 < 0.88	6.0	0.14	8.3	1.5	0.65	< 0.01	0.01	140		
93-148	C3-1	Ashley River	<	0.12 < 0.88	3.4	0.23	10.1	1.3	0.46	< 0.01	0.01	88		
93-149	C4-1	Ashley River	<	0.16 < 0.88	1.6	< 0.048	4.8	1.2	< 0.3	< 0.01	0.3	38		
93-150	CHP4	Plum Isl. Outfall	8.79 *											
93-151	C5-1	Ashley River	<	0.12 < 0.12	2.1	6.6	0.13	27.7	17.5	1.16	< 0.01	0.01	209	
93-152	D1-1	Ashley River	<	0.12 < 0.12	5	18.9	0.15	58.5	13.6	2.91	0.16	0.16	436	
93-153	A2-3	Leadenwah Creek												
93-154	A3-1	Leadenwah Creek												
93-155	A3-3	Leadenwah Creek												
93-156	A2-7	Leadenwah Creek												
93-157	A2-1	Leadenwah Creek	<	0.12 < 0.12	4.8	16.1	0.07	48.6	12.1	2.51	< 0.01	0.01	142	
93-158	A1-1	Leadenwah Creek	<	0.12 < 0.12	4.3	12.1	0.21	52.5	19.2	2.4	0.15	0.05	291	
93-159	A1-2	Leadenwah Creek	<	0.12 < 0.12	3.3	0.21	17.2	6.3	0.68	0.05	0.05	97		
93-160	CHP5	Brittle Bank Park	<	0.12 < 0.88										
93-161	CHP8	Brickyard Creek	<	0.12 < 0.88										
93-162	D4-1	Ashley River	<	0.12 < 0.88	4.5	0.05	15.2	4.9	0.65	0.05	0.05	136		
93-163	D4-2	Ashley River	<	0.12 < 0.88	2.0	< 0.048	9.1	2.6	< 0.3	< 0.01	0.01	34		
93-164	D4-4	Ashley River	<	0.19 < 4.5	20.0	0.29	69.1	25.2	2.79	0.1	0.1	415		
93-165	D2-1	Ashley River	<	0.19 < 4.5	0.15	12.7	5.4	0.43	< 0.01	0.01	0.01	100		
93-166	D2-3	Ashley River	<	0.12 < 0.88	3.5	0.12	14.7	10.1	0.76	0.03	0.03	131		
93-167	D2-2	Ashley River	<	0.12 < 0.88	5.8	0.14	11.6	12.5	0.45	0.02	0.02	85		
93-168	CHP6	Koppers Creosote	13.56 *											
93-169	D3-3	Ashley River	<	0.12 < 0.88	3.1	0.14	80.4	29.3	3.3	0.13	0.13	502		
93-170	D3-1	Ashley River	0.17	5.3	19.4	0.3	80.4	29.3	3.3	0.13	0.13	502		
93-171	CHP7	Dolphin Cv Marina	0.12	2.8	9.9	0.21	51.6	17.0	1.71	0.03	0.03	221		
93-172	D3-2	Ashley River	0.14	2.7	11.2	0.22	42.8	21.5	1.44	0.09	0.09	177		
93-173	E1-1	Cooper River	<	0.12 < 0.88	4.2	12.5	0.27	67.4	23.7	2.57	0.11	0.11	372	
93-174	E2-2	Cooper River	<	0.12 < 0.12	1.5	7.8	0.49	46.4	9.7	0.6	< 0.01	0.01	116	
93-175	E2-1	Cooper River	<	0.12 < 0.12	1.2	3.0	< 0.048	13.8	3.2	0.63	< 0.01	0.01	78	
93-176	A1-8	Leadenwah Creek	14.00 *											
93-177	Ref. No.	North Inlet	<	0.12 < 0.12	3.2	9.4	0.11	37.7	8.5	1.88	0.05	0.05	112	
93-178	D1-3	Ashley River	<	0.12 < 0.88	1	1.6	0.13	17.5	7.7	0.66	< 0.01	0.01	116	
93-179	D1-4	Ashley River	10.48 *											
93-180	CHP3	Aquarium Site	13.96 *											
93-181	E1-3	Cooper River	<	0.12 < 0.88	4.9	18.6	0.28	60.1	22.0	2.94	0.11	0.11	437	
93-182	E1-2	Cooper River	<	0.12 < 0.12	5.7	0.08	15.2	3.5	0.72	0.03	0.03	123		
93-183	E3-1	Cooper River	11.90 *											
93-184	E3-3	Cooper River	<	0.12 < 0.12	2.3	9.1	0.14	31.8	8.6	1.37	0.04	0.04	178	
93-185	CHP2	Romney St.	<	0.12 < 0.12	3.7	9.4	0.1	48.1	7.1	2.97	0.11	0.11	276	
93-186	A3-2	Leadenwah Creek	<	0.12 < 0.12	5.5	19.9	0.23	66.6	14.8	3.02	0.71	0.71	493	
93-196	H4-2	Cooper River	<	0.12 < 0.88	2.2	< 0.048	5.4	1.6	0.35	< 0.01	0.01	0.01	90	

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	clutch size	Signif *	Ag (µg/g)	Al (%)	As (µg/g)	Cd (µg/g)	Cr (µg/g)	Cu (µg/g)	Fe (%)	Hg (µg/g)	Mn (µg/g)
93-226	CHP10	Detyen's Shipyards	<	0.12	2.2	5.1	0.14	34.9	75.5	1.47	0.08	135	
93-227	Ref. No.	North Inlet	<	0.12	< 0.88	2.7	< 0.048	10.0	2.2	0.49	< 0.01	26	
93-228	B1-1	Winyah Bay		0.16	7.7	18.9	0.42	97.8	36.2	5.46	0.26	726	
93-229	B3-1	Winyah Bay	100.67	13.24	0.11	8	21.1	0.23	75.2	29.3	4.61	0.19	999
93-230	B1-3	Winyah Bay		0.15	7	21.3	0.52	112.8	38.0	5.14	0.16	651	
93-231	B1-2	Winyah Bay		0.13	10.3	16.4	0.41	99.1	38.2	6.47	0.14	579	
93-232	CHP11	I of P Connector	<	0.12	5.6	21.3	0.55	63.7	16.5	3.15	0.4	231	
93-233	B7-1	Winyah Bay	<	0.12	6.5	14.1	0.11	67.8	23.3	3.84	< 0.01	952	
93-234	B6-2	Winyah Bay	<	0.12	5.8	12.0	0.16	60.5	24.1	3.62	0.12	1123	
93-235	B5-1	Winyah Bay	<	0.12	2.8	4.1	< 0.048	28.1	9.7	1.51	< 0.01	173	
93-236	B4-1	Winyah Bay	<	0.12	6.1	16.9	0.16	49.5	24.3	3.53	0.68	503	
93-237	B2-1	Winyah Bay		0.14	8.1	17.6	0.2	79.2	31.0	4.57	0.93	1005	
brez	ref1												
brez	ref2												
star	ref2												
star	ref2												
USC lab ct1 batch 2					13.92								
USC lab ct1 batch 3					12.06								
NIOL Ref 1 batch 1					17.80								
NIOL Ref 1 batch 2					13.55								
NIOL Ref 1 batch 3					15.39								
ns	non-significant												
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177.											
+		statistically significant for Dunnett's t-test at 0.05											
++		statistically significant for Dunnett's t-test at 0.01											
*		statistically significant for Mann-Whitney and Dunnett's											
**		statistically significant for Mann-Whitney, Dunnett's, and Distribution-free											

Duplicates													
93-165	D2-1	Ashley River											
93-165	D2-1	Ashley River											
93-166	D2-3	Ashley River											
93-166	D2-3	Ashley River											
93-167	D2-2	Ashley River											
93-167	D2-2	Ashley River											

ns non-significant
^ Microtox Tests: NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

*

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Ni ($\mu\text{g/g}$)	Pb ($\mu\text{g/g}$)	Se ($\mu\text{g/g}$)	Sn ($\mu\text{g/g}$)	Zn ($\mu\text{g/g}$)	% sand %	% silt %	% clay %	% fines %	% TOC %	Cd ($\mu\text{mole/g}$)	Cu ($\mu\text{mole/g}$)	
93-146	C1-1	Ashley River	< 1.4	4.0	0.11	7.8	9.4	95.29	3.00	1.72	4.71	0.11	0.0007	*	0.0018 *
93-147	C2-4	Ashley River	< 1.4	5.5	0.22	< 7.8	18.1	96.84	0.11	3.05	3.16	0.19	0.0021	0.0060	
93-148	C3-1	Ashley River	< 1.5	3.5	0.12	< 7.8	13.2	96.22	0.81	2.97	3.78	0.13	0.0007	0.0029	
93-149	C4-1	Ashley River	< 1.4	3.8	< 0.11	< 7.8	6.7	94.50	0.20	5.29	5.50	0.16	0.0009	*	0.0042 *
93-150	CHP4	Plum Isl. Outfall	< 1.4												
93-151	C5-1	Ashley River													
93-152	D1-1	Ashley River	7.4	63.1	0.21	24.5	38.0	79.26	9.13	11.61	20.74	1.04	0.0030	*	0.7430
93-153	A2-3	Leadenwah Creek	18.6	20.8	0.37	45.2	62.3	10.72	38.46	50.82	89.28	3.45	0.0016	*	0.0547
93-154	A3-1	Leadenwah Creek													
93-155	A3-3	Leadenwah Creek													
93-156	A2-7	Leadenwah Creek													
93-157	A2-1	Leadenwah Creek													
93-158	A1-1	Leadenwah Creek	14	18.2	0.42	33.8	52.0	29.06	25.77	45.17	70.94	2.71	0.0059	*	0.0152 *
93-159	A1-2	Leadenwah Creek													
93-160	CHP5	Brittle Bank Park	14.7	37.0	0.46	40.7	71.1	35.41	18.67	45.93	64.59	1.90	0.0025	*	0.0594
93-161	CHP8	Brickyard Creek	6	12.9	0.36	< 7.8	31.7	87.53	1.53	10.94	12.47	0.77	0.0015	*	0.0318
93-162	D4-1	Ashley River													
93-163	D4-2	Ashley River	2.9	9.6	0.21	< 7.8	40.1	80.60	15.12	4.28	19.40	0.60	0.0010	*	0.0284
93-164	D4-4	Ashley River	< 1.4	5.3	< 0.11	< 7.8	10.8	92.88	6.38	0.74	7.12	0.43	0.0028	*	0.0047 *
93-165	D2-1	Ashley River	18.2	37.7	0.44	49.8	87.7	6.50	55.75	37.75	93.50	3.65	0.0016	*	0.1470
93-166	D2-3	Ashley River	3.5	10.4	0.25	< 7.8	19.0	90.47	4.53	5.00	9.53	0.43	0.0024	*	0.2401
93-167	D2-2	Ashley River	5.4	16.6	0.35	11.6	45.6	82.78	9.76	7.46	17.22	0.96	0.0039	*	0.0146
93-168	CHP6	Koppers Creosote	4.7	15.5	0.34	11.8	34.5	96.15	2.04	1.81	3.85	0.37	0.0004	*	0.0615
93-169	D3-3	Ashley River	21.6	42.1	0.67	49.1	113.3	2.76	48.64	48.60	97.24	4.59	0.0013	*	0.1733
93-170	D3-1	Ashley River	11.3	30.8	0.42	26.8	64.8	54.59	19.39	26.02	45.41	1.92	0.0045	*	0.0116 *
93-171	CHP7	Dolphin Cv Marina	10.1	34.6	0.39	23.8	59.0	55.53	12.41	32.06	44.47	2.02	0.0054	*	0.0810
93-172	D3-2	Ashley River	16.8	36.6	0.48	34.8	84.7	21.53	25.87	52.60	78.47	3.24	0.0108	*	0.0278 *
93-173	E1-1	Cooper River	4.3	18.1	0.31	< 7.8	49.0	89.14	6.01	4.85	10.86	0.46	0.0014	*	0.0226
93-174	E2-2	Cooper River	17.9	5.1	0.24	< 7.8	45.6	38.72	21.03	40.25	61.28	1.36	0.0013	*	0.0044 *
93-175	E2-1	Cooper River													
93-176	A1-8	Leadenwah Creek	2.6	7.6	0.12	13	15.4	86.58	3.60	9.82	13.42	0.58	0.0009	*	0.0189 *
93-177	Ref. No.	North Inlet	13.2	17.2	0.23	20.7	19.1	54.65	6.72	38.63	45.35	1.81	0.0013	*	0.0049 *
93-178	D1-3	Ashley River	4.2	73.9	0.21	16.5	28.4	86.37	3.93	9.70	13.63	0.65	0.0032	*	0.8456
93-179	D1-4	Ashley River	< 1.4	5.1	0.13	< 7.8	12.2	96.46	3.35	0.19	3.54	0.18	0.0013	*	0.0040
93-180	CHP3	Aquarium Site	20.3	39.8	0.31	38.3	85.2	4.21	31.79	63.99	95.79	3.23	0.0030	*	0.0947
93-181	E1-3	Cooper River	2.5	10.8	0.13	12	21.7	86.35	5.24	8.41	13.65	0.52	0.0005	*	0.0356
93-182	E1-2	Cooper River													
93-183	E3-1	Cooper River	13.5	8.3	0.25	40.5	38.3	53.65	17.29	29.06	46.35	1.03	0.0006	*	0.0017 *
93-184	E3-3	Cooper River	8.6	11.7	0.25	< 7.8	35.3	66.91	16.88	16.21	33.09	1.38	0.0027	*	0.1655
93-185	CHP2	Romney St.	11.8	14.7	0.20	33.7	43.7	21.48	32.38	46.14	78.52	1.12	0.0005	*	0.0099
93-186	A3-2	Leadenwah Creek	19.7	31.3	0.59	45.3	66.3	9.30	27.69	63.01	90.70	3.47	0.0020	*	0.2182
93-196	H4-2	Cooper River	< 1.4	5.2	< 0.11	< 7.8	15.4	93.75	1.10	5.15	6.25	0.30	0.0013	*	0.0029 *

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Ni ($\mu\text{g/g}$)	Pb ($\mu\text{g/g}$)	Se ($\mu\text{g/g}$)	Sn ($\mu\text{g/g}$)	Zn ($\mu\text{g/g}$)	% sand %	% silt %	% clay %	% fines %	% TOC %	Cd ($\mu\text{mole/g}$)	Cu ($\mu\text{mole/g}$)	
93-197	H4-3	Cooper River	21.8	31.5	0.41	44.7	84.7	11.71	19.98	68.31	88.29	3.09	0.0012 *	0.1138	
93-198	H4-5	Cooper River													
93-199	H5-2	Cooper River													
93-200	H5-4	Cooper River	4.9	7.8	0.17	12.6	23.5	82.59	4.14	13.27	17.41	0.53	0.0031 *	0.0081 *	
93-201	H5-8	Cooper River													
93-202	H6-1	Cooper River	6.8	7.9	0.39	<	7.8	25.2	73.70	3.99	22.31	26.30	0.60	0.0015 *	0.4343
93-203	H6-2	Cooper River	13.1	14.7	0.38	28.6	51.1	52.38	16.26	31.36	47.62	1.70	0.0024 *	0.0269	
93-204	H6-3	Cooper River	< 1.4	2.5	< 0.11	< 7.8	8.4	93.60	0.99	5.41	6.40	0.13	0.0002 *	0.0215	
93-205	H7-1	Cooper River	9.7	10.7	0.31	< 7.8	44.5	28.90	13.11	57.99	71.10	1.59	0.0034 *	0.0089 *	
93-206	H8-1	Cooper River	22.9	29.7	0.87	50.4	93.0	3.54	24.68	71.77	96.46	4.65	0.0065 *	3.0184	
93-207	H2-2	Cooper River	< 1.4	2.7	0.14	< 7.8	7.7	97.13	2.60	0.27	2.87	0.18	0.0010 *	0.0055 *	
93-208	H2-3	Cooper River	8.6	9.7	0.39	14.9	33.2	82.20	2.14	15.66	17.80	1.55	0.0093 *	0.0108 *	
93-209	H2-6	Cooper River	26.9	34.6	0.47	54.9	91.7	20.04	26.16	53.81	79.96	2.61	0.0028 *	0.1426	
93-210	CHP9	Wallace Channel	16.1	19.8	0.48	40.4	69.8	19.66	24.14	56.20	80.34	4.14	0.0033 *	0.0784	
93-211	H3-1	Cooper River													
93-212	H3-3	Cooper River	10.3	10.9	0.34	20.9	38.3	64.95	17.23	17.82	35.05	1.18	0.0007 *	0.0019 *	
93-213	H3-5	Cooper River													
93-214	F1-1	Wando River	3	6.2	0.38	7.8	15.5	95.09	0.36	4.55	4.91	0.22	0.0015 *	0.0091	
93-215	F1-2	Wando River	10.8	15.4	0.27	22.8	56.6	43.61	14.63	41.77	56.39	1.84	0.0039 *	0.0148 *	
93-216	F1-3	Wando River													
93-217	H1-1	Cooper River	6.2	10.2	0.43	17.4	26.9	80.81	6.29	12.90	19.19	0.83	0.0038 *	0.0098 *	
93-218	H1-4	Cooper River	8.8	4.4	0.73	9.9	22.9	86.56	4.28	9.16	13.44	0.56	0.0024 *	0.0063	
93-219	H1-5	Cooper River	20.8	28.3	0.31	47	76.0	5.06	33.46	61.48	94.94	3.10	0.0025 *	0.0162	
93-220	F2-1	Wando River													
93-221	F2-2	Wando River	3.8	< 0.11	< 7.8	12.9	95.55	1.51	2.94	4.45	0.32	0.0003 *	0.0077		
93-222	F2-4	Wando River	1.5	4.7	0.17	< 7.8	16.1	91.73	6.00	2.27	8.27	0.30	0.0008 *	0.0609	
93-223	G1-1	Cooper River	22.2	38.9	0.34	53.9	85.9	0.53	29.84	69.63	99.47	3.25	0.0050 *	0.0837	
93-224	G2-1	Cooper River	23.3	41.5	0.61	56.7	97.2	0.35	40.09	59.57	99.65	2.47	0.0048 *	0.1005	
93-225	CHP1	Shipyard Creek	24	42.6	0.53	97	105.8	0.07	36.78	63.15	99.93	3.58	0.0040 *	0.1653	

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Ni ($\mu\text{g/g}$)	Pb ($\mu\text{g/g}$)	Se ($\mu\text{g/g}$)	Sn ($\mu\text{g/g}$)	Zn ($\mu\text{g/g}$)	% sand	% silt	% clay	% fines	% TOC %	Cd ($\mu\text{mole/g}$)	Cu ($\mu\text{mole/g}$)	
93-226	CHP10	Detyen's Shipyard	10.4	19.4	0.56	21.9	67.8	52.33	14.65	33.02	47.67	2.09	0.0044 *	0.0114 *	
93-227	Ref. No.	North Inlet	4.2	7.5	0.11	<	7.8	16.3	84.81	1.70	13.49	15.19	0.70	0.0023 *	0.0041 *
93-228	B1-1	Winyah Bay	26.6	53.8	0.53	57.8	183.5	0.37	45.97	53.66	99.63	4.98	0.0122 *	0.1747	
93-229	B3-1	Winyah Bay	29.7	35.3	0.65	62.1	103.2	2.95	32.81	64.24	97.05	4.82	0.0005 *	0.0424	
93-230	B1-3	Winyah Bay	29.8	54.2	0.48	66.6	196.3	0.28	39.69	60.03	99.72	4.75	0.0095 *	0.0245 *	
93-231	B1-2	Winyah Bay	30	44.1	0.52	59.5	176.9	0.69	31.10	68.21	99.31	4.87	0.0110 *	0.0283 *	
93-232	CHP11	I of P Connector	20.7	19.7	0.36	38.8	66.9	30.40	18.65	50.95	69.60	2.57	0.0037 *	0.5526	
93-233	B7-1	Winyah Bay	24.6	31.7	0.35	47.7	91.7	15.14	26.99	57.87	84.86	3.71	0.0039 *	0.0958	
93-234	B6-2	Winyah Bay	25	29.5	0.48	56.7	91.4	34.54	24.92	40.53	65.46	3.06	0.0006 *	0.0576	
93-235	B5-1	Winyah Bay	10.4	11.1	0.21	22.2	31.1	68.19	8.59	23.22	31.81	0.80	0.0025 *	0.0364	
93-236	B4-1	Winyah Bay	25.8	35.0	0.53	46.5	90.7	1.61	60.75	37.64	98.39	3.44	0.0109 *	0.0280 *	
93-237	B2-1	Winyah Bay	30.4	45.6	0.72	76.5	123.9	0.01	22.46	77.53	99.99	4.88	0.0068 *	0.0090 *	
brez	ref1														
star	ref2														
star	ref2														
USC lab	ctl batch 2														
USC lab	ctl batch 3														
NIOL	Ref 1 batch 1														
NIOL	Ref 1 batch 2														
NIOL	Ref 1 batch 3														
ns	non-significant														
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177.													
+		statistically significant for Dunnett's t-test at 0.05													
++		statistically significant for Dunnett's t-test at 0.01													
*		statistically significant for Mann-Whitney													
**		statistically significant for Mann-Whitney and Dunnett's													
***		statistically significant for Mann-Whitney, Dunnnett's, and Distribution-free													
Duplicates															
93-165	D2-1	Ashley River													
93-165	D2-1	Ashley River													
93-166	D2-3	Ashley River													
93-166	D2-3	Ashley River													
93-167	D2-2	Ashley River													
93-167	D2-2	Ashley River													

ns non-significant
^ Microtox Tests: NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

*

** statistically significant for Mann-Whitney

*** statistically significant for Mann-Whitney and Dunnnett's

statistically significant for Mann-Whitney, Dunnnett's, and Distribution-free

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Ni (µmole/g)	Pb (µmole/g)	Zn (µmole/g)	Hg (µmole/g)	SEM (µmole/g)	AVS (µmole/g)	SEM:AVS ratio	NAPHTHALENE GC, ug/kg	
93-146	C1-1	Ashley River	0.0282	0.0115 *	0.0008 *	0.0000 *	0.043	0.072	0.59801	< 5	
93-147	C2-4	Ashley River	0.0309	0.0113 *	0.0008 *	0.0000 *	0.051	0.148	0.34606	< 5	
93-148	C3-1	Ashley River	0.0262	0.0108	0.1550	0.0000 *	0.196	0.026	7.44533	t 6.1	
93-149	C4-1	Ashley River	0.0354	0.0145 *	0.0011 *	0.0000 *	0.056	0.973	0.05759	7.3	
93-150	CHP4	Plum Isl. Outfall	0.0354	0.0145 *	0.0011 *	0.0000 *	0.056	0.973	0.05759	7.3	
93-151	C5-1	Ashley River	0.1350	0.8605	0.4055	0.0001 *	2.147	1.332	1.61207	t 31.3	
93-152	D1-1	Ashley River	0.0390	0.0261 *	0.0019 *	0.0000 *	0.123	0.171	0.72054	20.2	
93-153	A2-3	Leadenwah Creek								16.5	
93-154	A3-1	Leadenwah Creek								19.4	
93-155	A3-3	Leadenwah Creek								14.8	
93-156	A2-7	Leadenwah Creek								18.9	
93-157	A2-1	Leadenwah Creek								18.3	
93-158	A1-1	Leadenwah Creek	0.0930	*	0.1147 *	1.3768	0.0001 *	1.606	9.597	0.16731	
93-159	A1-2	Leadenwah Creek								9.7	
93-160	CHP5	Brittle Bank Park	0.2925	0.0399 *	0.5564	0.0001 *	0.951	0.702	1.35404	t 12.7	
93-161	CHP8	Brickyard Creek	0.0610	0.0242 *	0.3836	0.0000 *	0.502	1.638	0.30656	38.9	
93-162	D4-1	Ashley River								9.4	
93-163	D4-2	Ashley River	0.0196	*	0.0168 *	0.0012 *	0.0000 *	0.067	0.329	0.20382	
93-164	D4-4	Ashley River	0.0290	*	0.0299 *	0.1888	0.0000 *	0.255	1.705	0.14967	
93-165	D2-1	Ashley River	0.0246	*	0.0819	0.8398	0.0000 *	1.095	0.178	6.14921	
93-166	D2-3	Ashley River	0.2202	*	0.0106 *	0.2217	0.0000 *	0.695	0.401	1.73458	
93-167	D2-2	Ashley River	0.0412	*	0.0305 *	0.4687	0.0000 *	0.559	0.683	0.81797	
93-168	CHP6	Koppers Creosote	0.0159		0.0350	0.0004 *	0.0000 *	0.113	0.188	0.60236	
93-169	D3-3	Ashley River	0.0556		0.0805	1.0011	0.0000 *	1.312	0.224	5.85407	
93-170	D3-1	Ashley River	0.0708	*	0.0730 *	0.0054 *	0.0001 *	0.165	8.717	0.01897	
93-171	CHP7	Dolphin Cv Marina	0.0998	*	0.0878 *	0.0065 *	0.0001 *	0.281	6.442	0.04358	
93-172	D3-2	Ashley River	0.1697	*	0.1750 *	0.0129 *	0.0003 *	0.396	0.000	- t	
93-173	E1-1	Cooper River	0.0436	*	0.1242	0.5883	0.0000 *	0.780	0.430	1.81271	
93-174	E2-2	Cooper River	0.0710	0.0216 *	0.4189	0.0000 *	0.517	0.788	0.65651	11	
93-175	E2-1	Cooper River								29.2	
93-176	A1-8	Leadenwah Creek	0.0290	*	0.0143 *	0.0011 *	0.0000 *	0.064	0.100	0.64305	
93-177	Ref. No.	North Inlet	0.0302	*	0.0312 *	0.0023 *	0.0000 *	0.070	0.171	0.40929	
93-178	D1-3	Ashley River	3.7439	0.1341	0.3351	0.0000 *	5.062	1.139	4.44348	t < 5	
93-179	D1-4	Ashley River	0.0313	0.0125 *	0.2105	0.0000 *	0.260	0.083	3.14726	t 22.6	
93-180	CHP3	Aquarium Site	0.1276	0.0427 *	0.6171	0.0001 *	0.885	2.570	0.34448	67.3	
93-181	E1-3	Cooper River	0.0136	*	0.0094 *	0.1845	0.0000 *	0.244	0.056	4.32417	
93-182	E1-2	Cooper River								25.1	
93-183	E3-1	Cooper River	0.0190	*	0.0105 *	0.3230	0.0000 *	0.355	0.059	6.00830	
93-184	E3-3	Cooper River	0.0852	*	0.0433 *	0.3767	0.0001 *	0.674	2.475	0.27210	
93-185	CHP2	Romney St.	0.0392	0.0089 *	0.2657	0.0000 *	0.324	0.021	15.32747	6.8	
93-186	A3-2	Leadenwah Creek	0.3177	0.0431 *	0.0014 *	0.0000 *	0.582	0.087	6.65992	t 8	
93-196	H4-2	Cooper River	0.0308	*	0.0182 *	0.1759	0.0001 *	0.229	0.428	0.53587	25.8

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Ni (μmole/g)	Pb (μmole/g)	Zn (μmole/g)	Hg (μmole/g)	SEM (μmole/g)	AVS (μmole/g)	SEM:AVS ratio	NAPHTHALENE GC, ug/kg
93-197	H4-3	Cooper River	0.0939	0.0453 *	0.5453	0.0000	*	0.800	0.182	4.39323 t
93-198	H4-5	Cooper River	0.0493	* 0.0509	*	0.0038	*	0.0001	*	0.115
93-199	H5-2	Cooper River	0.1492	0.0243 *	1.1461	0.0000	*	1.756	1.305	1.34550 t
93-200	H5-4	Cooper River	0.1562	0.0397 *	0.4002	0.0001	*	0.625	2.965	0.21093
93-201	H5-8	Cooper River	0.0072	* 0.0032	*	0.0002	*	0.0000	*	0.032
93-202	H6-1	Cooper River	0.0544	* 0.0910	*	0.5725	0.0001	*	0.730	11.037
93-203	H6-2	Cooper River	3.8163	0.1306 *	6.4915	0.0001	*	13.463	1.884	7.14442 t
93-204	H6-3	Cooper River	0.0479	0.0149 *	0.1164	0.0000	*	0.186	0.200	0.92948
93-205	H7-1	Cooper River	0.0792	* 0.0679	*	0.6308	0.0001	*	0.798	0.820
93-206	H8-1	Cooper River	0.2843	0.0456 *	0.8945	0.0001	*	1.370	2.750	0.49804
93-207	H2-2	Cooper River	0.2587	0.0540 *	0.6281	0.0001	*	1.023	0.866	1.18007 t
93-208	H2-3	Cooper River	0.0115	* 0.0264	*	0.2516	0.0000	*	0.292	0.951
93-209	H2-6	Cooper River	0.0139	* 0.0122	*	0.2547	0.0000	*	0.291	0.30707
93-210	CHP9	Wallace Channel	0.1784	0.0627 *	0.5042	0.0001	*	0.764	6.716	0.11377
93-211	H3-1	Cooper River	0.0601	* 0.0620	*	0.4439	0.0001	*	0.580	4.442
93-212	H3-3	Cooper River	0.0322	* 0.0079	*	0.2677	0.0000	*	0.316	0.263
93-213	H3-5	Cooper River	0.0228	* 0.1030	*	0.2375	0.0000	*	0.382	0.487
93-214	F1-1	Wando River	0.0205	0.0053 *	0.1868	0.0000	*	0.221	0.030	7.30357 t
93-215	F1-2	Wando River	0.0971	0.0137 *	0.2367	0.0000	*	0.409	0.187	2.18315 t
93-216	F1-3	Wando River	0.4468	0.0765 *	0.7552	0.0001	*	1.367	1.529	0.89430
93-217	H1-1	Cooper River	2.7185	0.0788 *	0.7092	0.0001	*	3.612	2.731	1.32276 t
93-224	G2-1	Cooper River	0.2777	0.0656 *	1.3933	0.0001	*	1.906	0.658	2.89583 t
93-225	CHP1	Shipyard Creek								70.2

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	Ni (μmole/g)	Pb (μmole/g)	Zn (μmole/g)	Hg (μmole/g)	SEM (μmole/g)	AVS (μmole/g)	SEM:AVS ratio	NAPHTHALENE GC, ug/kg
93-226	CHP10	Deyen's Shipyard	0.0697 *	0.0937 *	0.1834	0.0001 *	0.363	9.775	0.03710	19.6
93-227	Ref. No.	North Inlet	0.0253 *	0.0261 *	0.2960	0.0000 *	0.354	1.876	0.18859	
93-228	B1-1	Winyah Bay	0.1735 *	0.0955 *	0.0071 *	0.0001 *	0.463	2.001	0.23140	26.3
93-229	B3-1	Winyah Bay	0.0744	0.0176 *	0.3707	0.0000 *	0.506	0.000 -	t	52.9
93-230	B1-3	Winyah Bay	0.1500 *	0.2307 *	1.6684	0.0002 *	2.083	42.674	0.04882	35.8
93-231	B1-2	Winyah Bay	0.1729 *	0.1782 *	0.0132 *	0.0003 *	0.404	72.677	0.00556	39.9
93-232	CHP11	I of P Connector	0.3511	0.0598 *	0.5437	0.0005	1.511	0.440	3.43662 t	13.8
93-233	B7-1	Winyah Bay	0.1213 *	0.0633 *	0.7876	0.0015	1.073	1.522	0.70520	35.2
93-234	B6-2	Winyah Bay	0.0648	0.0420	0.3523	0.0000 *	0.517	0.000 -	t	9.6
93-235	B5-1	Winyah Bay	0.2507	0.0249 *	0.3902	0.0000 *	0.705	1.696	0.41550	23.6
93-236	B4-1	Winyah Bay	0.1713 *	0.1766 *	0.0131 *	0.0003 *	0.400	30.117	0.01329	15.8
93-237	B2-1	Winyah Bay	0.6795	0.0567 *	0.8496	0.0001 *	1.602	2.096	0.76433	15.4
brez	ref1									
brez	ref1									
slar	ref2									
slar	ref2									
USC lab cfl batch 2										
USC lab cfl batch 3										
NIOL Ref 1 batch 1										
NIOL Ref 1 batch 2										
NIOL Ref 1 batch 3										
ns	non-significant									
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.								
+	statistically significant for Dunnett's t-test at 0.05									
++	statistically significant for Dunnett's t-test at 0.01									
*	statistically significant for Mann-Whitney									
**	statistically significant for Mann-Whitney and Dunnett's									
***	statistically significant for Mann-Whitney, Dunnett's, and Distribution-free									
Duplicates										
93-165	D2-1	Ashley River								
93-165	D2-1	Ashley River								
93-166	D2-3	Ashley River								
93-166	D2-3	Ashley River								
93-167	D2-2	Ashley River								
93-167	D2-2	Ashley River								

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	2-METHYLNAPHTHALENE GC, ug/kg	1-METHYLNAPHTHALENE GC, ug/kg	2,6-DIMETHYLNAPHTHALENE GC, ug/kg	BIPHENYL GC, ug/kg	ACENAPHTHYLENE GC, ug/kg	ACENAPHTHENE GC, ug/kg
93-146	C1-1	Ashley River	3.1 <	2	1.9 <	3 <	1 <	3
93-147	C2-4	Ashley River	3.0 <	2 <	2 <	3	1.1 <	3
93-148	C3-1	Ashley River	3.3 <	2 <	2 <	3	1.2 <	3
93-149	C4-1	Ashley River	3.2 <	2 <	2 <	3	1 <	3
93-150	CHP4	Plum Isl. Outfall	3.9	2.4	2 <	3 <	1	3
93-151	C5-1	Ashley River	15.0	7.1	6.5	3.8	27	11.4
93-152	D1-1	Ashley River	10.0	4.7	6.3	3.9	16.2	22.5
93-153	A2-3	Leadenwah Creek	9.6	5.7	4.9 <	3 <	1 <	3
93-154	A3-1	Leadenwah Creek	10.4	5.7	5.5 <	3 <	1 <	3
93-155	A3-3	Leadenwah Creek	8.5	4.6	4 <	3	3.3 <	3
93-156	A2-7	Leadenwah Creek	9.7	5	4.8 <	3 <	1 <	3
93-157	A2-1	Leadenwah Creek	9.5	5	5 <	3 <	1 <	3
93-158	A1-1	Leadenwah Creek	5.9	2.9	2.6 <	3 <	1 <	3
93-159	A1-2	Leadenwah Creek	6.6	3.5	3.6 <	3 <	1 <	3
93-160	CHP5	Brittle Bank Park	25.5	12.9	14.4	7.9	41.4	14.5
93-161	CHP8	Brickyard Creek	6.8	3.9	3.5 <	3	5.1	5.4
93-162	D4-1	Ashley River	9.5	5.3	6.9 <	3	6	69.9
93-163	D4-2	Ashley River	7.9	4.2	3.6 <	3	6.9 <	3
93-164	D4-4	Ashley River	46.7	12.1	17.4	15.85	56.65	37.3
93-165	D2-1	Ashley River	16.6	8.5	7.8	4.75	46.05	41.5
93-166	D2-3	Ashley River	98.3	50.85	58.05	28.6	103.65	95.25
93-167	D2-2	Ashley River	20.1	15	18.7	7.1	40.8	187.8
93-168	CHP6	Koppers Creosote	41.1	19	18.6	9.7	50.6	35
93-169	D3-3	Ashley River	21.1	9.5	8.4	3.9	22.7	8
93-170	D3-1	Ashley River	20.2	15.6	11.6	7.5	53	67.3
93-171	CHP7	Dolphin Cv Marina	37.0	17.4	17.9	9	34.7	40.1
93-172	D3-2	Ashley River	12.2	5.8	4.9 <	3	1.8 <	3
93-173	E1-1	Cooper River	8.4	4.4	5.4 <	3	1.6	6.4
93-174	E2-2	Cooper River	11.3	6.5	5.7	3.7	10.4	11.4
93-175	E2-1	Cooper River	4.1	2.2	1.8 <	3 <	1 <	3
93-176	A1-8	Leadenwah Creek						
93-177	Ref. No.	North Inlet						
93-178	D1-3	Ashley River	3.0 <	2 <	3 <	3 <	1 <	3
93-179	D1-4	Ashley River	12.8	6.8 <	2	4.2	11.1 <	3
93-180	CHP3	Aquarium Site	43.2	25	33.3	20.5	36.9	304.9
93-181	E1-3	Cooper River	13.9	6.9	6.8	3.1 <	3	5.8
93-182	E1-2	Cooper River						
93-183	E3-1	Cooper River	9.1	4.4	4 <	3 <	1 <	3
93-184	E3-3	Cooper River	24.9	12.5	11.8	7.2	19.7	26.1
93-185	CHP2	Romney St.	6.2	3.4	4.3 <	3	2.7	18.4
93-186	A3-2	Leadenwah Creek	5.4	3.2	3.5 <	3	2.1 <	3
93-196	H4-2	Cooper River	16.9	10.1	10.9	5.8 <	1 <	3

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	2-METHYLNAPHTHALENE GC, ug/kg	1-METHYLNAPHTHALENE GC, ug/kg	2,6-DIMETHYLNAPHTHALENE GC, ug/kg	BIPHENYL GC, ug/kg	ACENAPHTHYLENE GC, ug/kg	ACENAPHTHENE GC, ug/kg
93-197	H4-3	Cooper River	93.2	53.5	52	30.7	70.9	118.9
93-198	H4-5	Cooper River	138.4	62.9	168	46.5	26.9	54.2
93-199	H5-2	Cooper River	31.3	17	21.2	8.2	<	3
93-200	H5-4	Cooper River					<	
93-201	H5-8	Cooper River						
93-202	H6-1	Cooper River	9.3	5.2	5.6	<	3	3.7
93-203	H6-2	Cooper River	20.3	12.3	2	<	1	9.9
93-204	H6-3	Cooper River	8.5	4.6	3.8	<	1	3
93-205	H7-1	Cooper River	11.2	6.9	2	2.9	9.3	3
93-206	H8-1	Cooper River	29.8	16.3	2	7.1	<	3
93-207	H2-2	Cooper River	17.7	8	7.3	3.9	3.2	3
93-208	H2-3	Cooper River	23.1	11.3	12.3	6.5	8	20
93-209	H2-6	Cooper River	15.8	6.8	9.9	5	7.3	12.9
93-210	CHP9	Wallace Channel	24.9	14.2	12.5	5	<	6.7
93-211	H3-1	Cooper River	32.4	14.4	11.7	10.2	34.4	39.1
93-212	H3-3	Cooper River	9.9	5.3	7	3.9	7.9	11.4
93-213	H3-5	Cooper River	42.2	26.1	20.7	19.2	20.8	114.1
93-214	F1-1	Wando River	5.1	2.9	2.8	<	3	1.6
93-215	F1-2	Wando River	14.7	7	7.9	3.4	<	3
93-216	F1-3	Wando River	5.0	2.8	2.3	<	1	3
93-217	H1-1	Cooper River	9.7	4.3	4.3	3	6.5	3
93-218	H1-4	Cooper River	8.8	4.8	4.4	<	3	15.3
93-219	H1-5	Cooper River	51.8	24.7	25.5	16.5	32.8	29.7
93-220	F2-1	Wando River	11.1	5.8	5	<	2.2	3
93-221	F2-2	Wando River	5.2	2.8	2.5	<	1	3
93-222	F2-4	Wando River	6.8	3.5	3.1	<	1	3
93-223	G1-1	Cooper River	39.3	17.2	22.8	11.8	51.4	3
93-224	G2-1	Cooper River	45.5	19.2	20.9	14.9	45.7	3
93-225	CHP1	Shipyard Creek	59.1	22.7	30.4	18.5	56	17.9

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	2-METHYLNAPHTHALENE GC, ug/kg	1-METHYLNAPHTHALENE GC, ug/kg	2,6-DIMETHYLNAPHTHALENE GC, ug/kg	BIPHENYL GC, ug/kg	ACENAPHTHYLENE GC, ug/kg	ACENAPHTHENE GC, ug/kg
93-226	CHP10	Detven's Shipyard	19.6	14.6	20.3	4.4	<	1
93-227	Ref. No.	North Inlet						10.6
93-228	B1-1	Winyah Bay	20.0	10	12.5	5.1	<	7.7
93-229	B3-1	Winyah Bay	31.2	21.6	2	3	<	3
93-230	B1-3	Winyah Bay	32.9	17.3	24.7	8.9	<	3
93-231	B1-2	Winyah Bay	34.8	16.2	20.4	6.4	<	3
93-232	CHP11	I of P Connector	10.4	5.4	4.6	3	<	3
93-233	B7-1	Winyah Bay	27.7	15.6	12.5	4.6	<	3
93-234	B6-2	Winyah Bay	6.6	3.6	4.2	3	<	3
93-235	B5-1	Winyah Bay	4.2	2.4	2	3	<	3
93-236	B4-1	Winyah Bay	8.7	4.7	4.7	3	<	3
93-237	B2-1	Winyah Bay	12.2	7.2	7.7	3	<	3
brez	ref1							
brez	ref11							
star	ref12							
star	ref12							
USC	lab ctl batch 2							
USC	lab ctl batch 3							
NIOL	Ref 1 batch 1							
NIOL	Ref 1 batch 2							
NIOL	Ref 1 batch 3							
ns	non-significant							
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.						
+	statistically significant for Dunnett's t-test at 0.05							
++	statistically significant for Dunnett's t-test at 0.01							
*	statistically significant for Mann-Whitney							
**	statistically significant for Mann-Whitney and Dunnetts							
***	statistically significant for Mann-Whitney, Dunnetts, and Distribution-free							
Duplicates								
93-165	D2-1	Ashley River						
93-165	D2-1	Ashley River						
93-166	D2-3	Ashley River						
93-166	D2-3	Ashley River						
93-167	D2-2	Ashley River						
93-167	D2-2	Ashley River						

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab.	STATION No.	Location	2,3,5-TRIME NAP GC, ug/kg	FLUORENE GC, ug/kg	PHENANTHRENE GC, ug/kg	1-METHYLPHENANTHRENE GC, ug/kg	ANTHRACENE GC, ug/kg	FLUORANTHENE GC, ug/kg	PYRENE GC, ug/kg
93-146	C1-1	Ashley River <	1 <	1	5.4 <	2	2.1	7.3	6.6
93-147	C2-4	Ashley River	1.4 <	1	3 <	2 <	2	7.5	7.1
93-148	C3-1	Ashley River <	1 <	1	3 <	2	3.6	7.9	7.5
93-149	C4-1	Ashley River <	1 <	1	2 <	2	2.5	8.2	8.2
93-150	CHP4	Plum Isl. Outfall <	1 <	1	1.8 <	2 <	2	8.5	12.9
93-151	C5-1	Ashley River	1.5	4.2	43.8	2.5	2.1	70.1	60.3
93-152	D1-1	Ashley River	1.9	19.7	67.7	10.2	64.5	364.0	893.6
93-153	A2-3	Leadenwah Creek	1.2	2.4	7.6 <	2	3.6	17.8	17.9
93-154	A3-1	Leadenwah Creek	2.8	2.5	9.3 <	2	4.3	20.6	16.5
93-155	A3-3	Leadenwah Creek	1.6	1.7	7.6 <	2	14.8	16.5	15.0
93-156	A2-7	Leadenwah Creek	1.3	2.6	12.4 <	2	10	24.3	22.1
93-157	A2-1	Leadenwah Creek	1	1.8	9.7 <	2	5.4	22.1	18.4
93-158	A1-1	Leadenwah Creek <	1	1.2	8.4 <	2	4.2	18.6	17.0
93-159	A1-2	Leadenwah Creek	2.1 <	1	6.5 <	2	2.6	13.2	15.0
93-160	CHP5	Brittle Bank Park	5	22.1	108.4	17.8	120.5	322.4	2516.9
93-161	CHP8	Brickyard Creek	1	5	17.6	2.4	17.2	81.7	87.5
93-162	D4-1	Ashley River	2.6	21.4	41.8	6.8	20.5	158.0	140.8
93-163	D4-2	Ashley River	1	3.1	15.6 <	2	13.8	47.1	71.8
93-164	D4-4	Ashley River <	3.9	106.05	413.85	39.8	1930.65	1045.4	826.0
93-165	D2-1	Ashley River	2.9	23.35	147.5	19.65	329.8	497.6	519.2
93-166	D2-3	Ashley River	11.4	59.05	226.8	59.95	477.05	843.9	986.6
93-167	D2-2	Ashley River	3.7	64.6	467.9	41.8	158	829.7	701.9
93-168	CHP6	Koppers Creosote	5	36.4	188.3	29.3	327.5	840.4	727.0
93-169	D3-3	Ashley River	1.5	6	43.2	6.8	36.5	191.5	300.5
93-170	D3-1	Ashley River	2.1	19.9	94.1	12.8	81	279.1	278.6
93-171	CHP7	Dolphin Cv Marina	4.4	41.8	213.9	21.7	261.2	447.5	431.2
93-172	D3-2	Ashley River	1	2.8	10.8 <	2	11.2	25.4	25.0
93-173	E1-1	Cooper River	2.3	8.3	33.8	3.6	26.1	70.9	53.4
93-174	E2-2	Cooper River	1.7	6.9	28.5	5.4	15.6	102.6	92.9
93-175	E2-1	Cooper River	1 <	1 <	2 <	2	2	4.0	3.9
93-176	A1-8	Leadenwah Creek <							
93-177	Ref. No.	North Inlet							
93-178	D1-3	Ashley River <	1	8.2	70.5	24.3	184.4	220.8	359.3
93-179	D1-4	Ashley River <	1	3	17.8 <	2	11.6	37.8	43.8
93-180	CHP3	Aquarium Site	11.4	178.4	761	96.8	706	1979.2	1217.2
93-181	E1-3	Cooper River	1	5.1	28.4	2.3	14.7	55.0	58.6
93-182	E1-2	Cooper River							
93-183	E3-1	Cooper River							
93-184	E3-3	Cooper River							
93-185	CHP2	Romney St.							
93-186	A3-2	Leadenwah Creek							
93-196	H4-2	Cooper River <							

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leademwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	2,3,5-TRIME NAP GC, ug/kg	FLUORENE GC, ug/kg	PHENANTHRENE GC, ug/kg	1-METHYLPHENANTHRENE GC, ug/kg	ANTHRACENE GC, ug/kg	FLUORANTHENE GC, ug/kg	PYRENE GC, ug/kg
93-197	H4-3	Cooper River	<	1	105.5	657.5	53.5	742.3	1504.4
93-198	H4-5	Cooper River	<	30	46.2	246.3	33.1	106.9	1053.5
93-199	H5-2	Cooper River	<	1 <	6.3 <	6.3	2	8.1	1319.7
93-200	H5-4	Cooper River	<						19.1
93-201	H5-8	Cooper River	1.6	3.2	11.9 <	2	5.7	34.7	1086.5
93-202	H6-1	Cooper River	1	6.3	37	4.4	51.3	150.5	37.4
93-203	H6-2	Cooper River	<	1	1.8 <	2	2	9.1	143.7
93-204	H6-3	Cooper River	<	1	1.8 <	2	2	9.1	9.0
93-205	H7-1	Cooper River	1	4.1	14.5 <	2	30.1	45.6	38.7
93-206	H8-1	Cooper River	1	10.9	46.7	2.2	72.2	131.3	119.9
93-207	H2-2	Cooper River	<	5	18.7 <	2	31.6	26.7	21.7
93-208	H2-3	Cooper River	1.2	8.5	35.4	4.6	45.6	127.4	117.8
93-209	H2-6	Cooper River	6.5	12.1	34.5 <	2	22.4	146.7	145.8
93-210	CHP9	Wallace Channel	2.4	6.5	65.5	5.9	57.3	227.3	195.9
93-211	H3-1	Cooper River	2.6	18.6	127.3	13.2	223.5	2305.1	1422.0
93-212	H3-3	Cooper River	<	1	13.4	52.6	7.7	84.9	165.7
93-213	H3-5	Cooper River	4.5	54.8	259.9	21.8	201.4	516.8	416.5
93-214	F1-1	Wando River	1.1 <	1	4.3 <	2	3.6	11.3	11.2
93-215	F1-2	Wando River	1.6	8	17.4	3.1	68.6	69.7	61.9
93-216	F1-3	Wando River	<	1 <	2 <	2 <	2 <	3.0 <	2.0
93-217	H1-1	Cooper River	1.5	5.1	22.3	2.7	27.1	54.0	42.4
93-218	H1-4	Cooper River	1.2	2.6	23.4	4	12.4	175.1	134.5
93-219	H1-5	Cooper River	<	1	58.2	26.4	378.8	443.8	353.4
93-220	F2-1	Wando River	<	1 <	2 <	2 <	2	3.8	5.0
93-221	F2-2	Wando River	<	1	3.1 <	2 <	2	6.3	5.7
93-222	F2-4	Wando River	<	1	2.7 <	2	2	13.8	13.0
93-223	G1-1	Cooper River	6.3	31.6	172.4	18.2	289.1	405.2	280.1
93-224	G2-1	Cooper River	2.2	66	261.3	24.4	558.7	454.5	335.4
93-225	CHP1	Shipyard Creek	4.5	66.7	271.8	25.4	575	499.1	344.1

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	2,3,5-TRIME NAP GC, ug/kg	FLUORENE GC, ug/kg	PHENANTHRENE GC, ug/kg	1-METHYL PHENANTHRENE GC, ug/kg	ANTHRACENE GC, ug/kg	FLUORANTHENE GC, ug/kg	PYRENE GC, ug/kg
93-226	CHP10	Detyen's Shipyard	7	11.7	120.5	36.4	16.4	191.7	193.5
93-227	Ref. No.	North Inlet							
93-228	B1-1	Winyah Bay	7.3	6.6	39.1 <	2	9.8	117.1	101.2
93-229	B3-1	Winyah Bay	<	1	2.5	16.6 <	2	24.7	35.2
93-230	B1-3	Winyah Bay		10	12	95.5 <	2	114.3	254.7
93-231	B1-2	Winyah Bay	6.9	6.5	51.7 <	2	14.1	176.1	151.1
93-232	CHP11	I of P Connector	3 <	1	4.3 <	2	2.4	10.0	7.7
93-233	B7-1	Winyah Bay	<	1	1.7	5.1 <	2	13.8	19.2
93-234	B6-2	Winyah Bay		1.8 <	1	4.1 <	2	7.3	12.0
93-235	B5-1	Winyah Bay	<	1	1.4	9.8 <	2	2.6	8.2
93-236	B4-1	Winyah Bay		1.4	3.5	10.9 <	2	32.1	34.8
93-237	B2-1	Winyah Bay	7 <	1	10.9 <	2	2	12.9	18.7
brez	ref1								
brez	ref1								
star	ref2								
star	ref2								
USC	lab ctl batch 2								
USC	lab ctl batch 3								
NIOL	Ref 1 batch 1								
NIOL	Ref 1 batch 2								
NIOL	Ref 1 batch 3								
ns	non-significant								
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.							
+	statistically significant for Dunnett's t-test at 0.05								
++	statistically significant for Dunnett's t-test at 0.01								
*	statistically significant for Mann-Whitney								
**	statistically significant for Mann-Whitney and Dunnett's								
***	statistically significant for Mann-Whitney, Dunnnett's, and Distribution-free								
Duplicates									
93-165	D2-1	Ashley River							
93-165	D2-1	Ashley River							
93-166	D2-3	Ashley River							
93-166	D2-3	Ashley River							
93-167	D2-2	Ashley River							
93-167	D2-2	Ashley River							

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	BENZ(A)ANTHRACENE GC, ug/kg	CHRYSENE GC, ug/kg	BENZO(B)FLUORANTHENE GC, ug/kg	BENZO(K)FLUORANTHENE GC, ug/kg	BENZO(E)PYRENE GC, ug/kg	BENZO(A)PYRENE GC, ug/kg
93-146	C1-1	Ashley River	<	4	2.1	3.2 <	3 <	3 <
93-147	C2-4	Ashley River	4.5	3.1	6.4 <	3	2.8	6.7
93-148	C3-1	Ashley River	<	4	3.3	3.8 <	3 <	5
93-149	C4-1	Ashley River	5.2	4.1	6.7	3.3	3.2	6.6
93-150	CHP4	Plum Isl. Outfall	4.2	4.0	7.5	3.2	3.5	5.4
93-151	C5-1	Ashley River	23.4	27.4	319.4	87	176.4	196.4
93-152	D1-1	Ashley River	182.9	190.9	209.1	62.8	110.9	153
93-153	A2-3	Leadenwah Creek	6.5	6.6	8.7	4.2	8.2	15.1
93-154	A3-1	Leadenwah Creek	9.3	9.3	12.3	4.7	7.2	14.5
93-155	A3-3	Leadenwah Creek	14.9	25.4	23.3	10.1	11.2	20.5
93-156	A2-7	Leadenwah Creek	17.2	25.7	13.9	7.8	10.9	21.5
93-157	A2-1	Leadenwah Creek	7.6	7.9	11.5	6.4	8.2	13
93-158	A1-1	Leadenwah Creek	10.2	8.8	14.3	6.1	6.2	13.8
93-159	A1-2	Leadenwah Creek	6.4	5.1	10.1	3	6.7	10.3
93-160	CHP5	Brittle Bank Park	352.8	642.6	681.7	306.6	258.4	364.3
93-161	CHP8	Brickyard Creek	26.5	34.0	45.2	16.7	28.6	38.5
93-162	D4-1	Ashley River						0
93-163	D4-2	Ashley River	28.4	26.5	41.8	15.2	38.2	56.5
93-164	D4-4	Ashley River	39.4	43.0	90.9	23.5	26.4	41.1
93-165	D2-1	Ashley River	427.65	596.5	578.1	218.8	294.45	428
93-166	D2-3	Ashley River	288.6	348.0	453.3	154.55	208.75	303.6
93-167	D2-2	Ashley River	647.15	1150.3	1170.9	423.3	525.2	710.9
93-168	CHP6	Koppers Creosote	256.7	300.3	422.5	127.9	270.3	359.1
93-169	D3-3	Ashley River	690.7	891.2	805.5	314.3	344.2	448.6
93-170	D3-1	Ashley River	75.9	134.0	205.6	59.6	131.6	167.7
93-171	CHP7	Dolphin Cv Marina	78.8	106.5	122.1	38.3	89.5	128.6
93-172	D3-2	Ashley River	175.5	282.1	271.5	88.7	193.3	258.7
93-173	E1-1	Cooper River	10.2	8.2	10.4	3.2	5.9	10.4
93-174	E2-2	Cooper River	17.2	13.0	13.0	5.1	13.7	21.5
93-175	E2-1	Cooper River	49.6	38.8	57.7	17.1	32	63.4
93-176	A1-8	Leadenwah Creek	< 4	2.2	3.7 <	3	3 <	5
93-177	Ref. No.	North Inlet						0
93-178	D1-3	Ashley River	195.3	436.6	1083.8	902.3	567.8	840.7
93-179	D1-4	Ashley River	25.8	28.8	46.3	16.9	11.7	22
93-180	CHP3	Aquarium Site	542.3	807.4	754.7	224.8	432.8	538.1
93-181	E1-3	Cooper River	36.8	28.9	68.0	25.8	21.6	52.8
93-182	E1-2	Cooper River	9.4	9.3	12.5	4.6	7.1	11.8
93-183	E3-1	Cooper River	95.9	88.6	98.6	30.6	110.4	165
93-184	E3-3	Cooper River	33.3	31.8	26.5	11.3	13.4	24.6
93-185	CHP2	Romney St.	15.1	14.0	20.7	7.8	10.8	20.1
93-186	A3-2	Leadenwah Creek	10.8	8.2	15.1	6.6	3	7.7
93-196	H4-2	Cooper River						

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	BENZ(A)ANTHRACENE GC, ug/kg	CHRYSENE GC, ug/kg	BENZO(B)FLUORANTHENE GC, ug/kg	BENZO(K)FLUORANTHENE GC, ug/kg	BENZO(E)PYRENE GC, ug/kg	BENZO(A)PYRENE GC, ug/kg
93-197	H4-3	Cooper River	669.4	561.4	458.4	215.1	133.3	0
93-198	H4-5	Cooper River	244.4	505.1	420.8	148.5	294.2	266.1
93-199	H5-2	Cooper River	18.1	14.8	23.7	9.6	7.9	276.2
93-200	H5-4	Cooper River						17.5
93-201	H5-8	Cooper River						0
93-202	H6-1	Cooper River	13.5	9.5	16.0	5	10.4	16.3
93-203	H6-2	Cooper River	82.2	71.6	94.3	30.7	34	60.9
93-204	H6-3	Cooper River	3.9	3.5	3.0	<	3	5
93-205	H7-1	Cooper River	26.5	27.0	31.2	11.2	8.1	18.6
93-206	H8-1	Cooper River	73.3	95.5	100.9	36.9	32.6	52.1
93-207	H2-2	Cooper River	10.9	10.2	8.2	3.7	3.7	7.3
93-208	H2-3	Cooper River	32	29.3	31.0	9.5	25	38.1
93-209	H2-6	Cooper River	32.3	25.2	30.3	10	42	68.2
93-210	CHP9	Wallace Channel	67.4	113.7	96.2	32.1	84.4	109.9
93-211	H3-1	Cooper River	1071.9	1491.4	685.1	297.6	283.4	368.8
93-212	H3-3	Cooper River	64.4	64.6	62.5	20.8	37.3	57.7
93-213	H3-5	Cooper River	253.8	291.7	215.1	72.8	123.2	173.8
93-214	F1-1	Wando River	6	5.3	7.7	3	3.3	7.5
93-215	F1-2	Wando River	31.3	29.2	32.9	15	19.4	37.8
93-216	F1-3	Wando River	4	< 1.0	3.0	<	3	< 5
93-217	H1-1	Cooper River	30.7	42.2	29.4	12.6	18.1	34.2
93-218	H1-4	Cooper River	44.3	87.2	80.5	26.1	31.9	41.9
93-219	H1-5	Cooper River	217.3	267.6	212.0	80.8	103.6	176
93-220	F2-1	Wando River	4	4.2	4.6	<	3	5
93-221	F2-2	Wando River	4	2.8	4.4	<	3	< 5
93-222	F2-4	Wando River	7.1	4.7	10.9	3.5	4.4	7.7
93-223	G1-1	Cooper River	201.4	244.5	161.0	76.6	97.6	157.2
93-224	G2-1	Cooper River	274	403.3	276.0	116.7	139.9	202.1
93-225	CHP1	Shipyard Creek	302	457.4	305.6	101.1	148.9	222.8

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	BENZ(A)ANTHRACENE GC, ug/kg	CHRYSENE GC, ug/kg	BENZO(B)FLUORANTHENE GC, ug/kg	BENZO(K)FLUORANTHENE GC, ug/kg	BENZO(E)PYRENE GC, ug/kg	BENZO(A)PYRENE GC, ug/kg
93-226	CHP10	Detten's Shipyard	74.5	96.1	109.2	38.5	99	126.1
93-227	Ref. No.	North Inlet						0
93-228	B1-1	Winyah Bay	33	29.5	33.5	11.3	36.7	56.9
93-229	B3-1	Winyah Bay	12.1	15.1	29.2	13.1	6.5	13.1
93-230	B1-3	Winyah Bay	80.3	121.3	87.1	32.1	105.9	136.6
93-231	B1-2	Winyah Bay	34.9	36.8	49.6	14.9	54.4	83.5
93-232	CHP11	I of P Connector	3.8	3.7	8.1	3	3.3	6.4
93-233	B7-1	Winyah Bay	4.4	5.5	8.9	3.6	5.8	9.1
93-234	B6-2	Winyah Bay	<	4	3.3	5.7	<	5
93-235	B5-1	Winyah Bay	<	4	1.9	4.4	<	5
93-236	B4-1	Winyah Bay	8.7	8.6	10.0	4.6	7.6	5
93-237	B2-1	Winyah Bay	<	4	4.5	3.2	4.6	8.8
brez	ref1							
brez	ref1							
slar	ref2							
slar	ref2							
USC	lab ctl batch 2							
USC	lab ctl batch 3							
NIOL	Ref 1 batch 1							
NIOL	Ref 1 batch 2							
NIOL	Ref 1 batch 3							
ns	non-significant							
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.						
+	statistically significant for Dunnett's t-test at 0.05							
++	statistically significant for Dunnett's t-test at 0.01							
*	statistically significant for Mann-Whitney							
**	statistically significant for Mann-Whitney and Dunnetts							
***	statistically significant for Mann-Whitney, Dunnetts, and Distribution-free							
Duplicates								
93-165	D2-1	Ashley River						
93-165	D2-1	Ashley River						
93-166	D2-3	Ashley River						
93-166	D2-3	Ashley River						
93-167	D2-2	Ashley River						
93-167	D2-2	Ashley River						

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	PERYLENE GC, ug/kg	INDENO[1,2,3-C]DIPYRENE GC, ug/kg	DIBENZ[A,H]ANTHRACENE GC, ug/kg	BENZO[G,H]PERYLENE GC, ug/kg	Total PAHs GC, ug/kg		PHENANTHRENE HPLC, ug/kg	1-METHYLPHENANTHRENE HPLC, ug/kg	ANTHRACENE HPLC, ug/kg
							GC, ug/kg	HPLC, ug/kg			
93-146	C1-1	Ashley River	3.2 <	5	<	1	<	3	35.0 <	5.7	6
93-147	C2-4	Ashley River	3.5	5.1 <	1 <	3	52.3	3	2.7	2.7	27
93-148	C3-1	Ashley River	<	3 <	3 <	3	33.7	<	5.7	2.7	8.3
93-149	C4-1	Ashley River	3.6 <	5 <	1 <	3	62.8	<	5.7	2.7	8.3
93-150	CHP4	Plum Isl. Outfall	3.3 <	5 <	1 <	3	69.9	<	5.7	2.7	8.3
93-151	C5-1	Ashley River	28.6	138.6	28.9	114.3	1445.8	23	69	53	53
93-152	D1-1	Ashley River	48.4	89.3	16.5	66.6	2635.8	23	69	10	10
93-153	A2-3	Leadenwah Creek	31.2	10.2 <	1	7.9	185.9	<	5.7	5.7	10
93-154	A3-1	Leadenwah Creek	17.5	6.5	3.9	5.8	188.0	10.7	241.9	120	20
93-155	A3-3	Leadenwah Creek	15.9	17.4 <	1	8.8	247.8	6	19	19	19
93-156	A2-7	Leadenwah Creek	25	6 <	1	8.8	205.2	<	5.7	5.7	8.3
93-157	A2-1	Leadenwah Creek	36.8	9.5 <	1	8.3	144.1	<	9	9	9
93-158	A1-1	Leadenwah Creek	7.3 <	5 <	1	8	138.3	37	106	106	106
93-159	A1-2	Leadenwah Creek	15.9	7 <	1	128.6	6334.7	6	19	19	19
93-160	CHP5	Brittle Bank Park	96.8	163.6	70.8	19.7	498.7	19.7	19	19	19
93-161	CHP8	Brickyard Creek	15.1	27.8 <	1	19.7	19.7	19.7	19	19	19
93-162	D4-1	Ashley River	19.6	35.9	6.5	24.3	798.8	20	45	45	18
93-163	D4-2	Ashley River	12.9	23.8	4.4	16.5	509.2	<	5.7	5.7	11
93-164	D4-4	Ashley River	125.2	250.45	54	198.3	7767.9	<	179	179	1339
93-165	D2-1	Ashley River	181.55	31.6	126.4	3885.0	39	163	163	392	392
93-166	D2-3	Ashley River	77.35	86.65	325.15	9079.6	97	298	298	280	280
93-167	D2-2	Ashley River	192.25	414.5	153.3	4830.5	59	542	542	147	147
93-168	CHP6	Koppers Creosote	104.1	197.6	75.3	205	6554.1	368	190	190	288
93-169	D3-3	Ashley River	138.8	245.1	22.4	98.4	1771.1	8	48	48	28
93-170	D3-1	Ashley River	47.9	131.2	96.5	76	1770.5	13	105	105	74
93-171	CHP7	Dolphin Cv Marina	37.3	97.7	36.7	147.7	3350.5	70	233	233	1261
93-172	D3-2	Ashley River	75.8	104.1	1.6	4.7	188.6	<	5.7	5.7	9
93-173	E1-1	Cooper River	6.9	9	3.4	6.5	422.7	8	39	39	22
93-174	E2-2	Cooper River	78.1	15.8	39	29.3	748.6	<	2.7	2.7	8.3
93-175	E2-1	Cooper River	83.7	39	6.3	1	<	3	33.0 <	5.7 <	8.3
93-176	A1-8	Leadenwah Creek	4.1 <	5 <	1 <	3	33.0 <	<	2.7	2.7	8.3
93-177	Ref. No.	North Inlet									
93-178	D1-3	Ashley River	207.4	438.7	96	316.5	5952.4	131	86	86	160
93-179	D1-4	Ashley River	7.4	19	6.4	10.2	365.9	45	18	18	8.3
93-180	CHP3	Aquarium Site	166.8	350.6	68.4	267.1	9634.0	207	752	752	652
93-181	E1-3	Cooper River	24.8	44.1	7.9	25.8	562.2	41	32	32	11
93-182	E1-2	Cooper River	106 <	5	1.7	5	264.3	<	5.7	5.7	9
93-183	E3-1	Cooper River	72.6	125.8	21.1	96.7	2064.2	13	181	181	68
93-184	E3-3	Cooper River	100.2	13.3	3.7	10	640.4	18	76	76	34
93-185	CHP2	Romney St.	5	3.3	9.4	9.4	221.5	<	5.7	5.7	15
93-186	A3-2	Leadenwah Creek	31.7 <	7.4	6.6 <	3.4	216.2	18	18	18	33
93-196	H4-2	Cooper River									

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	PERYLENE GC, ug/kg	INDENO(1,2,3-C,D)PYRENE GC, ug/kg	DIBENZ(A,H)ANTHRACENE GC, ug/kg	BENZO(G,H,I)PERYLENE GC, ug/kg	Total PAHs GC, ug/kg	FLUORENE HPLC, ug/kg	PHENANTHRENE HPLC, ug/kg	1-METHYLPHENANTHRENE HPLC, ug/kg	ANTHACENE HPLC, ug/kg
93-197	H4-3	Cooper River	62.4	143.7	27.3	77.8	7270.2	236	71.9	743	
93-198	H4-5	Cooper River	94.5	186.8	47.3	160.3	5842.6	44	253	120	
93-199	H5-2	Cooper River	10.9	15.3 <	1	8.1	322.4	64	8	8.3	
93-200	H5-4	Cooper River									
93-201	H5-8	Cooper River									
93-202	H6-1	Cooper River	26.6	14.1 <	1	8.5	262.9 <	5.7	15	8.3	
93-203	H6-2	Cooper River	42.8	37.8 <	1	9.3	934.4	79	43	49	
93-204	H6-3	Cooper River	4.5 <	5 <	1	3	60.0 <	5.7 <	2.7	8.3 <	
93-205	H7-1	Cooper River	14.6	13.4 <	1	7.4	340.1	55	18	28	
93-206	H8-1	Cooper River	54.8	36.4 <	1	22.2	995.8	105	51	60	
93-207	H2-2	Cooper River	3.4 <	5 <	1 <	3	205.5 <	5.7	21	28	
93-208	H2-3	Cooper River	24.9	23.4 <	1	14.5	683.1	11	40	39	
93-209	H2-6	Cooper River	80.5	39.6 <	1	33.9	823.5	19	46	21	
	CHP9	Wallace Channel	37.9	85.3	11.4	61.8	1350.7 <	5.7	77	49	
93-211	H3-1	Cooper River	107.5	148.4	51.4	116.1	8928.2				
93-212	H3-3	Cooper River	25.6	30.5	5.3	21.2	914.4	15	57	81	
93-213	H3-5	Cooper River	61.9	82.8	15.1	63.3	3173.2				
93-214	F1-1	Wando River	3.7 <	5 <	1 <	3	90.9 <	5.7	5	8.3	
93-215	F1-2	Wando River	16.5	23.3 <	1	13.1	503.1	10	21	60	
93-216	F1-3	Wando River	<	3 <	5 <	3	19.1				
93-217	H1-1	Cooper River	18.8	20.8	3.4	10.9	419.7 <	5.7	25	46	
93-218	H1-4	Cooper River	11.7	27.8	4.9	19.5	773.8 <	5.7	26	10	
93-219	H1-5	Cooper River	63.1	85.1	18.4	60.7	3039.5	61	278	433	
93-220	F2-1	Wando River	<	3 <	1 <	3	63.1 <	5.7 <	2.7	8.3 <	
93-221	F2-2	Wando River	<	3 <	5	1.1 <	3	40.5 <	5.7 <	8.3 <	
93-222	F2-4	Wando River	3.8 <	5	1.5 <	3	97.2 <	5.7	2.7	8.3	
93-223	G1-1	Cooper River	56	77.3	12.9	55.6	2535.1	37	174	264	
93-224	G2-1	Cooper River	66.6	109.5	21.7	31.4	3544.8	61	257	503	
93-225	CHP1	Shipyard Creek	73.9	112	22	77.2	3884.3	73	260	525	

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. STATION No.	Location	PERYLENE GC, ug/kg	INDENO(1,2,3-C,D)PYRENE GC, ug/kg	DIBENZ(A,H)ANTHRACENE GC, ug/kg	BENZO(G,H,I)PERYLENE GC, ug/kg	Total PAHs GC, ug/kg	FLUORENE HPLC, ug/kg	PHENANTHRENE HPLC, ug/kg	1-METHYLPHENANTHRENE HPLC, ug/kg	ANTHRACENE HPLC, ug/kg
93-226 CHP10	Deven's Shipyard	37.4	90.3	15.2	15.2	79	1431.7	<	5.7	123
93-227 Ref. No.	North Inlet									20
93-228 B1-1	Winyah Bay	109.2	47.5 <	1	19.1	30.7	751.1	9	46	25
93-229 B3-1	Winyah Bay	228	25.2			14.6	570.8	54	23	8.3
93-230 B1-3	Winyah Bay	302.1	99.3 <	1	90.2	1990.1				107
93-231 B1-2	Winyah Bay	125.6	77.6 <	1	52.8	1054.3	10	63	11	11
93-232 CHP11	I of P Connector	11.8	5.2	1.7	4.2	112.9	<	5.7	7	9
93-233 B7-1	Winyah Bay	114.2	10.2 <	1	5.3	302.4	<	5.7	9	8.3
93-234 B6-2	Winyah Bay	96.2 <	5 <	1	3	157.3	<	7	18	36
93-235 B5-1	Winyah Bay	194.4 <	167.4	11.9 <	1.1	3	271.9	<	5.7	8
93-236 B4-1	Winyah Bay				1	5	343.9	<	5.7	8
93-237 B2-1	Winyah Bay	92.7	8.6	2.2 <	3	223.7	<	5.7	13	11
brez	ref1									
star	ref2									
USC lab cfl batch 2										
USC lab cfl batch 3										
NIOL Ref 1 batch 1										
NIOL Ref 1 batch 2										
NIOL Ref 1 batch 3										
ns	non-significant									
~	Microtox Test: NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.									
+	statistically significant for Dunnnett's t-test at 0.05									
++	statistically significant for Dunnnett's t-test at 0.01									
*	statistically significant for Mann-Whitney									
**	statistically significant for Mann-Whitney and Dunnett's									
***	statistically significant for Mann-Whitney, Dunnnett's, and Distribution-free									
Duplicates										
93-165	D2-1	Ashley River								
93-166	D2-1	Ashley River								
93-166	D2-3	Ashley River								
93-166	D2-3	Ashley River								
93-167	D2-2	Ashley River								
93-167	D2-2	Ashley River								

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	FLUORANTHENE HPLC, ug/kg	PYRENE HPLC, ug/kg	BENZ(A)ANTHRACENE HPLC, ug/kg	CHRYSENE HPLC, ug/kg	BENZO(B)FLUORANTHENE HPLC, ug/kg
93-146	C1-1	Ashley River	8	7 <	4.8 <	7.3 <	8.8 <
93-147	C2-4	Ashley River	9 <	9 <	4.8 <	7.3 <	8.8 <
93-148	C3-1	Ashley River	9 <	9	6 <	7.3 <	8.8 <
93-149	C4-1	Ashley River	9 <	9	5 <	7.3 <	8.8 <
93-150	CHP4	Plum Isl. Outfall	9 <	13	5 <	7.3 <	8.8 <
93-151	C5-1	Ashley River	381	577	234	247	96
93-152	D1-1	Ashley River	21	20	10	12	16 <
93-153	A2-3	Leadenwah Creek					
93-154	A3-1	Leadenwah Creek					
93-155	A3-3	Leadenwah Creek					
93-156	A2-7	Leadenwah Creek					
93-157	A2-1	Leadenwah Creek					
93-158	A1-1	Leadenwah Creek	18	17	11	11	9 <
93-159	A1-2	Leadenwah Creek					
93-160	CHP5	Brittle Bank Park	340	1762	471	843	254
93-161	CHP8	Brickyard Creek	84	93	40	60	64
93-162	D4-1	Ashley River					
93-163	D4-2	Ashley River	192	224	75	79	61
93-164	D4-4	Ashley River	37	74	44	48	50
93-165	D2-1	Ashley River	936	875	643	787	295
93-166	D2-3	Ashley River	606	610	488	431	297
93-167	D2-2	Ashley River	803	1391	954	1783	589
93-168	CHP6	Koppers Creosote	1069	791	502	522	366
93-169	D3-3	Ashley River	1616	1821	2040	2463	641
93-170	D3-1	Ashley River	173	476	183	326	235
93-171	CHP7	Dolphin Cv Marina	301	297	133	179	134
93-172	D3-2	Ashley River	1659	1258	1040	1840	570
93-173	E1-1	Cooper River	21	23	11	9	16 <
93-174	E2-2	Cooper River	77	56	31	38 <	8.8
93-175	E2-1	Cooper River					

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	FLUORANTHENE HPLC, ug/kg	PYRENE HPLC, ug/kg	BENZ(A)ANTHRACENE HPLC, ug/kg	CHRYSENE HPLC, ug/kg	BENZO(B)FLUORANTHENE HPLC, ug/kg
93-176	A1-8	Leadenwah Creek	5.9 <	7.3 <	4.8 <	7.3	23 <
	Ref. No.	North Inlet					
93-177	D1-3	Ashley River	129	503	340	824	559
93-178	D1-4	Ashley River	18	36	16	18	12
93-179	CHP3	Aquarium Site	2554	1623	1190	1639	1035
93-180	E1-3	Cooper River	34	59	33	29	19
93-181	E1-2	Cooper River					
93-182	E3-1	Cooper River	16	24	14	17	164
93-183	E3-3	Cooper River	472	362	262	239	96
93-184	CHP2	Romney St.	103	69	34	35	25
93-185	A3-2	Leadenwah Creek	25	22	19	21	28
93-186	H4-2	Cooper River	7	14	8	<	8.8 <
93-196	H4-3	Cooper River					
93-197	H4-5	Cooper River	1659	956	614	643	154
93-198	H5-2	Cooper River	1602	981	438	781	323
93-199	H5-4	Cooper River	10	39	13	10	10
93-200	H5-8	Cooper River					
93-201	H6-1	Cooper River	29	44	19	13	9
93-202	H6-2	Cooper River	129	138	76	80	36
93-203	H6-3	Cooper River	5.9 <	7.3 <	4.8 <	7.3 <	8.8 <
93-204	H7-1	Cooper River	27	42	25	27	12
93-205	H8-1	Cooper River	78	119	69	91	54
93-206	H2-2	Cooper River	26	21	11	14 <	8.8 <
93-207	H2-3	Cooper River	137	119	52	49	42
93-208	H2-6	Cooper River	166	171	71	78	70
93-209	CHP9	Wallace Channel	227	189	96	193	119
93-210	H3-1	Cooper River					
93-211	H3-3	Cooper River	170	135	111	106	70
93-212	H3-5	Cooper River					
93-213	F1-1	Wando River	11	10	7	<	8.8 <
93-214	F1-2	Wando River	58	50	35	38	33
93-215	F1-3	Wando River					
93-216	H1-1	Cooper River	54	44	46	76	18
93-217	H1-4	Cooper River	209	100	59	114	32
93-218	H1-5	Cooper River	467	326	302	350	108
93-219	F2-1	Wando River	5.9 <	7.3 <	4.8 <	7.3 <	8.8 <
93-220	F2-2	Wando River	5.9 <	7.3 <	4.8 <	7.3 <	8.8 <
93-221	F2-4	Wando River	7 <	13	9 <	7.3 <	8.8 <
93-222	G1-1	Cooper River	389	264	289	318	109
93-223	G2-1	Cooper River	463	331	419	510	143

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	FLUORANTHENE HPLC, ug/kg	PYRENE HPLC, ug/kg	BENZ(A)ANTHRACENE HPLC, ug/kg	CHRYSENE HPLC, ug/kg	BENZO(B)FLUORANTHENE HPLC, ug/kg
93-225	CHP1	Shipyard Creek	494	337	452	559	147
93-226	CHP10	Detyens Shipyard	196	206	87	116	113
93-227	Ref. No.	North Inlet					
93-228	B1-1	Winyah Bay	114	108	52	83	39
93-229	B3-1	Winyah Bay	16	26	11	15 <	8.8 <
93-230	B1-3	Winyah Bay	360	325	167	325	172
93-231	B1-2	Winyah Bay	171	174	92	134	54
93-232	CHP11	I of P Connector	10	8 <	4.8 <	7.3	9 <
93-233	B7-1	Winyah Bay	5.9	13	6	8	16 <
93-234	B6-2	Winyah Bay	16	18 <	4.8 <	7.3	12 <
93-235	B5-1	Winyah Bay	17	16	7	8	14 <
93-236	B4-1	Winyah Bay	22	20	11	11	24
93-237	B2-1	Winyah Bay	14	15	7	10 <	8.8 <
brez	ref1						
brez	ref1						
slar	ref2						
slar	ref2						
USC lab ct	batch 2						
USC lab ct	batch 3						
NIOL Ref 1	batch 1						
NIOL Ref 1	batch 2						
NIOL Ref 1	batch 3						
ns	non-significant						
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.					

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

Duplicates

93-165	D2-1	Ashley River
93-165	D2-1	Ashley River
93-166	D2-3	Ashley River
93-166	D2-3	Ashley River
93-167	D2-2	Ashley River
93-167	D2-2	Ashley River

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	BENZO(K)FLUORANTHENE HPLC, ug/kg	BENZO(E)PYRENE HPLC, ug/kg	BENZO(A)PYRENE HPLC, ug/kg	PERYLENE HPLC, ug/kg	INDENO(1,2,3-C,D)PYRENE HPLC, ug/kg
93-146	C1-1	Ashley River	5.3	<	7.4	<	5.4
93-147	C2-4	Ashley River					v
93-148	C3-1	Ashley River	5.3	<	7.4	<	5.4
93-149	C4-1	Ashley River	5.3	<	7.4	<	5.4
93-150	CHP4	Plum Isl. Outfall	5.3		8		6
93-151	C5-1	Ashley River					v
93-152	D1-1	Ashley River	95		151	60	v
93-153	A2-3	Leadenwah Creek	5.3		12		v
93-154	A3-1	Leadenwah Creek					v
93-155	A3-3	Leadenwah Creek					v
93-156	A2-7	Leadenwah Creek					v
93-157	A2-1	Leadenwah Creek					v
93-158	A1-1	Leadenwah Creek	5.3	<	7.4	10	
93-159	A1-2	Leadenwah Creek					v
93-160	CHP5	Brittle Bank Park	282		467	159	
93-161	CHP8	Brickyard Creek	27		40	15	v
93-162	D4-1	Ashley River					v
93-163	D4-2	Ashley River	45		74	20	
93-164	D4-4	Ashley River	28		47	13	v
93-165	D2-1	Ashley River	281		618	160	
93-166	D2-3	Ashley River	215		468	98	
93-167	D2-2	Ashley River	493		1325	257	
93-168	CHP6	Koppers Creosote	297		483	161	
93-169	D3-3	Ashley River	527		898	183	
93-170	D3-1	Ashley River	136		238	54	
93-171	CHP7	Dolphin Cv Marina	103		148	33	
93-172	D3-2	Ashley River	279		461	124	
93-173	E1-1	Cooper River	5.3	<	7.4	7	v
93-174	E2-2	Cooper River	8		15	83	v
93-175	E2-1	Cooper River					

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	BENZO(K)FLUORANTHENE HPLC, ug/kg	BENZO(E)PYRENE HPLC, ug/kg	BENZO(A)PYRENE HPLC, ug/kg	PERYLENE HPLC, ug/kg	INDENO(1,2,3-C,D)PYRENE HPLC, ug/kg
93-176	A1-8	Leadenwah Creek	5.3	<	7.4	<	5.4
	Ref. No.						
93-177	Inlet.	North Inlet					
93-178	D1-3	Ashley River	524	914	236		
93-179	D1-4	Ashley River	8	16	6		
93-180	CHP3	Aquarium Site	681	885	209		
93-181	E1-3	Cooper River	17	41	25		
93-182	E1-2	Cooper River					
93-183	E3-1	Cooper River	7	12	106		
93-184	E3-3	Cooper River	136	231	72		
93-185	CHP2	Romney St.					
93-186	A3-2	Landfill	20	24	105		
93-196	H4-2	Leadenwah Creek	10	18	31		
93-197	H4-3	Cooper River	5.3	<	7.4	<	5.4
93-198	H4-5	Cooper River					
93-199	H5-2	Cooper River	125	215	74		
93-200	H5-4	Cooper River	268	260	114		
93-201	H5-8	Cooper River	7	11	12		
93-202	H6-1	Cooper River					
93-203	H6-2	Cooper River	9	16	27		
93-204	H6-3	Cooper River	23	41	41		
93-205	H7-1	Cooper River	5.3	<	7.4	<	5.4
93-206	H8-1	Cooper River	6	11	14		
93-207	H2-2	Cooper River	22	38	57		
93-208	H2-3	Cooper River	5.3	<	7.4	<	5.4
93-209	H2-6	Cooper River	23	36	32		
93-210	CHP9	Wallace Channel	37	64	104		
93-211	H3-1	Cooper River	68	106	47		
93-212	H3-3	Cooper River	45	69	29		
93-213	H3-5	Cooper River					
93-214	F1-1	Wando River					
93-215	F1-2	Wando River	5.3	<	7.4	<	5.4
93-216	F1-3	Wando River	13	23	15		
93-217	H1-1	Cooper River	18	7.4	24		
93-218	H1-4	Cooper River	29	51	13		
93-219	H1-5	Cooper River	91	163	88		
93-220	F2-1	Wando River	5.3	<	7.4	<	5.4
93-221	F2-2	Wando River	5.3	<	7.4	<	5.4
93-222	F2-4	Wando River	5.3	8	5		
93-223	G1-1	Cooper River	85	155	78		
93-224	G2-1	Cooper River	121	203	95		

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	BENZO(K)FLUORANTHENE HPLC, ug/kg	BENZO(E)PYRENE HPLC, ug/kg	BENZO(A)PYRENE HPLC, ug/kg	PERYLENE HPLC, ug/kg	INDENO(1,2,3-C,D)PYRENE HPLC, ug/kg
93-225	CHP1	Shipyard Creek	140	228	228	106	v
93-226	CHP10	Detyen's Shipyard	63	129	129	42	
93-227	Ref. No.	North Inlet					
93-228	B1-1	Winyah Bay	25	v	7.4	111	v
93-229	B3-1	Winyah Bay	5.3	v	10	208	v
93-230	B1-3	Winyah Bay	81	v	141	403	v
93-231	B1-2	Winyah Bay	44	v	111	154	v
93-232	CHP11	I of P Connector	5.3	v	7.4	11	v
93-233	B7-1	Winyah Bay	5.3	v	9	117	v
93-234	B6-2	Winyah Bay	5.3	v	7.4	228	v
93-235	B5-1	Winyah Bay	5.3	v	7.4	102	v
93-236	B4-1	Winyah Bay	6	v	7.4	7	v
93-237	B2-1	Winyah Bay	5.3	v	7.4	100	v
brez	ref1						
slar	ref2						
slar	ref2						
USC lab ct batch 2							
USC lab ct batch 3							
NIOL Ref 1 batch 1							
NIOL Ref 1 batch 2							
NIOL Ref 1 batch 3							
ns	non-significant						
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.					
+		statistically significant for Dunnett's t-test at 0.05					
++		statistically significant for Dunnett's t-test at 0.01					
*		statistically significant for Mann-Whitney and Dunnets					
**		statistically significant for Mann-Whitney, Dunnets, and Distribution-free					

Duplicates							
93-165	D2-1	Ashley River					
93-165	D2-1	Ashley River					
93-166	D2-3	Ashley River					
93-166	D2-3	Ashley River					
93-167	D2-2	Ashley River					
93-167	D2-2	Ashley River					

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	DIBENZ(A,H)ANTHRACENE HPLC, ug/kg	BENZO(G,H,I)PERYLENE HPLC, ug/kg	Total PAH HPLC, ug/kg	CL2(08) ug/kg	HEXACHLOROBENZENE ug/kg	LINDANE ug/kg	CL3(18) ug/kg
93-146	C1-1	Ashley River	7.3 <		8.2 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <
93-147	C2-4	Ashley River				< 0.13 <	0.06 <	0.08 <	0.15 <
93-148	C3-1	Ashley River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-149	C4-1	Ashley River	7.3 <	8.2 #REF!	0.13 <	0.06 <	0.08 <	0.15 <	
93-150	CHP4	Plum Isl. Outfall	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-151	C5-1	Ashley River				< 0.13 <	0.06 <	0.08 <	0.15 <
93-152	D1-1	Ashley River	7.3	67 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-153	A2-3	Leadenwah Creek	7.3	11 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-154	A3-1	Leadenwah Creek			< 0.13 <	0.06 <	0.08 <	0.15 <	
93-155	A3-3	Leadenwah Creek			< 0.13 <	0.1 <	0.08 <	0.15 <	
93-156	A2-7	Leadenwah Creek			< 0.13 <	0.06 <	0.08 <	0.15 <	
93-157	A2-1	Leadenwah Creek			< 0.38 <	0.06 <	0.08 <	0.15 <	
93-158	A1-1	Leadenwah Creek	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-159	A1-2	Leadenwah Creek			< 0.13 <	0.06 <	0.08 <	0.15 <	
93-160	CHP5	Brittle Bank Park	15	180 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-161	CHP8	Brickyard Creek	7.3	23 #REF!	0.17	0.07 <	0.08 <	0.15 <	
93-162	D4-1	Ashley River	9	39 #REF!	< 0.13 <	0.07 <	0.08 <	0.15 <	
93-163	D4-2	Ashley River	7.3	28 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-164	D4-4	Ashley River	86	385 #REF!	0.13 <	0.06 <	0.08 <	0.15 <	
93-165	D2-1	Ashley River	38	218 #REF!	< 0.13 <	0.07 <	0.08 <	0.15 <	
93-166	D2-3	Ashley River	118	603 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-167	D2-2	Ashley River							
93-168	CHP6	Koppers Creosote	62	258 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-169	D3-3	Ashley River	102	517 #REF!	0.59 <	0.06 <	0.08 <	0.15 <	
93-170	D3-1	Ashley River	31	133 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-171	CHP7	Dolphin Cv Marina	24	113 #REF!	0.13	0.44 <	0.08 <	0.15 <	
93-172	D3-2	Ashley River	50	233 #REF!	0.13	0.07 <	0.08 <	0.15 <	
93-173	E1-1	Cooper River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-174	E2-2	Cooper River	7.3	9 #REF!	< 0.13 <	0.06 <	0.08 <	0.15 <	
93-175	E2-1	Cooper River			< 0.13 <	0.06 <	0.08 <	0.15 <	

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab No.	STATION No.	Location	DIBENZ(A,H)ANTHRACENE HPLC, ug/kg	BENZO(G,H,I)PERYLENE HPLC, ug/kg	Total PAH HPLC, ug/kg	CL2(08) ug/kg	HEXYACHLOROBENZENE ug/kg	LINDANE ug/kg	CL3(18) ug/kg
93-176	A1-8	Leadenwah Creek	7.3 <		8.2 #REF!	< 0.13 <		0.06 <	0.08 < 0.15 <
93-177	Ref. No.	North Inlet							
93-178	D1-3	Ashley River	86	362 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-179	D1-4	Ashley River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-180	CHP3	Aquarium Site	119	567 #REF!	0.15	0.1 <	0.08 < 0.15 <		
93-181	E1-3	Cooper River	7.3	25 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-182	E1-2	Cooper River							
93-183	E3-1	Cooper River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-184	E3-3	Cooper River	7.3	164 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-185	CHP2	Romney St.							
93-186	A3-2	Leadenwah Creek	7.3	18 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-196	H4-2	Cooper River	7.3 <	16 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-197	H4-3	Cooper River							
93-198	H4-5	Cooper River	15	96 #REF!	0.66 <	0.06	0.51 < 0.15 <		
93-199	H5-2	Cooper River	7.3	142 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-200	H5-4	Cooper River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-201	H5-8	Cooper River							
93-202	H6-1	Cooper River	7.3	12 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-203	H6-2	Cooper River	7.3	18 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-204	H6-3	Cooper River	7.3 <	8.2 #REF!	0.21 <	0.06 <	0.08 < 0.15 <		
93-205	H7-1	Cooper River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-206	H8-1	Cooper River	7.3	23 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-207	H2-2	Cooper River	7.3 <	8.2 #REF!	0.2 <	0.06 <	0.08 < 0.15 <		
93-208	H2-3	Cooper River	7.3	21 #REF!	0.23 <	0.06 <	0.08 < 0.15 <		
93-209	H2-6	Cooper River	11	40 #REF!	0.3 <	0.06 <	0.08 < 0.15 <		
93-210	CHP9	Wallace Channel	11	81 #REF!	0.49 <	0.06 <	0.08 < 0.15 <		
93-211	H3-1	Cooper River							
93-212	H3-3	Cooper River	8	40 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-213	H3-5	Cooper River							
93-214	F1-1	Wando River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-215	F1-2	Wando River	7.3	16 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-216	F1-3	Wando River							
93-217	H1-1	Cooper River	7.3	13 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-218	H1-4	Cooper River	7.3	30 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-219	H1-5	Cooper River	7.3	69 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-220	F2-1	Wando River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-221	F2-2	Wando River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-222	F2-4	Wando River	7.3 <	8.2 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-223	G1-1	Cooper River	7.3	67 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		
93-224	G2-1	Cooper River	7.3	95 #REF!	< 0.13 <	0.06 <	0.08 < 0.15 <		

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	DIBENZ(A,H)ANTHRACENE HPLC, ug/kg	BENZO(G,H,I)PERYLENE HPLC, ug/kg	Total PAH HPLC, ug/kg	CL2(08) ug/kg	HEXACHLOROBENZENE ug/kg	LINDANE ug/kg	CL3(18) ug/kg
93-225	CHP1	Shipyard Creek	7.3	101	#REF!	<	0.06 <	0.08 <	0.15 <
93-226	CHP10	Detyen's Shipyard Ref. No.	13	94	#REF!	<	0.13 <	0.06 <	0.08 < 0.15 <
93-227	Inlet.	North Inlet							
93-228	B1-1	Winyah Bay	7.3	31	#REF!	<	0.13	0.09	0.23 < 0.15 <
93-229	B3-1	Winyah Bay	7.3	10	#REF!	<	0.13 <	0.06 <	0.08 < 0.15 <
93-230	B1-3	Winyah Bay	13	121	#REF!	0.38	0.24	0.86 <	0.15 <
93-231	B1-2	Winyah Bay	9	90	#REF!	<	0.13	0.22 <	0.08 < 0.15 <
93-232	CHP11	I of P Connector	7.3 <	8.2	#REF!	<	0.13 <	0.06 <	0.08 < 0.15 <
93-233	B7-1	Winyah Bay	7.3 <	8.2	#REF!	<	0.13	0.2 <	0.08 < 0.15 <
93-234	B6-2	Winyah Bay	7.3 <	8.2	#REF!	<	0.13 <	0.06 <	0.08 < 0.15 <
93-235	B5-1	Winyah Bay	7.3 <	8.2	#REF!	<	0.13 <	0.06 <	0.08 < 0.15 <
93-236	B4-1	Winyah Bay	7.3 <	8.2	#REF!	<	0.13 <	0.06 <	0.08 < 0.15 <
93-237	B2-1	Winyah Bay	7.3 <	8.2	#REF!	<	0.13	0.08 <	0.08 < 0.15 <
biez	ref1								
biez	ref1								
star	ref2								
USC lab ct	batch 2								
USC lab ct	batch 3								
NIOL Ref 1	batch 1								
NIOL Ref 1	batch 2								
NIOL Ref 1	batch 3								
ns	non-significant								
^	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.							
+		statistically significant for Dunnett's t-test at 0.05							
++		statistically significant for Dunnett's t-test at 0.01							
*		statistically significant for Mann-Whitney							
**		statistically significant for Mann-Whitney and Dunnets							
***		statistically significant for Mann-Whitney, Dunnets, and Distribution-free							
Duplicates									
93-165	D2-1	Ashley River							
93-165	D2-1	Ashley River							
93-166	D2-3	Ashley River							
93-166	D2-3	Ashley River							
93-167	D2-2	Ashley River							
93-167	D2-2	Ashley River							

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL3(28) ug/kg	HEPTACHLOR ug/kg	CL4(52) ug/kg	ALDRIN ug/kg	CL4(44) ug/kg	HEPTACHLOREPOXIDE ug/kg	CL4(66) ug/kg	2,4-DDE ug/kg	CL5(101) ug/kg
93-146	C1-1	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-147	C2-4	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-148	C3-1	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-149	C4-1	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-150	CHP4	Plum Isl. Outfall	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-151	C5-1	Ashley River	0.19 <	0.04 <	0.08 <	0.01 <	0.05 <	0.10 <	0.07 <	0.06 <	0.02 <
93-152	D1-1	Ashley River	0.19 <	0.04 <	0.17 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.1 <
93-153	A2-3	Leadenwah Creek	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-154	A3-1	Leadenwah Creek	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-155	A3-3	Leadenwah Creek	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-156	A2-7	Leadenwah Creek	0.19 <	0.04 <	0.09 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-157	A2-1	Leadenwah Creek	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-158	A1-1	Leadenwah Creek	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-159	A1-2	Leadenwah Creek	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-160	CHP5	Brittle Bank Park	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-161	CHP8	Brickyard Creek	0.19 <	0.04 <	0.09 <	0.01 <	0.05 <	0.10 <	0.07 <	0.06 <	0.23
93-162	D4-1	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-163	D4-2	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.08 <	0.06 <	0.14 <
93-164	D4-4	Ashley River	0.19 <	0.04 <	0.07 <	0.06 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-165	D2-1	Ashley River	0.19 <	0.04 <	0.19 <	0.01 <	0.05 <	0.10 <	0.26 <	0.06 <	0.02 <
93-166	D2-3	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.07 <	0.06 <	0.15 <
93-167	D2-2	Ashley River	0.19 <	0.04 <	0.1 <	0.01 <	0.05 <	0.10 <	0.13 <	0.06 <	0.3
93-168	CHP6	Koppers Creosote	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.18 <
93-169	D3-3	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-170	D3-1	Ashley River	0.19 <	0.04 <	0.16	0.07 <	0.05 <	0.10 <	0.1 <	0.06 <	0.5
93-171	CHP7	Dolphin Cv Marina	0.19 <	0.04 <	0.14	0.08 <	0.05 <	0.10 <	0.06 <	0.06 <	0.66
93-172	D3-2	Ashley River	0.19 <	0.04 <	0.23 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.55
93-173	E1-1	Cooper River	0.19 <	0.04 <	0.12 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.15 <
93-174	E2-2	Cooper River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.05 <
93-175	E2-1	Cooper River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL3(28) ug/kg	HEPTACHLOR ug/kg	CL4(52) ug/kg	ALDRIN ug/kg	CL4(44) ug/kg	HEPTACHLOREPOXIDE ug/kg	CL4(66) ug/kg	2,4-DDE ug/kg	CL5(101) ug/kg
93-176	A1-8 Ref. No.	Leadenwah Creek	0.19 <	0.04 <	0.07	0.07 <	0.05	<	0.10 <	0.06 <	0.06
93-177	Inlet.	North Inlet									
93-178	D1-3	Ashley River	0.19 <	0.04	0.42 <	0.01 <	0.05 <	0.10	0.17 <	0.06	0.29 <
93-179	D1-4	Ashley River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-180	CHP3	Aquarium Site	0.19 <	0.04	0.21	0.21 <	0.05 <	0.10 <	0.06 <	0.06	0.3
93-181	E1-3	Cooper River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-182	E1-2	Cooper River									
93-183	E3-1	Cooper River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-184	E3-3	Cooper River	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-185	CHP2	Romney St. Landfill	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-186	A3-2	Leadenwah Creek	0.19 <	0.04 <	0.07 <	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-196	H4-2	Cooper River									
93-197	H4-3	Cooper River									
93-198	H4-5	Cooper River	0.19 <	0.04	<	0.01 <	0.41 <	0.10	0.97 <	0.06	17.69 <
93-199	H5-2	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.07 <
93-200	H5-4	Cooper River									
93-201	H5-8	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-202	H6-1	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-203	H6-2	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-204	H6-3	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-205	H7-1	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-206	H8-1	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-207	H2-2	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-208	H2-3	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.08 <
93-209	H2-6	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-210	CHP9	Wallace Channel	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-211	H3-1	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.22 <
93-212	H3-3	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.1
93-213	H3-5	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.25
93-214	F1-1	Wando River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-215	F1-2	Wando River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-216	F1-3	Wando River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-217	H1-1	Cooper River	0.19 <	0.04	<	0.01 <	0.06 <	0.10 <	0.14 <	0.06	0.1 <
93-218	H1-4	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-219	H1-5	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-220	F2-1	Wando River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-221	F2-2	Wando River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-222	F2-4	Wando River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.04 <
93-223	G1-1	Cooper River	0.29 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.02 <
93-224	G2-1	Cooper River	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.17

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL3(28) ug/kg	HEPTACHLOR ug/kg	CL4(52) ug/kg	ALDRIN ug/kg	CL4(44) ug/kg	HEPTACHLOREPOXIDE ug/kg	CL4(66) ug/kg	2,4-DDE ug/kg	CL5(101) ug/kg
93-225	CHP1	Shipyard Creek	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06	0.16 <
93-226	CHP10	Detyen's Shipyard	0.19 <	0.04	<	0.01	0.18 <	0.10	0.33 <	0.06	0.8
93-227	Ref. Inlet.	North Inlet									
93-228	B1-1	Winyah Bay	0.19 <	0.04	<	0.01	0.16 <	0.10	1.18 <	0.06	0.42
93-229	B3-1	Winyah Bay	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-230	B1-3	Winyah Bay	0.24 <	0.04	<	0.01	0.44 <	0.10	2.11 <	0.06	1.84
93-231	B1-2	Winyah Bay	0.19 <	0.04	<	0.01	0.49 <	0.10 <	0.06 <	0.06 <	0.02 <
93-232	CHP11	I of P Connector	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-233	B7-1	Winyah Bay	0.19 <	0.04	<	0.01 <	0.05 <	0.10	0.31 <	0.06 <	0.02 <
93-234	B6-2	Winyah Bay	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-235	B5-1	Winyah Bay	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
93-236	B4-1	Winyah Bay	0.19 <	0.04	<	0.01 <	0.05 <	0.10	0.49 <	0.06 <	0.02 <
93-237	B2-1	Winyah Bay	0.19 <	0.04	<	0.01 <	0.05 <	0.10 <	0.06 <	0.06 <	0.02 <
brez	ref1										
star	ref1										
star	ref2										
star	ref2										
USC lab ct batch 2											
USC lab ct batch 3											
NIOL Ref 1 batch 1											
NIOL Ref 1 batch 2											
NIOL Ref 1 batch 3											
ns	non-significant										
Λ	Microtox Tests:	NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.									
+		statistically significant for Dunnett's t-test at 0.05									
++		statistically significant for Dunnett's t-test at 0.01									
*		statistically significant for Mann-Whitney									
**		statistically significant for Mann-Whitney and Dunnett's									
***		statistically significant for Mann-Whitney, Dunnett's, and Distribution-free									
Duplicates											
93-165	D2-1	Ashley River									
93-165	D2-1	Ashley River									
93-166	D2-3	Ashley River									
93-166	D2-3	Ashley River									
93-167	D2-2	Ashley River									
93-167	D2-2	Ashley River									

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab.	STATION No.	Location	alpha-Chlordane ug/kg	TRANS-NONACHLOR ug/kg	DIELDRIN ug/kg	4,4-DDE ug/kg	CL4(77) ug/kg	2,4-DDD ug/kg	CL5(118) ug/kg	4,4-DDT ug/kg	2,4-DDT ug/kg
93-146	C1-1	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.24 <	0.14
93-147	C2-4	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.24 <	0.14
93-148	C3-1	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.24 <	0.14
93-149	C4-1	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.24 <	0.14
93-150	CHP4	Plum Isl. Outfall	0.08 <	0.09 <	0.18 <	0.05 <	1.5 <	0.06 <	0.06 <	0.24 <	0.14
93-151	C5-1	Ashley River	0.11 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.24 <	0.14
93-152	D1-1	Ashley River	0.08 <	0.09 <	0.18 <	0.24 <	1.5 <	0.06 <	0.06 <	0.07 <	0.55 <
93-153	A2-3	Leadenwah Creek	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-154	A3-1	Leadenwah Creek	0.08 <	0.09 <	0.18 <	0.3 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-155	A3-3	Leadenwah Creek	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-156	A2-7	Leadenwah Creek	0.08 <	0.09 <	0.18 <	0.67 <	1.5 <	0.06 <	0.17 <	0.07 <	0.24 <
93-157	A2-1	Leadenwah Creek	0.08 <	0.09 <	0.18 <	0.71 <	1.5 <	0.06 <	0.14 <	0.07 <	0.24 <
93-158	A1-1	Leadenwah Creek	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-159	A1-2	Leadenwah Creek	0.08 <	0.09 <	0.18 <	0.73 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-160	CHP5	Brittle Bank Park	0.08 <	0.09 <	0.18 <	0.15 <	1.5 <	0.06 <	0.94 <	0.07 <	0.24 <
93-161	CHP8	Brickyard Creek	0.13 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.11	0.37 <
93-162	D4-1	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-163	D4-2	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-164	D4-4	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-165	D2-1	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.06 <	0.3
93-166	D2-3	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-167	D2-2	Ashley River	0.09 <	0.11 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.14	0.73 <
93-168	CHP6	Koppers Creosote	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.09 <	0.24 <
93-169	D3-3	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	1.83 <	0.07 <	0.24 <
93-170	D3-1	Ashley River	0.15 <	0.09 <	0.18 <	0.82 <	1.5 <	0.06 <	0.06 <	0.21 <	0.24 <
93-171	CHP7	Dolphin Cv Marina	0.28	0.2	0.18	0.89 <	1.5 <	0.06 <	0.06 <	0.17	0.96 <
93-172	D3-2	Ashley River	0.36	0.27 <	0.18	1.01 <	1.5	1.32	0.35	0.52 <	0.14
93-173	E1-1	Cooper River	0.08 <	0.09 <	0.18 <	0.18	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-174	E2-2	Cooper River	0.08 <	0.09 <	0.18 <	0.11 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <
93-175	E2-1	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab.	STATION No.	Location	alpha-Chlordane ug/kg	TRANS-NONACHLOR ug/kg	DIELDRIN ug/kg	4,4-DDE ug/kg	CL4(77) ug/kg	2,4-DDD ug/kg	C16(154) ug/kg	CL5(118) ug/kg	4,4-DDD ug/kg	2,4-DDT ug/kg
93-176	A1-8	Leadenwah Creek	0.11 <	0.09 <	0.18	1.38 <	1.5 <	0.06	0.13 <	0.07	0.81 <	0.14
93-177	Ref. No.	North Inlet										
93-178	D1-3	Ashley River	0.08 <	0.09 <	0.18	0.81 <	1.5 <	0.06	1.77	0.25 <	0.24 <	0.14
93-179	D1-4	Ashley River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-180	CHP3	Aquarium Site	0.25 <	0.09 <	0.18	0.49 <	1.5 <	0.06	0.56 <	0.07 <	0.24 <	0.14
93-181	E1-3	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-182	E1-2	Cooper River										
93-183	E3-1	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-184	E3-3	Cooper River	0.08 <	0.09 <	0.18	0.07 <	1.5 <	0.06	0.33 <	0.07 <	0.24 <	0.14
93-185	CHP2	Romney St.	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-186	A3-2	Leadenwah Creek	0.08 <	0.09 <	0.18	0.54 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-196	H4-2	Cooper River										
93-197	H4-3	Cooper River										
93-198	H4-5	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06	0.06 <	0.07 <	0.24 <	0.14
93-199	H5-2	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-200	H5-4	Cooper River										
93-201	H5-8	Cooper River										
93-202	H6-1	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-203	H6-2	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-204	H6-3	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-205	H7-1	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-206	H8-1	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-207	H2-2	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-208	H2-3	Cooper River	0.08 <	0.09 <	0.18	0.12 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-209	H2-6	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-210	CHP9	Wallace Channel	0.27 <	0.09 <	0.18	0.93 <	1.5 <	0.06 <	0.1	0.07 <	0.24 <	0.14
93-211	H3-1	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-212	H3-3	Cooper River	0.13 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-213	H3-5	Cooper River	0.09 <	0.09 <	0.18 <	0.24 <	1.5 <	0.06 <	0.24	0.15	0.45 <	0.14
93-214	F1-1	Wando River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-215	F1-2	Wando River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-216	F1-3	Wando River	0.08 <	0.09 <	0.18	0.06 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-217	H1-1	Cooper River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-218	H1-4	Cooper River	0.08 <	0.09 <	0.18 <	0.05 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-219	H1-5	Cooper River	0.08 <	0.09 <	0.18	0.25 <	1.5 <	0.06	2.35 <	0.07 <	0.24 <	0.14
93-220	F2-1	Wando River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-221	F2-2	Wando River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-222	F2-4	Wando River	0.08 <	0.09 <	0.18 <	0.03 <	1.5 <	0.06 <	0.06 <	0.07 <	0.24 <	0.14
93-223	G1-1	Cooper River	0.08 <	0.09 <	0.18	0.28 <	1.5 <	0.06	0.32 <	0.07 <	0.24 <	0.14
93-224	G2-1	Cooper River	0.16 <	0.09 <	0.18	0.4 <	1.5 <	0.06	0.47 <	0.07	0.28 <	0.14

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

+ statistically significant for Dunnett's t-test at 0.05

卷之三

ns non-significant

pared to reference 177. NMES 93-196 to 93-237 compared to reference 227.

***	statistically significant for Mann-Whitney and Dunnett's
****	statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL6(153) ug/kg	CL5(105) ug/kg	4,4-DDT ug/kg	CL6(138) ug/kg	CL5(126) ug/kg	CL7(187) ug/kg	CL6(128) ug/kg	CL7(180) ug/kg	MIREX ug/kg	CL7(170) ug/kg
93-146	C1-1	Ashley River <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-147	C2-4	Ashley River <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.08 <	0.07 <	0.11 <	0.16 <	0.16 <
93-148	C3-1	Ashley River <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-149	C4-1	Ashley River <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-150	CHP4	Plum Isl. Outfall <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.55 <
93-151	C5-1	Ashley River <	0.17 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.61 <
93-152	D1-1	Ashley River <	0.20 <	0.12 <	0.06 <	0.18 <	0.13 <	0.05 <	0.07 <	0.21 <	0.16 <	1.73 <
93-153	A2-3	Leadenwah Creek <	0.10 <	0.12 <	0.32 <	0.32 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.28 <
93-154	A3-1	Leadenwah Creek <	0.10 <	0.12 <	0.64 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.18 <
93-155	A3-3	Leadenwah Creek <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-156	A2-7	Leadenwah Creek <	0.10 <	0.12 <	0.42 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.21 <
93-157	A2-1	Leadenwah Creek <	0.10 <	0.12 <	0.41 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.17 <
93-158	A1-1	Leadenwah Creek <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-159	A1-2	Leadenwah Creek <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-160	CHP5	Brittle Bank Park <	0.33 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-161	CHP8	Brickyard Creek <	0.38 <	0.12 <	0.1 <	0.4 <	0.13 <	0.11 <	0.07 <	0.22 <	0.16 <	0.16 <
93-162	D4-1	Ashley River <										
93-163	D4-2	Ashley River <	0.29 <	0.12 <	0.09	0.25 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-164	D4-4	Ashley River <	0.22 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-165	D2-1	Ashley River <	1.06 <	0.12 <	0.02 <	0.18 <	0.13 <	0.18 <	0.07 <	0.11 <	0.16 <	0.16 <
93-166	D2-3	Ashley River <	0.18 <	0.12 <	0.07 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-167	D2-2	Ashley River <	0.10 <	0.12 <	0.02 <	0.18 <	0.31	0.15 <	0.07 <	0.11 <	0.16 <	0.16 <
93-168	CHP6	Koppers Creosote <	0.35 <	0.12 <	0.02 <	0.18 <	0.13 <	0.11 <	0.07 <	0.11 <	0.16 <	0.16 <
93-169	D3-3	Ashley River <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.2 <	0.16 <
93-170	D3-1	Ashley River <	0.10 <	0.12 <	0.02 <	0.42 <	0.13	0.17 <	0.07 <	0.3 <	0.16 <	0.16 <
93-171	CHP7	Dolphin Cv Marina <	0.10 <	0.12 <	0.31	0.58 <	0.13	0.05 <	0.07 <	0.28 <	0.16 <	0.17 <
93-172	D3-2	Ashley River <	0.10 <	0.12 <	0.02	0.77 <	0.13 <	0.05 <	0.07 <	0.31 <	0.16 <	0.16 <
93-173	E1-1	Cooper River <	0.10 <	0.12 <	0.02 <	0.18 <	0.13	0.06 <	0.07 <	0.11 <	0.16 <	0.16 <
93-174	E2-2	Cooper River <	0.12 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-175	E2-1	Cooper River <	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.24 <

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL6(153) ug/kg	CL5(105) ug/kg	4,4-DDT ug/kg	CL6(138) ug/kg	CL5(126) ug/kg	CL7(187) ug/kg	CL6(128) ug/kg	CL7(180) ug/kg	MIREX ug/kg	CL7(170) ug/kg
93-176	A1-8	Leadenwah Creek	< 0.10 <	0.12	0.59	0.52 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-177	Ref. No.	North Inlet										
93-178	D1-3	Ashley River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	4.39 <
93-179	D1-4	Ashley River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-180	CHP3	Aquarium Site	< 0.10 <	0.12 <	0.02 <	0.77 <	0.13 <	0.05 <	0.07 <	0.33 <	0.16 <	0.16 <
93-181	E1-3	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-182	E1-2	Cooper River										
93-183	E3-1	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-184	E3-3	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-185	CHP2	Romney St. Landfill	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.21 <
93-186	A3-2	Leadenwah Creek	< 0.10 <	0.12 <	0.02 <	0.31 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.17 <
93-196	H4-2	Cooper River										
93-197	H4-3	Cooper River										
93-198	H4-5	Cooper River										
93-199	H5-2	Cooper River	50.79 <	0.12 <	0.02 <	0.18 <	9.1	21.48 <	0.07	50.7 <	0.16 <	0.16 <
93-200	H5-4	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-201	H5-8	Cooper River										
93-202	H6-1	Cooper River	0.19 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-203	H6-2	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.31 <
93-204	H6-3	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-205	H7-1	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-206	H8-1	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-207	H2-2	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-208	H2-3	Cooper River	0.25 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-209	H2-6	Cooper River	0.38 <	0.12 <	0.02 <	0.18 <	0.13 <	0.21 <	0.07 <	0.11 <	0.16 <	0.16 <
93-210	CHP9	Wallace Channel	0.23 <	0.12	0.53 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-211	H3-1	Cooper River	0.15 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-212	H3-3	Cooper River	0.30 <	0.12 <	0.02 <	0.19 <	0.13 <	0.05 <	0.07 <	0.14 <	0.16 <	0.9 <
93-213	H3-5	Cooper River	0.23 <	0.12	1.32 <	0.18 <	0.13 <	0.09 <	0.07 <	0.11 <	0.16 <	0.16 <
93-214	F1-1	Wando River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-215	F1-2	Wando River	< 0.10 <	0.12	0.79	0.78 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.34 <
93-216	F1-3	Wando River	0.19 <	0.12	0.08 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.19 <
93-217	H1-1	Cooper River	0.26 <	0.12 <	0.02 <	0.25 <	0.13 <	0.06 <	0.07 <	0.11 <	0.16 <	0.39 <
93-218	H1-4	Cooper River	< 0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-219	H1-5	Cooper River	0.52 <	0.12 <	0.02 <	0.31 <	0.13 <	0.1	0.07 <	0.11 <	0.16 <	2.06 <
93-220	F2-1	Wando River	< 0.10 <	0.12	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-221	F2-2	Wando River	< 0.10 <	0.12	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-222	F2-4	Wando River	0.11 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-223	G1-1	Cooper River	0.56 <	0.12 <	0.02 <	0.4 <	0.13	0.08 <	0.07 <	0.11 <	0.16 <	1.91 <
93-224	G2-1	Cooper River	0.48 <	0.12	0.58 <	0.13 <	0.05 <	0.07 <	0.07 <	0.11 <	0.16 <	2.34 <

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL6(153) ug/kg	CL5(105) ug/kg	4,4-DDT ug/kg	CL6(138) ug/kg	CL5(126) ug/kg	CL7(187) ug/kg	CL6(128) ug/kg	CL7(180) ug/kg	MIREX ug/kg	CL7(170) ug/kg
93-225	CHP1	Shipyard Creek	0.53 <	0.12 <	0.02	0.48 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16	3.01 <
93-226	CHP10	Detyen's Shipyard	0.23 <	0.12 <	0.02	0.97 <	0.13 <	0.05 <	0.07	0.21 <	0.16 <	0.16 <
93-227	Ref. No.	North Inlet										
93-228	B1-1	Winyah Bay	<	0.10 <	0.12	0.8 <	0.18 <	0.13 <	0.05 <	0.07	0.16	0.27 <
93-229	B3-1	Winyah Bay	0.74 <	0.12	0.63 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <	0.16 <
93-230	B1-3	Winyah Bay	<	0.10 <	0.12	2.09	2.09	1.72 <	0.05 <	0.07	1.03 <	0.16 <
93-231	B1-2	Winyah Bay	1.16 <	0.12	0.98 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16	0.57
93-232	CHP11	I of P Connector	<	0.10 <	0.12 <	0.02 <	0.18 <	0.13 <	0.05 <	0.07 <	0.11 <	0.16 <
93-233	B7-1	Winyah Bay	0.60 <	0.12	0.5 <	0.18	0.58 <	0.05 <	0.07	0.11 <	0.16 <	0.16 <
93-234	B6-2	Winyah Bay	<	0.10 <	0.12	0.3 <	0.18	0.2 <	0.05 <	0.07	0.11 <	0.16 <
93-235	B5-1	Winyah Bay	<	0.10 <	0.12 <	0.02 <	0.18	0.21 <	0.05 <	0.07	0.11 <	0.16 <
93-236	B4-1	Winyah Bay	<	0.10 <	0.12	0.37	0.37 <	0.13 <	0.05 <	0.07	0.33 <	0.16 <
93-237	B2-1	Winyah Bay	<	0.10 <	0.12	0.27 <	0.18 <	0.13 <	0.05 <	0.07	0.11 <	0.16 <
brez	ref1											
star	ref2											
star	ref2											
USC lab ct batch 2												
USC lab ct batch 3												
NIOL Ref 1 batch 1												
NIOL Ref 1 batch 2												
NIOL Ref 1 batch 3												

ns non-significant
 ^ Microtox Tests: NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

Duplicates

93-165	D2-1	Ashley River
93-165	D2-1	Ashley River
93-166	D2-3	Ashley River
93-166	D2-3	Ashley River
93-167	D2-2	Ashley River
93-167	D2-2	Ashley River

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL8(195) ug/kg	CL9(206) ug/kg	CL10(209) ug/kg	Sum of 20 Congeners ug/kg	2x	total PCBs ug/kg	Pesticide Total : ug/kg	DDT Total :
93-146	C1-1	Ashley River	0.12 <	0.10 <	0.10	0.00	2	0	0	0.80
93-147	C2-4	Ashley River	0.12 <	0.10 <	0.10	0.19	2	0.385530709	0.80	0.55
93-148	C3-1	Ashley River	0.12 <	0.10 <	0.10	0.00	2	0	0.80	0.55
93-149	C4-1	Ashley River	0.12 <	0.10 <	0.10	1.03	2	2.052954758	0.80	0.55
93-150	CHP4	Plum Isl. Outfall	0.12 <	0.10 <	0.10	1.60	2	3.197182663	0.80	0.57
93-151	C5-1	Ashley River	0.12 <	0.10 <	0.10	1.04	2	2.078493554	0.83	0.55
93-152	D1-1	Ashley River	0.12 <	0.10 <	0.10	3.27	2	6.533081511	0.80	1.11
93-153	A2-3	Leadenwah Creek	0.12 <	0.10 <	0.10	0.92	2	1.835076923	0.80	0.85
93-154	A3-1	Leadenwah Creek	0.12 <	0.10 <	0.10	1.11	2	2.222102767	0.80	1.44
93-155	A3-3	Leadenwah Creek	0.12 <	0.10 <	0.10	0.10	2	0.195778656	0.84	0.55
93-156	A2-7	Leadenwah Creek	0.12 <	0.10 <	0.10	1.55	2	3.103564626	0.80	1.59
93-157	A2-1	Leadenwah Creek	0.12 <	0.10 <	0.10	1.99	2	3.977926531	0.80	1.62
93-158	A1-1	Leadenwah Creek	0.12 <	0.10 <	0.10	0.00	2	0	0	0.80
93-159	A1-2	Leadenwah Creek	0.12 <	0.10 <	0.10	0.73	2	1.460291262	0.80	1.25
93-160	CHP5	Brittle Bank Park	0.12	0.11	0.12	1.65	2	3.303091892	0.80	0.67
93-161	CHP8	Brickyard Creek	0.12 <	0.10 <	0.10	2.44	2	4.881875	0.86	0.76
93-162	D4-1	Ashley River								
93-163	D4-2	Ashley River	0.12 <	0.10 <	0.10	1.01	2	2.017217235	0.81	0.62
93-164	D4-4	Ashley River	0.12 <	0.10 <	0.10	0.28	2	0.554591549	0.85	0.55
93-165	D2-1	Ashley River	0.14	0.25 <	0.10	4.13	2	8.252705882	0.80	2.06
93-166	D2-3	Ashley River	0.12 <	0.10 <	0.10	0.61	2	1.216055556	0.81	0.6
93-167	D2-2	Ashley River	0.12 <	0.10 <	0.10	2.07	2	4.136666667	0.83	1.04
93-168	CHP6	Koppers Creosote	0.12 <	0.10 <	0.10	0.74	2	1.473536878	0.80	0.55
93-169	D3-3	Ashley River	0.18	0.19	0.65	3.64	2	7.277577465	0.84	0.55
93-170	D3-1	Ashley River	0.12 <	0.10 <	0.10	2.91	2	5.820275689	0.93	1.34
93-171	CHP7	Dolphin Cv Marina	0.12 <	0.10 <	0.10	5.21	2	10.41151055	1.56	2.42
93-172	D3-2	Ashley River	0.22 <	0.10 <	0.10	7.30	2	14.59800709	1.27	3.07
93-173	E1-1	Cooper River	0.12 <	0.10 <	0.10	0.52	2	1.048682779	0.80	0.72
93-174	E2-2	Cooper River	0.12 <	0.10 <	0.10	0.27	2	0.545277162	0.80	0.63
93-175	E2-1	Cooper River	0.12 <	0.10 <	0.10	0.24	2	0.47342711	0.80	0.55

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL8(195) ug/kg	CL9(206) ug/kg	CL10(209) ug/kg	Sum of 20 Congeners ug/kg	2x	total PCBs ug/kg	Pesticide Total : ug/kg	DDT Total :
93-176	A1-8	Leadenwah Creek	0.12 <	0.10	0.10	0.10	3.76	2	7.522648118	0.89
93-177	Ref. Inlet.	North Inlet								
93-178	D1-3	Ashley River	0.12 <	0.10 <	0.10	0.10	8.09	2	16.17438571	0.80
93-179	D1-4	Ashley River	0.12 <	0.10 <	0.10	0.10	0.00	0	0.80	0.55
93-180	CHP3	Aquarium Site	0.12 <	0.10 <	0.10	0.10	3.38	2	6.759096774	1.21
93-181	E1-3	Cooper River	0.12 <	0.10 <	0.10	0.10	0.00	0	0.80	0.55
93-182	E1-2	Cooper River								0
93-183	E3-1	Cooper River	0.12 <	0.10 <	0.10	0.10	0.00	0	0.80	0.55
93-184	E3-3	Cooper River	0.12 <	0.10 <	0.10	0.10	0.40	2	0.809868263	0.80
93-185	CHP2	Romney St.	0.12 <	0.10 <	0.10	0.10	0.21	2	0.412982456	0.80
93-186	A3-2	Landfill								0.55
93-196	H4-2	Leadenwah Creek	0.12 <	0.10 <	0.10	0.10	0.17	2	0.336735376	0.80
93-197	H4-3	Cooper River								1.06
93-198	H4-5	Cooper River	4.73	1.65	0.11	0.11	164.35	2	328.703444	3.51
93-199	H5-2	Cooper River	0.12 <	0.10 <	0.10	0.10	0.07	2	0.145799012	0.80
93-200	H5-4	Cooper River								0.55
93-201	H5-8	Cooper River								0.55
93-202	H6-1	Cooper River	0.12 <	0.10 <	0.10	0.10	0.19	2	0.37703125	0.80
93-203	H6-2	Cooper River	0.12 <	0.10 <	0.10	0.10	0.43	2	0.866367123	0.80
93-204	H6-3	Cooper River	0.12 <	0.10 <	0.10	0.10	0.21	2	0.41233281	0.80
93-205	H7-1	Cooper River	0.12 <	0.10 <	0.10	0.10	0.00	0	0.80	0.55
93-206	H8-1	Cooper River	0.12 <	0.10 <	0.10	0.10	0.00	0	0.80	0.55
93-207	H2-2	Cooper River	0.12 <	0.10 <	0.10	0.10	0.20	2	0.392996923	0.80
93-208	H2-3	Cooper River	0.12 <	0.10 <	0.10	0.10	0.69	2	1.380666667	0.80
93-209	H2-6	Cooper River	0.12	0.25	0.56	0.56	1.70	2	3.395779614	0.80
93-210	CHP9	Wallace Channel	0.12 <	0.10	0.10	0.10	3.20	2	6.397879518	0.99
93-211	H3-1	Cooper River	0.12 <	0.10	0.14	0.14	0.78	2	1.563707547	0.80
93-212	H3-3	Cooper River	0.12 <	0.10 <	0.10	0.10	1.76	2	3.525135593	0.85
93-213	H3-5	Cooper River	0.12 <	0.10 <	0.10	0.10	3.33	2	6.651623762	0.82
93-214	F1-1	Wando River	0.12 <	0.10 <	0.10	0.10	0.00	2	0	0.80
93-215	F1-2	Wando River	0.12 <	0.10 <	0.10	0.10	1.92	2	3.837478992	0.80
93-216	F1-3	Wando River	0.12 <	0.10 <	0.10	0.10	0.68	2	1.359530139	0.87
93-217	H1-1	Cooper River	0.12 <	0.10 <	0.10	0.10	1.55	2	3.106951965	0.80
93-218	H1-4	Cooper River	0.12 <	0.10 <	0.10	0.10	0.12	2	0.243929825	0.80
93-219	H1-5	Cooper River	0.12 <	0.10	0.11	0.11	5.83	2	11.656	0.80
93-220	F2-1	Wando River	0.12 <	0.10 <	0.10	0.10	0.00	2	0	0.80
93-221	F2-2	Wando River	0.12 <	0.10 <	0.10	0.10	0.00	2	0	0.80
93-222	F2-4	Wando River	0.12 <	0.10 <	0.10	0.10	0.14	2	0.289651877	0.80
93-223	G1-1	Cooper River	0.12 <	0.10 <	0.10	0.10	3.84	2	7.679730337	0.80
93-224	G2-1	Cooper River	0.12 <	0.10 <	0.10	0.10	5.64	2	11.28842105	0.88

Appendix B1 continued. Toxicity and chemistry data from Charleston Harbor, Leadenwah Creek, and Winyah Bay.

NMFS Lab. No.	STATION No.	Location	CL8(195) ug/kg	CL9(206) ug/kg	CL10(209) ug/kg	Sum of 20 Congeners ug/kg	2x ug/kg	total PCBs ug/kg	Pesticide Total : ug/kg	DDT Total :
93-225	CHP1	Shipyard Creek	0.12 <	0.10 <	0.10	5.48	2	10.95254658	0.80	0.91
93-226	CHP10	Detyen's Shipyard	0.12 <	0.10 <	0.10	4.19	2	8.378943005	1.08	0.55
93-227	Ref. No.	North Inlet								
93-228	B1-1	Winyah Bay	0.12	0.12 <	0.10	5.61	2	11.21590909	1.30	2.1
93-229	B3-1	Winyah Bay	0.12	0.10 <	0.10	1.64	2	3.288482412	0.80	1.16
93-230	B1-3	Winyah Bay	0.2	0.10 <	0.10	21.22	2	42.4365	3.50	5.17
93-231	B1-2	Winyah Bay	0.13	0.10 <	0.10	4.50	2	9.0005255714	0.96	1.51
93-232	CHP11	I of P Connector	0.12 <	0.10 <	0.10	0.19	2	0.387918644	0.80	0.71
93-233	B7-1	Winyah Bay	0.12	0.10 <	0.10	2.36	2	4.7208	0.94	1.03
93-234	B6-2	Winyah Bay	0.12	0.10 <	0.10	1.01	2	2.027653179	0.80	1.13
93-235	B5-1	Winyah Bay	0.12	0.10 <	0.10	0.21	2	0.427706619	0.80	0.55
93-236	B4-1	Winyah Bay	0.12	0.16 <	0.10	2.72	2	5.434941176	0.80	1.47
93-237	B2-1	Winyah Bay	0.12	0.13	0.11	1.14	2	2.286772532	0.82	1.21
brez	ref1									
brez	ref1									
star	ref2									
star	ref2									
USC lab ct batch 2										
USC lab ct batch 3										
NIOL Ref 1 batch 1										
NIOL Ref 1 batch 2										
NIOL Ref 1 batch 3										

ns non-significant

^ Microtox Tests: NMFS 93-146 to 93-186 compared to reference 177. NMFS 93-196 to 93-237 compared to reference 227.

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

Duplicates

93-165	D2-1	Ashley River
93-165	D2-1	Ashley River
93-166	D2-3	Ashley River
93-166	D2-3	Ashley River
93-167	D2-2	Ashley River
93-167	D2-2	Ashley River

Appendix B2. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Lab. No. Number	Location	% Amph surv. as % of control	Amph Surv (% of sample)	Signif. (% of control)	Microtox EC50 (mg/ml)	Microtox (273; 216 and 242)° in 100% pw	Microtox % ref. (273; 216 and 242)° in 100% pw	Signif. % urch fert. in 100% pw
94-216	Ref. No. Inlet			11.28816667	0.84	100.0000059			
94-217	E1,2-1	South Channel	93	100	ns	7.758366667	0.56	70.34589692	*
94-218	E1,3-1	South Channel	93	100	ns	67.66666667	4.84	613.5405251	ns
94-219	E1-1	South Channel	93	100	ns	4.8972	0.38	44.40340876	*
94-220	C1	Back River	85	91	@	0.979766667	0.07	8.883643672	**
94-221	E2-2	South Channel	95	102	ns	51	3.82	462.4221691	ns
94-222	E2-3	South Channel	92	99	ns	3.742533333	0.29	33.93392906	*
94-223	E2-1	South Channel	87	94	ns	69.30493333	5.24	628.394855	ns
94-224	C3	Back River	90	97	ns	67.66666667	5.03	613.5405251	ns
94-225	C2	Back River	85	91	ns	1.055433333	0.08	9.569721007	**
94-226	D1,1-1	Lower Channel	85	91	ns	1.192733333	0.09	10.81463402	**
94-227	D1,2-2	Lower Channel	94	101	ns	67.66666667	5.00	613.5405251	ns
94-228	D1,3-1	Lower Channel	90	97	ns	1.1924	0.09	10.81161166	**
94-229	D2,1-1	Lower Channel	89	94	ns	67.66666667	5.04	613.5405251	ns
94-230	D2,2-2	Lower Channel	91	96	ns	101	7.54	915.7772369	ns
94-231	D2,3-1	Lower Channel	83	87	ns	8.0994	0.61	73.43808072	*
94-242	Ref. No. Inlet			10.7696	0.82	95.406			
94-243	E3-2	South Channel	89	94	ns	3.068633333	0.24	27.82360946	*
94-244	E3-5	South Channel	90	95	ns	0.084266667	0.01	0.764054408	**
94-245	D3,1-4	Lower Channel	84	88	@	51	3.88	462.4221691	*
94-246	D3,2-1	Lower Channel	95	100	ns	30.13476667	2.25	273.2349837	ns
94-247	D3,3-2	Lower Channel	91	96	ns	2.1175	0.16	19.19958712	**
94-248	E3-9	South Channel	91	96	ns	1.616433333	0.12	14.65636487	**
94-266	B5-1	City Channel	96	102	ns	66.74363333	5.08	650.4382358	ns
94-267	B5-2	City Channel	98	104	ns	95.094	6.77	926.7217037	ns
94-268	G1	Ocean Terminal	91	97	ns	2.079633333	0.16	20.26669764	**
94-269	G2	Ocean Terminal	96	102	ns	7.769366667	0.59	75.71498427	ns
94-270	H1	Steven Terminal	97	103	ns	3.181533333	0.25	31.00506857	**
94-271	H2	Steven Terminal	92	98	ns	2.7673	0.21	26.96823113	**

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	% Amph surv. as % of control	Amph Surv Signif.	Microtox EC50 (% of sample)	Microtox (mg/ml)	Microtox % ref. (273, 216 and 242) ^r	Signif. in 100% pw
94-272	H3	Steven Terminal		2.507133333	0.19	24.43282304		
94-273	Ref. No. Inlet			10.261333	0.77	100.0000032		
94-274	B9-1	City Channel	94	100	0.11	14.14273695	* *	48.4
94-275	B9-2	City Channel	91	97	0.21	26.32211624	* *	8.5
94-276	B9-3	City Channel	94	100	0.10	13.70062415	* *	31.6
94-277	F1	Dundee Canal	95	101	0.47	60.4476371	ns	7.2
94-278	F2	Dundee Canal	95	101	0.32	41.8317971	* *	10.4
94-279	B5-3	City Channel	95	101	0.07	10.2621495	***	8.6
94-280	B7-1	City Channel	98	101	0.07	9.215501859	***	71.8
94-281	B8-1	City Channel	95	98	51	3.81	497.0114507	ns
94-282	B8-2	City Channel	96	99	4.22	541.4221849	ns	98.4
94-283	B2-1	City Channel	96	99	0.17	21.8948162	**	3.4
94-284	B2-3	City Channel	96	99	0.18	22.51364418	**	5.2
94-285	B2-4	City Channel	96	99	0.57	72.28463073	ns	7.4
94-286	B8-3	City Channel	94	97	0.41	52.13325273	ns	10.4
94-287	A2-2	Upper River	81	87	0.68	86.61447787	ns	99.8
94-288	A2-3	Upper River	88	95	0.23	30.27189547	**	98.4
94-289	A2-4	Upper River	84	90	*	51	4.00	497.0114507
94-290	B6-1	City Channel	94	101	ns	84.33333333	6.38	821.8555361
94-291	B6-2	City Channel	89	96	ns	10.65346667	0.80	103.8214691
94-292	B6-3	City Channel	91	98	ns	1.5819	0.12	15.41612576
94-293	A1-1	Upper River	4	**	51	3.89	497.0114507	ns
94-294	A1-2	Upper River	83	89	@	51	3.87	497.0114507
94-295	A1-3	Upper River	78	84	@	3.1399	0.23	30.59933831
94-296	B7-2	City Channel	89	96	ns	67.66668667	3.93	659.4334934
94-297	B7-3	City Channel	97	109	ns	67.66668667	5.16	659.4334934
94-298	B1-1	City Channel	90	101	ns	51	3.81	497.0114507
94-299	B1-2	City Channel	89	100	ns	51	3.79	497.0114507
94-300	B1-5	City Channel	92	103	ns	51	3.83	497.0114507
94-301	B3-3	City Channel	90	101	ns	1.647266667	0.12	16.05314501
94-302	B3-5	City Channel	93	104	ns	2.6534	0.20	25.85823889
94-303	B3-6	City Channel	86	97	ns	4.636533333	0.34	45.1845129
94-304	B4-4	City Channel	93	104	ns	1.954066667	0.15	19.04300997
94-305	B4-5	City Channel	96	108	ns	2.064166667	0.15	20.11595999
94-306	B4-6	City Channel	91	102	ns	5.0255	0.38	48.97511853
	Ref. Redfish Bay (upper Savannah)				0.82			98.8
	Ref. No. Inlet (upper Savannah)				0.77			98.4
	Ref. No. Inlet (Lower Savannah)							99.6
	Ref. Redfish Bay (Lower Savannah)							99.3

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No. *RE-RUN 1-24-94	STATION Number	Location	% Amph surv. as % of control	Amph Surv (µg/g)	Signif. (% of sample)	Microtox EC50 (mg/ml)	Microtox (mg/ml)	Microtox % ref. (273; 216 and 242)^\n	Signif. in 100% pw
94-295	A1-3								
94-295R*	A1-3								
94-293	A1-1								
94-293	A1-1								
94-294	A1-2								
94-294	A1-2								
94-301	B3-3								
94-301	B3-3								
94-302	B3-5								
94-302	B3-5								
94-303	B3-6								
94-303	B3-6								
94-304	B4-4								
94-304	B4-4								
94-305	B4-5								
94-305	B4-5								
94-306	B4-6								
94-306	B4-6								
94-267	B5-2								
94-267	B5-2								
94-279	B5-3								
94-279	B5-3								
94-274	B9-1								
94-274	B9-1								
94-244	E3-5								
94-244	E3-5								
94-277	F1								
94-277	F1								
94-269	G2								
94-269	G2								
94-271	H2								
94-271	H2								

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.
MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^\n Microtox Tests: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.
ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnell's t-test at 0.01

* statistically significant for Mann-Whitney and Dunnets

** statistically significant for Mann-Whitney, Dunnets, and Distribution-free

@ sample mean was statistically different from control

- sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	Signif. (Ref. Redfish Bay) in 50% pw.	% urch fert. in 25% pw.	Signif. (Ref. Redfish Bay)	% urch fert. in 25% pw.	Signif. (Ref. Redfish Bay) in 100% pw.	Signif. (Ref. Redfish Bay)
94-216	Ref. No. Inlet						91.2	
94-217	E1,2-1	South Channel	ns	100.0	ns	99.4	ns	96.8
94-218	E1,3-1	South Channel	ns	99.6	ns	99.6	ns	97.6
94-219	E1-1	South Channel	ns	99.8	ns	99.6	ns	64.6
94-220	C1	Back River	ns	99.6	ns	95.2	ns	91.4
94-221	E2-2	South Channel	ns	99.4	ns	99.4	ns	98.4
94-222	E2-3	South Channel	ns	99.2	ns	98.8	ns	36.8
94-223	E2-1	South Channel	ns	99.4	ns	99.6	ns	97.8
94-224	C3	Back River	ns	99.6	ns	100.0	ns	98.6
94-225	C2	Back River	ns	99.8	ns	98.8	ns	64.4
94-226	D1,1-1	Lower Channel	ns	100.0	ns	99.4	ns	0
94-227	D1,2-2	Lower Channel	ns	99.2	ns	99.2	ns	96.4
94-228	D1,3-1	Lower Channel	++	98.6	ns	99.2	ns	3.6
94-229	D2,1-1	Lower Channel	ns	99.6	ns	99.4	ns	96.2
94-230	D2,2-2	Lower Channel	ns	99.2	ns	99.4	ns	98.2
94-231	D2,3-1	Lower Channel	ns	98.4	ns	96.8	ns	98.2
94-242	Ref. No. Inlet							
94-243	E3-2	South Channel	ns	99.8	ns	99.0	ns	97.8
94-244	E3-5	South Channel	ns	99.6	ns	99.2	ns	1
94-245	D3,1-4	Lower Channel	ns	100.0	ns	99.8	ns	97.4
94-246	D3,2-1	Lower Channel	ns	99.6	ns	99.4	ns	98
94-247	D3,3-2	Lower Channel	ns	99.4	ns	99.6	ns	19
94-248	E3-9	South Channel	ns	99.4	ns	100.0	ns	3
94-266	B5-1	City Channel	ns	99.4	ns	99	ns	0
94-267	B5-2	City Channel	ns	98.2	ns	98.2	ns	0
94-288	G1	Ocean Terminal	++	97.2	++	98.8	ns	0
94-289	G2	Ocean Terminal	ns	94.4	ns	98.2	ns	0.2
94-270	H1	Steven Terminal	++	94.6	ns	98.2	ns	0
94-271	H2	Steven Terminal	ns	96.4	ns	99.4	ns	0

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	Signif. (Ref. Redfish Bay) in 50% pw.	% urch fert. in 25% pw.	Signif. (Ref. Redfish Bay)	% urch fert. in 25% pw.	Signif. (Ref. Redfish Bay) in 100% pw	Signif. (Ref. Redfish Bay)
94-272	H3	Steven Terminal						
94-273	Ref. No. Inlet							
94-274	B9-1	City Channel	++#	96.8	ns	99.8	ns#	0
94-275	B9-2	City Channel	++#	96.6	ns#	99.0	ns#	0
94-276	B9-3	City Channel	++#	97.4	ns#	99.0	ns#	0
94-277	F1	Dundee Canal	++#	95.2	ns#	99.4	ns#	2.6
94-278	F2	Dundee Canal	++#	93.2	ns#	97.0	ns#	++#
94-279	B5-3	City Channel	++	87.4	ns	94.8	rs	2
94-280	B7-1	City Channel	++	96.6	ns	96.4	rs	++
94-281	B8-1	City Channel	ns	98	ns	99.6	rs	++
94-282	B8-2	City Channel	ns	99.2	ns	98.8	rs	0.4
94-283	B2-1	City Channel	++	86.75	ns	96.8	rs	++
94-284	B2-3	City Channel	++	67	++	95	rs	++
94-285	B2-4	City Channel	++	88	ns	98.2	rs	++
94-286	B8-3	City Channel	++#	80.2	++#	99.4	ns#	++#
94-287	A2-2	Upper River	ns	99.2	ns	99	rs	90.4
94-288	A2-3	Upper River	ns	99.4	ns	98.4	rs	25.2
94-289	A2-4	Upper River	ns	99.2	ns	99.8	rs	88.8
94-290	B6-1	City Channel	ns	98.8	ns	99.6	rs	0
94-291	B6-2	City Channel	ns	99.8	ns	99.2	rs	++
94-292	B6-3	City Channel	ns	98.2	ns	98.8	rs	++
94-293	A1-1	Upper River	ns	99.2	ns	98.6	rs	ns
94-294	A1-2	Upper River	ns	99.4	ns	98.8	rs	++
94-295	A1-3	Upper River	ns	98.4	ns	99.4	rs	95
94-296	B7-2	City Channel	ns	99.2	ns	98.6	rs	0
94-297	B7-3	City Channel	ns	98	ns	99	rs	++
94-298	B1-1	City Channel	ns	99	ns	98.6	rs	93.2
94-299	B1-2	City Channel	ns	99	ns	98.2	rs	92
94-300	B1-5	City Channel	ns	99.6	ns	98.6	rs	92.8
94-301	B3-3	City Channel	++	78.6	++	98.4	rs	0.8
94-302	B3-5	City Channel	++	53.2	++	96.6	rs	0
94-303	B3-6	City Channel	++	72.8	++	96.8	rs	++
94-304	B4-4	City Channel	ns	99.4	ns	98.4	rs	ns
94-305	B4-5	City Channel	++	94.2	ns	96.2	rs	92.8
94-306	B4-6	City Channel	++	95	ns	98.8	rs	0
	Ref. Redfish Bay (upper Savannah)			99.4		99.3		91.2
	Ref. No. Inlet (upper Savan			98.6		99.2		1.5
	Ref. No. Inlet (Lower Savan			100		99.8		31.2
	Ref. Redfish Bay (Lower Savannah)			99.0		99.3		99.2

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

4-271 H2

MSL samples 94-266 to 94-306 collected from Upper Savannah.

MSL samples 94-300 collected from Upper Savannah.

protox Tests: Upper Savannah samples co-

OTU tests. Up to 30% samples were significant

ion-significant

statistically significant for Dunnett's t-test at 0.05

statistically significant for Dunnett's t-test at 0.01

statistically significant for Dunnett's t-test at 0.01

statistically significant for Mann-Whitney

statistically significant for Mann-Whitney and Dunnell

statistically significant for Mann-Whitney and Dunnett

statistically significant for Mann-Whitney, Dunnett's,

sample mean was statistically different from control

sample mean was statistically different from control

sample mean 80% or less than the control

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Ref. No. Number	Location	% urchdevp in 50% pw.	Signif. (Ref. Redfish Bay)	%urchdevp in 25% pw.	Signif. (Ref. Redfish Bay)	UAN, amph start mg/l	UAN, amph end mg/l
94-216	Ref. No. Inlet		93.5		93.5			
94-217	E1,2-1	South Channel	98.6	ns	98	ns	ns	ns
94-218	E1,3-1	South Channel	98.2	ns	97.6	ns	ns	ns
94-219	E1-1	South Channel	98.4	ns	97.4	ns	ns	ns
94-220	C1	Back River	98	ns	98.6	ns	ns	ns
94-221	E2-2	South Channel	98.2	ns	98	ns	ns	ns
94-222	E2-3	South Channel	96.8	ns	97.6	ns	ns	ns
94-223	E2-1	South Channel	98.6	ns	98.2	ns	ns	ns
94-224	C3	Back River	98	ns	96.4	ns	ns	ns
94-225	C2	Back River	97.6	ns	96.6	ns	ns	ns
94-226	D1,1-1	Lower Channel	91.6	ns	97.6	ns	ns	ns
94-227	D1,2-2	Lower Channel	96.4	ns	98.8	ns	ns	ns
94-228	D1,3-1	Lower Channel	0	++	14.6	++	++	++
94-229	D2,1-1	Lower Channel	97.8	ns	98.4	ns	ns	ns
94-230	D2,2-2	Lower Channel	98.2	ns	98.2	ns	ns	ns
94-231	D2,3-1	Lower Channel	97	ns	98.2	ns	ns	ns
94-242	Ref. No. Inlet							
94-243	E3-2	South Channel	95.8	ns	98.8	ns	ns	ns
94-244	E3-5	South Channel	0	++	79.2	++	++	++
94-245	D3,1-4	Lower Channel	98.4	ns	96.8	ns	ns	ns
94-246	D3,2-1	Lower Channel	97.8	ns	98.8	ns	ns	ns
94-247	D3,3-2	Lower Channel	0	++	97.2	ns	ns	ns
94-248	E3-9	South Channel	0	++	97.2	ns	ns	ns
94-266	B5-1	City Channel	76.8	+	94.8	ns	ns	ns
94-267	B5-2	City Channel	62.4	++	91.2	ns	ns	ns
94-268	G1	Ocean Terminal	0	++	78.4	+	+	+
94-269	G2	Ocean Terminal	0	++	28.8	++	++	++
94-270	H1	Steven Terminal	0	++	0.6	++	++	++
94-271	H2	Steven Terminal	0	++	89.8	ns	ns	ns

Appendix B continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No. *RE-RUN 1-24-94	STATION Number	Location	UAN, amph ave. mg/l	Pwater UAN ug/l	Ag ($\mu\text{g/g}$)	Al (%)	As ($\mu\text{g/g}$)	Cd ($\mu\text{g/g}$)	Cr ($\mu\text{g/g}$)	Cu ($\mu\text{g/g}$)	Fe (%)	Hg ($\mu\text{g/g}$)
--	-------------------	----------	------------------------	--------------------	---------------------------	-----------	---------------------------	---------------------------	---------------------------	---------------------------	-----------	---------------------------

MSL	STATION	1-METHYLNAPHTHALE BIPHENYL GC, ug/kg	GC, ug/kg	2,6-dimene nap GC, ug/kg	GC, ug/kg	ACENAPHTHACENAPHTHENE 2,3,5-trime nap GC, ug/kg	GC, ug/kg
94-295	A1-3	2	<	2	<	1	<
94-295R*	A1-3	2	<	3	<	3	<
94-293	A1-1	2	<	3	<	3	<
94-293	A1-1	2	<	3	<	3	<
94-294	A1-2	2	<	3	<	3	<
94-294	A1-2	2	<	3	<	3	<
94-301	B3-3	2	<	2	<	1	<
94-301	B3-3	2	<	2	<	1	<
94-302	B3-5	2	<	2	<	1	<
94-302	B3-5	2	<	2	<	1	<
94-303	B3-6	2	<	2	<	1	<
94-303	B3-6	2	<	2	<	1	<
94-304	B4-4	2	<	2	<	1	<
94-304	B4-4	2	<	2	<	1	<
94-305	B4-5	2	<	2	<	1	<
94-305	B4-5	2	<	2	<	1	<
94-306	B4-6	2	<	2	<	1	<
94-306	B4-6	2	<	2	<	1	<
94-267	B5-2	2	<	2	<	1	<
94-267	B5-2	2	<	2	<	1	<
94-279	B5-3	2	<	2	<	1	<
94-279	B5-3	2	<	2	<	1	<
94-274	B9-1	2	<	2	<	1	<
94-274	B9-1	2	<	2	<	1	<
94-244	E3-5	2	<	2	<	1	<
94-244	E3-5	2	<	2	<	1	<
94-277	F1	2	<	2	<	1	<
94-277	F1	2	<	2	<	1	<
94-269	G2	2	<	2	<	1	<
94-269	G2	2	<	2	<	1	<
94-271	H2	2	<	2	<	1	<
94-271	H2	2	<	2	<	1	<

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.
MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Microtox Tests: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.
ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney and Dunnett's

** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

@ sample mean was statistically different from control

- sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Lab. Id. No. Number	Location	Mn ($\mu\text{g/g}$)	Ni ($\mu\text{g/g}$)	Pb ($\mu\text{g/g}$)	Se ($\mu\text{g/g}$)	Sn ($\mu\text{g/g}$)	Zn ($\mu\text{g/g}$)	% sand	% silt	% clay
94-216	Ref. No. Inlet										
94-217	E1,2-1	South Channel	121	2.15	7.1	0.14	<	7.8	29.9	88.9	3.7
94-218	E1,3-1	South Channel	259	9.6	11.1	< 0.03	<	7.8	39.6	77.7	4.6
94-219	E1-1	South Channel	54	< 1.9	2.8	0.07	<	7.8	12.9	92.5	2.0
94-220	C1	Back River	272	12.25	17.1	0.4		38.8	73.6	44.1	13.1
94-221	E2-2	South Channel	209	11.88	10.2	0.2		25.5	40.4	44.7	12.0
94-222	E2-3	South Channel	200	8.25	7.9	0.22		16.9	36.0	63.6	9.5
94-223	E2-1	South Channel	72	< 1.9	4.3	< 0.03	<	7.8	11.8	93.9	0.7
94-224	C3	Back River	66	< 1.9	2.3	< 0.03	<	7.8	8.2	96.7	1.4
94-225	C2	Back River	369	16.34	23.0	0.41		44.4	81.4	21.4	21.2
94-226	D1,1-1	Lower Channel	312	9.42	11.7	< 0.03		23.8	41.2	56.8	15.2
94-227	D1,2-2	Lower Channel	735	12.14	11.5	0.25		30.2	41.9	67.4	4.3
94-228	D1,3-1	Lower Channel	547	15.55	18.6	0.04		35.6	72.4	25.8	22.6
94-229	D2,1-1	Lower Channel	634	15.88	14.0	0.66		30.8	50.8	49.4	12.4
94-230	D2,2-2	Lower Channel	152	5.92	7.8	< 0.03		14.6	24.5	77.4	6.0
94-231	D2,3-1	Lower Channel	90	3.03	4.5	0.09	<	7.8	16.1	86.7	5.6
94-242	Ref. No. Inlet										
94-243	E3-2	South Channel	206	3	3.8	< 0.03	<	7.8	18.5	94.8	1.8
94-244	E3-5	South Channel	486	16.13	17.5	0.56		42.8	67.5	12.1	16.4
94-245	D3,1-4	Lower Channel	73	< 1.9	1.9	< 0.03	<	7.8	5.3	99.0	0.9
94-246	D3,2-1	Lower Channel	73	< 1.9	2.2	0.11	<	7.8	9.6	94.7	1.5
94-247	D3,3-2	Lower Channel	322	12.24	11.9	< 0.03		19.4	45.0	42.8	14.6
94-248	E3-9	South Channel	561	16.58	18.4	0.11		28.3	74.5	13.2	22.9
94-266	B5-1	City Channel	637	19.26	19.6	< 0.03		40.1	89.5	5.9	29.5
94-267	B5-2	City Channel	879	24.96	24.7	< 0.03		46	108.8	0.2	24.8
94-268	G1	Ocean Terminal	990	21.33	26.2	1		50.3	112.8	1.0	31.8
94-269	G2	Ocean Terminal	1005	19.3	22.9	0.64		44.7	102.0	1.9	27.6
94-270	H1	Steven Terminal	599	23.22	32.7	0.07		59.8	129.5	0.4	18.5
94-271	H2	Steven Terminal	904	24.28	28.0	0.17		49.3	119.5	2.8	32.0

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	Mn ($\mu\text{g/g}$)	Ni ($\mu\text{g/g}$)	Pb ($\mu\text{g/g}$)	Se ($\mu\text{g/g}$)	Sn ($\mu\text{g/g}$)	Zn ($\mu\text{g/g}$)	% sand	% silt	% clay
94-272	H3	Steven Terminal	862	23.32	36.4	0.92	46.8	149.0	6.4	26.4	67.2
94-273	Ref. No. Inlet										
94-274	B9-1	City Channel	441	11.32	13.0	0.08	21.9	69.2	49.3	30.7	20.0
94-275	B9-2	City Channel	643	13.8	17.4	0.61	37.9	74.1	27.7	26.5	45.8
94-276	B9-3	City Channel	485	1.2	14.5	0.72	28	65.2	38.6	19.7	41.7
94-277	F1	Dundee Canal	883	24.12	24.0	0.08	59.7	105.2	7.7	16.5	75.8
94-278	F2	Dundee Canal	658	17.48	24.1	0.36	52.4	83.5	10.7	22.1	67.2
94-279	B5-3	City Channel	549	16.43	20.5	0.2	33.7	81.9	30.5	21.6	47.9
94-280	B7-1	City Channel	321	9.95	14.5	0.11	20.3	47.6	67.1	6.7	26.2
94-281	B8-1	City Channel	1114	22.46	24.5	0.03	47.7	105.6	5.7	50.3	44.1
94-282	B8-2	City Channel	1252	18.36	20.5	0.71	48.1	89.0	14.4	40.1	45.5
94-283	B2-1	City Channel	492	10.59	12.0	0.21	9.7	50.3	79.1	7.0	13.9
94-284	B2-3	City Channel	885	22.79	23.3	0.21	45	105.3	15.7	28.3	56.0
94-285	B2-4	City Channel	816	14.4	16.8	0.2	34.5	66.1	75.9	11.9	12.2
94-286	B8-3	City Channel	625	13.22	16.0	0.15	35	71.6	54.3	18.3	27.4
94-287	A2-2	Upper River	1107	27.02	26.3	0.23	66.8	87.4	16.2	23.5	60.3
94-288	A2-3	Upper River	468	10.39	10.4	0.21	26	46.5	65.3	19.0	15.7
94-289	A2-4	Upper River	195	15.04	12.4	0.67	34.3	49.4	52.7	11.0	36.4
94-290	B6-1	City Channel	784	14.92	19.7	0.17	34.8	16.2	2.5	29.1	68.4
94-291	B6-2	City Channel	848	17.44	22.1	0.42	40.6	84.6	16.5	37.0	46.5
94-292	B6-3	City Channel	901	18.12	23.1	< 0.03	40.5	92.8	41.1	12.9	46.0
94-293	A1-1	Upper River	430	21.38	20.0	0.54	48	72.7	47.4	9.6	43.0
94-294	A1-2	Upper River	271	6.24	8.3	< 0.03	26	27.3	75.5	9.2	15.4
94-295	A1-3	Upper River	696	20.26	22.3	0.52	49.5	72.6	35.4	24.6	40.1
94-296	B7-2	City Channel	1137	16.4	19.1	0.63	40.2	79.4	7.9	28.8	63.4
94-297	B7-3	City Channel	1103	23.14	24.1	0.79	58.9	113.3	1.8	33.8	64.4
94-298	B1-1	City Channel	254	7.25	9.3	0.27	22.1	38.3	75.1	7.4	17.6
94-299	B1-2	City Channel	111	< 1.9	2.7	0.05	< 7.8	9.6	96.7	1.9	1.4
94-300	B1-5	City Channel	130	2.2	5.2	< 0.03	< 7.8	12.1	96.7	1.9	1.4
94-301	B3-3	City Channel	273	5.44	6.8	0.27	12.1	40.4	77.9	8.5	13.6
94-302	B3-5	City Channel	1024	22.06	24.9	0.22	49.6	114.4	8.7	24.3	67.0
94-303	B3-6	City Channel	1013	21.78	25.1	0.92	50.1	106.2	12.5	29.5	58.0
94-304	B4-4	City Channel	127	3.09	7.5	0.04	9	21.1	93.3	2.9	3.8
94-305	B4-5	City Channel	200	4.71	7.2	< 0.03	< 7.8	37.8	88.2	5.2	6.6
94-306	B4-6	City Channel	713	18.22	19.8	< 0.03	36.3	86.8	32.4	24.1	43.5
		Ref. Redfish Bay (upper Savannah)									
		Ref. No. Inlet (upper Savannah)									
		Ref. No. Inlet (Lower Savannah)									
		Ref. Redfish Bay (Lower Savannah)									

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

note: MLS samples 94-266 to 94-306 collected from Upper Savannah.
MLS samples 94-217 to 94-248 samples collected from Lower Savannah.

Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.

*Significant
†Statistically significant for Dunnell's t-test at 0.05
‡Statistically significant for Dunnett's t-test at 0.01

*Statistically significant for Mann-Whitney and Dunnets

ole mean was statistically different from control
ole assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	% fines	% TOC	Cd (umole/g)	Cu (umole/g)	Ni (umole/g)	Pb (umole/g)	Zn (umole/g)	Hg (umole/g)
94-216	Ref. No. Inlet			<0.003	*	0.054	<0.052	*	<0.054	*
94-217	E1,2-1	South Channel	11.1	0.36	0.002	*	0.357	0.068	0.043	*
94-218	E1,3-1	South Channel	22.3	1.75	0.005	*	<0.012	*	<0.078	*
94-219	E1-1	South Channel	7.5	0.35	<0.000	*	0.002	*	<0.007	*
94-220	C1	Back River	55.9	1.84	0.064	*	0.531	0.728	*	1.330
94-221	E2-2	South Channel	55.3	1.42	0.003	*	<0.007	*	<0.043	*
94-222	E2-3	South Channel	36.4	1.18	<0.004	*	<0.009	*	<0.057	*
94-223	E2-1	South Channel	6.1	0.29	0.002	*	<0.002	*	<0.013	*
94-224	C3	Back River	3.3	0.11	0.001	*	<0.002	*	<0.012	*
94-225	C2	Back River	78.7	0.00	0.021	*	<0.038	*	<0.231	*
94-226	D1,1-1	Lower Channel	43.2	1.42	<0.013	*	<0.034	*	<0.209	*
94-227	D1,2-2	Lower Channel	32.6	2.11	<0.003	*	0.073	<0.045	*	<0.047
94-228	D1,3-1	Lower Channel	74.2	2.08	<0.008	*	<0.020	*	<0.122	*
94-229	D2,1-1	Lower Channel	50.6	2.28	0.005	*	<0.009	*	<0.055	*
94-230	D2,2-2	Lower Channel	22.6	1.11	<0.002	*	<0.004	*	<0.024	*
94-231	D2,3-1	Lower Channel	13.3	0.04	<0.001	*	0.023	<0.021	*	0.026
94-242	Ref. No. Inlet			0.003	*	<0.007	*	<0.043	*	<0.044
94-243	E3-2	South Channel	5.2	0.60	<0.003	*	0.035	0.056	*	<0.048
94-244	E3-5	South Channel	87.9	0.28	<0.024	*	<0.062	*	<0.381	*
94-245	D3,1-4	Lower Channel	1.0	0.28	<0.000	*	<0.001	*	<0.007	*
94-246	D3,2-1	Lower Channel	5.3	0.49	<0.001	*	0.007	<0.011	*	<0.008
94-247	D3,3-2	Lower Channel	57.3	1.88	<0.021	*	<0.053	<0.325	*	<0.335
94-248	E3-9	South Channel	86.8	2.53	<0.019	*	<0.048	*	<0.294	*
94-266	B5-1	City Channel	94.1	2.47	0.009	*	0.195	0.086	<0.019	*
94-267	B5-2	City Channel	99.8	3.70	<0.002	*	0.155	0.030	*	0.034
94-268	G1	Ocean Terminal	99.0	3.50	<0.039	*	<0.100	<0.610	*	<0.629
94-269	G2	Ocean Terminal	98.2	3.62	<0.011	*	0.560	<0.175	*	<0.181
94-270	H1	Steven Terminal	99.6	3.04	<0.011	*	0.259	<0.177	*	<0.183
94-271	H2	Steven Terminal	97.2	3.44	<0.012	*	<0.030	*	<0.183	*

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

240

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No. *RE-RUN 1-24-94	STATION Lab. Id. No. Number	Location	% fines	% TOC	Cd (umole/g)	Cu (umole/g)	Ni (umole/g)	Pb (umole/g)	Zn (umole/g)	Hg (umole/g)
--	-----------------------------------	----------	---------	-------	-----------------	-----------------	-----------------	-----------------	-----------------	-----------------

MSL	STATION	BENZO(A)PY PERYLENE			INDENO(1,2,3-(DIBENZO(G,H,I) Total			CL2(08)			HEXACHLOROBENZENE		
		GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg
94-295	A1-3	13	782	<	5	<	1	12	970	0.431	<	0.145	
94-295R*	A1-3	18	964	<	16	<	1	12	1236	0.274	<	0.225	
94-293	A1-1												
94-293	A1-1												
94-294	A1-2												
94-294	A1-2												
94-301	B3-3												
94-301	B3-3												
94-302	B3-5												
94-302	B3-5												
94-303	B3-6												
94-303	B3-6												
94-304	B4-4												
94-304	B4-4												
94-305	B4-5												
94-305	B4-5												
94-306	B4-6												
94-306	B4-6												
94-267	B5-2												
94-267	B5-2												
94-279	B5-3												
94-279	B5-3												
94-274	E9-1												
94-274	E9-1												
94-244	E3-5												
94-244	E3-5												
94-277	F1												
94-277	F1												
94-269	G2												
94-269	G2												
94-271	H2												
94-271	H2												

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.

MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Microtox Tests: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.
ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney, Dunnets, and Distribution-free

** statistically significant for Mann-Whitney, Dunnets, and Distribution-free

@ sample mean was statistically different from control

~ sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	SEM (umole/g)	AVS (umole/g)	SEM:AVS ratio	NAPHTHALENE		2-METHYLNAPHTHALENE		1-METHYLNAPHTHALENE	
						GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg
94-216	Ref. No. Inlet		0.268	0.599	0.447						
94-217	E1,2-1	South Channel	1.149	0.064	17.847 t	23					32
94-218	E1,3-1	South Channel	0.177	0.252	0.701	112					152
94-219	E1-1	South Channel	0.065	1.027	0.064	7					9
94-220	C1	Back River	15.422	34.244	0.450	7					10
94-221	E2-2	South Channel	0.340	0.116	2.937 t	90					117
94-222	E2-3	South Channel	0.134	6.016	0.022	231					274
94-223	E2-1	South Channel	0.055	0.037	1.496 t	233					331
94-224	C3	Back River	0.027	0.002	11.298 t	5					4
94-225	C2	Back River	0.989	23.118	0.043	24					37
94-226	D1,1-1	Lower Channel	1.013	11.947	0.085	43					64
94-227	D1,2-2	Lower Channel	0.540	0.407	1.328 t	37					55
94-228	D1,3-1	Lower Channel	0.533	13.092	0.041	5					6
94-229	D2,1-1	Lower Channel	0.130	2.035	0.064	13					15
94-230	D2,2-2	Lower Channel	0.056	0.337	0.166	16					17
94-231	D2,3-1	Lower Channel	0.234	0.669	0.349	32					22
94-242	Ref. No. Inlet		0.253	0.478	0.531						
94-243	E3-2	South Channel	0.308	0.580	0.531	15					16
94-244	E3-5	South Channel	0.890	22.525	0.040	13					18
94-245	D3,1-4	Lower Channel	0.034	0.003	9.912 t	7					8
94-246	D3,2-1	Lower Channel	0.154	0.235	0.657	8					9
94-247	D3,3-2	Lower Channel	0.762	6.670	0.114	21					20
94-248	E3-9	South Channel	0.667	29.475	0.023	36					38
94-266	B5-1	City Channel	0.928	0.007	129.187 t	30					30
94-267	B5-2	City Channel	0.914	0.080	11.495 t	100					118
94-268	G1	Ocean Terminal	1.388	8.032	0.173	62					51
94-269	G2	Ocean Terminal	1.562	6.691	0.233	60					72
94-270	H1	Steven Terminal	1.393	6.517	0.214	17					43
94-271	H2	Steven Terminal	0.431	4.293	0.100	9					24
											17

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id No.	STATION Number	Location	SEM ($\mu\text{mole/g}$)	AVS ($\mu\text{mole/g}$)	SEM:AVS ratio	NAPHTHALENE GC, ug/kg	NAPHTHALENE GC, ug/kg	2-METHYLNAPHTHALENE GC, ug/kg	1-METHYLNAPHTHALENE GC, ug/kg
94-272	H3	Steven Terminal	1.652	14.713	0.112	5	5	5	5
94-273	Ref. No. Inlet		0.380	3.668	0.103				
94-274	B9-1	City Channel	0.929	6.010	0.155	13	20	14	14
94-275	B9-2	City Channel	0.654	1.500	0.436	5	7	7	7
94-276	B9-3	City Channel	0.606	8.919	0.068	5	16	12	12
94-277	F1	Dundee Canal	0.880	1.692	0.520	19	53	38	38
94-278	F2	Dundee Canal	1.021	2.617	0.390	45	51	28	28
94-279	B5-3	City Channel	1.689	38.151	0.044	43	43	35	35
94-280	B7-1	City Channel	1.139	19.809	0.058	11	13	7	7
94-281	B8-1	City Channel	0.957	0.158	6.067 t	30	28	15	15
94-282	B8-2	City Channel	0.032	0.040	0.816	33	35	21	21
94-283	B2-1	City Channel	1.055	6.155	0.171	18	23	13	13
94-284	B2-3	City Channel	1.832	5.808	0.316	34	34	19	19
94-285	B2-4	City Channel	0.845	2.057	0.411	64	84	48	48
94-286	B8-3	City Channel	1.199	3.448	0.348	33	29	17	17
94-287	A2-2	Upper River	0.814	2.181	0.373	17	18	8	8
94-288	A2-3	Upper River	1.137	1.246	0.913	18	20	11	11
94-289	A2-4	Upper River	missing	0.028	missing	5	10	7	7
94-290	B6-1	City Channel	1.583	0.119	13.275 t	6	7	4	4
94-291	B6-2	City Channel	2.592	0.348	7.454 t	34	34	17	17
94-292	B6-3	City Channel	2.022	1.251	1.616 t	5	12	11	11
94-293	A1-1	Upper River	missing	0.298	missing	5	4	4	4
94-294	A1-2	Upper River	0.382	1.103	0.346	5 <	3 <	2 <	2 <
94-295	A1-3	Upper River	0.742	10.745	0.069	5 <	3 <	2 <	2 <
94-296	B7-2	City Channel	1.211	0.013	93.556 t	5 <	3 <	2 <	2 <
94-297	B7-3	City Channel	missing	0.051	missing	11	20	16	16
94-298	B1-1	City Channel	0.528	0.603	0.875	5 <	3	2	2
94-299	B1-2	City Channel	0.101	0.041	2.452 t	8	7	5	5
94-300	B1-5	City Channel	missing	0.030	missing	5 <	3 <	2 <	2 <
94-301	B3-3	City Channel	2.718	4.226	0.643	5 <	3 <	2 <	2 <
94-302	B3-5	City Channel	0.862	10.163	0.085	5 <	3 <	2 <	2 <
94-303	B3-6	City Channel	1.281	3.272	0.392	5	11	12	12
94-304	B4-4	City Channel	0.741	7.665	0.097	5 <	3 <	2 <	2 <
94-305	B4-5	City Channel	0.817	3.069	0.266	24	26	21	21
94-306	B4-6	City Channel	1.489	6.079	0.245	29	31	17	17
		Ref. Redfish Bay (upper Savannah)							
		Ref. No. Inlet (upper Savannah)							
		Ref. No. Inlet (Lower Savannah)							
		Ref. Redfish Bay (Lower Savannah)							

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

Note: MSI samples 94-266 to 94-306 collected from Upper Savannah.
MSI samples 94-217 to 94-248 samples collected from lower Savannah.

Microtox Tests: Upper Savannah samples compared to reference 273; lower Savannah samples compared to references 216 and 242.

s	non-significant	statistically significant for Dunnnett's t-test at 0.05	statistically significant for Dunnnett's t-test at 0.01	statistically significant for Mann-Whitney	statistically significant for Mann-Whitney and Dunnnett	sample mean was statistically different from control	sample assayed and statistically compared to Red
+							
*							
**							

卷之三

244

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Ref. No. Inlet	Location	BIPHENYL		2,6-dime nap		ACENAPHTHYLENE		ACENAPHTHENE		2,3,5-trime nap	
			Number	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg
94-216												
94-217	E1,2-1	South Channel	10		18		4		5		2	
94-218	E1,3-1	South Channel	23		50	v	1	v	3		7	
94-219	E1-1	South Channel	4		4		16		4		2	
94-220	C1	Back River	6		4		4		4		2	
94-221	E2-2	South Channel	16		36	v	1	v	3		1	
94-222	E2-3	South Channel	41		81	v	1	v	3		1	
94-223	E2-1	South Channel	50		115		34	v	3		1	
94-224	C3	Back River	3		2		1	v	3		1	
94-225	C2	Back River	17		27		9		64		10	
94-226	D1,1-1	Lower Channel	12		22		53	v	3		1	
94-227	D1,2-2	Lower Channel	8		19	v	1	v	3		2	
94-228	D1,3-1	Lower Channel	5		11		10	v	3		1	
94-229	D2,1-1	Lower Channel	3		6	v	1	v	3		2	
94-230	D2,2-2	Lower Channel	3		6	v	1	v	3		1	
94-231	D2,3-1	Lower Channel	4		8	v	1	v	3		2	
94-242	Ref. No. Inlet											
94-243	E3-2	South Channel	3		6		22	v	3		1	
94-244	E3-5	South Channel	5		8		3	v	3		1	
94-245	D3,1-4	Lower Channel	3		3		2	v	3		1	
94-246	D3,2-1	Lower Channel	3		4	v	1	v	3		1	
94-247	D3,3-2	Lower Channel	4		5	v	1	v	3		3	
94-248	E3-9	South Channel	7		15	v	1	v	5		6	
94-266	B5-1	City Channel	10		8		5		33		6	
94-267	B5-2	City Channel	23		46		21		221		7	
94-268	G1	Ocean Terminal	21		27		6		116		5	
94-269	G2	Ocean Terminal	19		17		9		77		3	
94-270	H1	Steven Terminal	20		30		30		66		8	
94-271	H2	Steven Terminal	12		17		14		98		4	

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

Ref. Redfish Bay (upper Savannah)

Ref. No. Inlet (upper Savannah)

Ref. No. Inlet (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	BIPHENYL GC, ug/kg	2,6-dime nap GC, ug/kg	ACENAPHTHENE GC, ug/kg	ACENAPHTHENE GC, ug/kg
*RE-RUN 1-24-94						

MSL	STATION	HEPTACHLORREPOXIDE CL4(66) ug/kg	CL4(66) ug/kg	2,4-DDE ug/kg	CL5(101) ug/kg	Alpha Chlordane ug/kg
94-295	A1-3	0.210	0.909	<	0.092	<
94-295R*	A1-3	0.210	3.706	<	1.144	0.359
94-293	A1-1					0.896
94-293	A1-1					0.345
94-294	A1-2					0.345
94-294	A1-2					
94-301	B3-3					
94-301	B3-3					
94-302	B3-5					
94-302	B3-5					
94-303	B3-6					
94-303	B3-6					
94-304	B4-4					
94-304	B4-4					
94-305	B4-5					
94-305	B4-5					
94-306	B4-6					
94-306	B4-6					
94-267	B5-2					
94-267	B5-2					
94-267	B5-2					
94-279	B5-3					
94-279	B5-3					
94-274	B9-1					
94-274	B9-1					
94-244	E3-5					
94-244	E3-5					
94-277	F1					
94-277	F1					
94-269	G2					
94-269	G2					
94-271	H2					
94-271	H2					

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.
MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Microtox Tests: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.
ns non-significant

+ statistically significant for Dunnell's t-test at 0.05
++ statistically significant for Dunnell's t-test at 0.01

*

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free
@ sample mean was statistically different from control
~ sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	FLUORENE		PHENANTHRENE		1-METHYLPHENANTHRENE		ANTHRACENE		FLUORANTHENE		PYRENE	
			GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg
94-216	Ref. No. Inlet													
94-217	E1,2-1	South Channel	5	14	<				3	5		13	18	
94-218	E1,3-1	South Channel	6	14			2	<	2	2		6	<	
94-219	E1-1	South Channel	4	49			16		18		165	150		
94-220	C1	Back River	4	19			4		7	7	43	51		
94-221	E2-2	South Channel	1	3	<		2	<	2	2		3	2	<
94-222	E2-3	South Channel	2	9	<		2	<	2	2		9	12	<
94-223	E2-1	South Channel	1	3	<		2	<	2	2		5	6	<
94-224	C3	Back River	2	27	<		2		7	7	20	17		
94-225	C2	Back River	43	341			51		30		362	324		
94-226	D1,1-1	Lower Channel	4	22	<		2		11		50	49		
94-227	D1,2-2	Lower Channel	2	6	<		2	<	2		3	4	<	
94-228	D1,3-1	Lower Channel	2	14	<		2		7	7	47	47		
94-229	D2,1-1	Lower Channel	1	3	<		2	<	2	2		3	<	
94-230	D2,2-2	Lower Channel	2	6	<		2	<	2	2		7	9	<
94-231	D2,3-1	Lower Channel	2	7	<		2		2	2		15	19	
94-242	Ref. No. Inlet													
94-243	E3-2	South Channel	2	4			3		7		73	87		
94-244	E3-5	South Channel	3	12			3		5		25	25		
94-245	D3,1-4	Lower Channel	1	4	<		2	<	2		3	3	<	
94-246	D3,2-1	Lower Channel	1	2	<		2	<	2		3	<	2	<
94-247	D3,3-2	Lower Channel	3	9			3		3		16	17		
94-248	E3-9	South Channel	6	15			6		12		31	34		
94-266	B5-1	City Channel	24	76			17		29		140	138		
94-267	B5-2	City Channel	89	554			27		191		465	372		
94-268	G1	Ocean Terminal	79	451			25		73		411	317		
94-269	G2	Ocean Terminal	38	252			15		44		233	193		
94-270	H1	Steven Terminal	42	373			43		150		986	828		
94-271	H2	Steven Terminal	50	301			23		71		461	382		

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	FLUORENE GC, ug/kg	PHENANTHRENE GC, ug/kg	1-METHYLPHENANTHRENE GC, ug/kg	ANTHRACENE GC, ug/kg	FLUORANTHENE GC, ug/kg	PYRENE GC, ug/kg
			258	44	21	73	21	380
94-272	H3	Steven Terminal						
94-273	Ref. No. Inlet							
94-274	B9-1	City Channel	7	61	8	22	134	129
94-275	B9-2	City Channel	10	84	9	21	127	108
94-276	B9-3	City Channel	12	60	2	26	135	114
94-277	F1	Dundee Canal	21	178	2	61	291	260
94-278	F2	Dundee Canal	24	145	2	55	278	258
94-279	B5-3	City Channel	96	123	2	52	250	169
94-280	B7-1	City Channel	6	14	2	8	49	45
94-281	B8-1	City Channel	17	138	20	37	188	165
94-282	B8-2	City Channel	89	495	2	71	493	383
94-283	B2-1	City Channel	10	56	6	20	68	62
94-284	B2-3	City Channel	18	130	18	69	173	157
94-285	B2-4	City Channel	166	875	39	139	717	532
94-286	B8-3	City Channel	27	180	18	58	260	233
94-287	A2-2	Upper River	6	33	4	10	49	54
94-288	A2-3	Upper River	23	201	19	40	298	256
94-289	A2-4	Upper River	7	36	2	11	99	119
94-290	B6-1	City Channel	1	7	7	2	6	< 2
94-291	B6-2	City Channel	12	80	10	48	150	140
94-292	B6-3	City Channel	19	118	13	25	178	187
94-293	A1-1	Upper River	1	5	2	2	10	17
94-294	A1-2	Upper River	1	5	2	2	6	< 2
94-295	A1-3	Upper River	2	16	2	5	28.5	42
94-296	B7-2	City Channel	5	100	12	24	199	202
94-297	B7-3	City Channel	20	160	14	34	248	229
94-298	B1-1	City Channel	3	18	2	5	25	26
94-299	B1-2	City Channel	1	5	2	2	6	< 7
94-300	B1-5	City Channel	1	9	2	<	12	13
94-301	B3-3	City Channel	3	18	3	9	34	30
94-302	B3-5	City Channel	19	198	2	43	396	330
94-303	B3-6	City Channel	48	457	41	96	645	526
94-304	B4-4	City Channel	4	31	2	7	45	39
94-305	B4-5	City Channel	4	31	5	12	81	69
94-306	B4-6	City Channel	29	159	2	29	191	163
		Ref. Redfish Bay (upper Savannah)						
		Ref. No. Inlet (upper Savannah)						
		Ref. No. Inlet (Lower Savannah)						
		Ref. Redfish Bay (Lower Savannah)						

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. id. No.	STATION Number	Location	PHENANTHRENE		1-METHYLPHENANTHRENE		ANTHRACENE		FLUORANTHENE		PYRENE	
			GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg
*RE-RUN 1-24-94												
MSL	STATION				DIELDRIN	4,4-DDE		CL4(77)	2,4-DDI CL5(118)		4,4-DDD	
94-295	A1-3			<	0.562	0.403	<	1.533	0.598	<	0.416	<
94-295R*	A1-3				0.403	0.371		7.977	1.566		2.976	<
94-293	A1-1								0.794			
94-293	A1-1											
94-294	A1-2											
94-294	A1-2											
94-301	B3-3											
94-301	B3-3											
94-302	B3-5											
94-302	B3-5											
94-303	B3-6											
94-303	B3-6											
94-304	B4-4											
94-304	B4-4											
94-305	B4-5											
94-305	B4-5											
94-306	B4-6											
94-306	B4-6											
94-267	B5-2											
94-267	B5-2											
94-279	B5-3											
94-279	B5-3											
94-274	B9-1											
94-274	B9-1											
94-244	E3-5											
94-244	E3-5											
94-277	F1											
94-277	F1											
94-269	G2											
94-269	G2											
94-271	H2											
94-271	H2											

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.

MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Microtox Test: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney and Dunnett's

** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

@ sample mean was statistically different from control

~ sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	BENZ(A)ANTHRACENE		CHRYSENE		BENZO(B)FLUORANTHENE		BENZO(K)FLUORANTHENE	
			GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg
94-216	Ref. No. Inlet									
94-217	E1,2-1	South Channel	4		3		7		3	
94-218	E1,3-1	South Channel	4	<	1	<	3		3	<
94-219	E1-1	South Channel	118		116		143		59	
94-220	C1	Back River	19		18		33		9	
94-221	E2-2	South Channel	4	<	1	<	3		3	<
94-222	E2-3	South Channel	4		3		4		3	<
94-223	E2-1	South Channel	4		2		3		3	<
94-224	C3	Back River	7		7		12		5	
94-225	C2	Back River	115		169		219		69	
94-226	D1,1-1	Lower Channel	26		27		88		39	
94-227	D1,2-2	Lower Channel	4	<	1	<	3		3	<
94-228	D1,3-1	Lower Channel	17		16		46		37	
94-229	D2,1-1	Lower Channel	4	<	1	<	3		3	<
94-230	D2,2-2	Lower Channel	4		3		8		6	
94-231	D2,3-1	Lower Channel	7		6		16		12	
94-242	Ref. No. Inlet									
94-243	E3-2	South Channel	159		145		179		90	
94-244	E3-5	South Channel	8		9		20		18	
94-245	D3,1-4	Lower Channel	4	<	1	<	3		3	<
94-246	D3,2-1	Lower Channel	4	<	1	<	3		3	<
94-247	D3,3-2	Lower Channel	8		8		20		14	
94-248	E3-9	South Channel	17		27		31		29	
94-246	B5-1	City Channel	55		58		89		63	
94-267	B5-2	City Channel	181		193		212		164	
94-268	G1	Ocean Terminal	120		117		199		158	
94-269	G2	Ocean Terminal	79		82		107		76	
94-270	H1	Steven Terminal	353		442		552		500	
94-271	H2	Steven Terminal	167		166		220		175	

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.						
MSL Lab. Id. No.	STATION Number	Location	BENZ(A)ANTHRACENE		BENZO(B)FLUORANTHENE	
			GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg
944-272	H3	Steven Terminal	180	232	235	215
944-273	Ref. No. Inlet					
944-274	B9-1	City Channel	55	61	81	57
944-275	B9-2	City Channel	34	37	51	48
944-276	B9-3	City Channel	35	38	52	37
944-277	F1	Dundee Canal	126	125	187	144
944-278	F2	Dundee Canal	113	171	261	207
944-279	B5-3	City Channel	8	<	3	<
944-280	B7-1	City Channel	18	23	36	26
944-281	B8-1	City Channel	58	74	107	75
944-282	B8-2	City Channel	125	189	201	143
944-283	B2-1	City Channel	29	34	44	41
944-284	B2-3	City Channel	54	73	80	57
944-285	B2-4	City Channel	173	214	201	159
944-286	B8-3	City Channel	105	115	209	166
944-287	A2-2	Upper River	23	37	69	49
944-288	A2-3	Upper River	123	139	153	121
944-289	A2-4	Upper River	36	56	76	54
944-290	B6-1	City Channel	4	<	1	25
944-291	B6-2	City Channel	73	130	111	17
944-292	B6-3	City Channel	81	96	129	79
944-293	A1-1	Upper River	7	8	18	91
944-294	A1-2	Upper River	4	4	7	13
944-295	A1-3	Upper River	14	20	31.5	5
944-296	B7-2	City Channel	112	127	149	14
944-297	B7-3	City Channel	97	92	174	61
944-298	B1-1	City Channel	7	11	15	61
944-299	B1-2	City Channel	4	2	4	3
944-300	B1-5	City Channel	4	4	5	3
944-301	B3-3	City Channel	7	8	10	3
944-302	B3-5	City Channel	121	150	183	4
944-303	B3-6	City Channel	261	252	283	62
944-304	B4-4	City Channel	16	17	18	224
944-305	B4-5	City Channel	30	29	37	8
944-306	B4-6	City Channel	63	65	84	16
		Ref. Redfish Bay (upper Savannah)				31
		Ref. No. Inlet (upper Savannah)				
		Ref. No. Inlet (Lower Savannah)				
		Ref. Redfish Bay (lower Savannah)				

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	BENZ(A)ANTHRACENE GC, ug/kg	CHRYSENE GC, ug/kg	BENZO(B)FLUORANTHENE GC, ug/kg	BENZO(K)FLUORANTHENE GC, ug/kg
			CL6(1) ug/kg	CL6(138) ug/kg	CL5(10:4,4-DDT ug/kg	CL5(1) ug/kg
*RE-RUN 1-24-94						
MSL	STATION		2,4-DDT ug/kg	CL6(153) ug/kg	CL5(10:4,4-DDT ug/kg	CL5(1) ug/kg
94-295	A1-3		0.382	< 1.322	0.086	0.791
94-295R*	A1-3		0.382	3.550	< 0.086	3.238
94-293	A1-1					
94-293	A1-1					
94-294	A1-2					
94-294	A1-2					
94-301	B3-3					
94-301	B3-3					
94-302	B3-5					
94-302	B3-5					
94-303	B3-6					
94-303	B3-6					
94-304	B4-4					
94-304	B4-4					
94-305	B4-5					
94-305	B4-5					
94-306	B4-6					
94-306	B4-6					
94-267	B5-2					
94-267	B5-2					
94-279	B5-3					
94-279	B5-3					
94-274	B9-1					
94-274	B9-1					
94-244	E3-5					
94-244	E3-5					
94-277	F1					
94-277	F1					
94-269	G2					
94-269	G2					
94-271	H2					
94-271	H2					

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.
MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Microtox Tests: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.
ns non-significant

+ statistically significant for Dunnett's t-test at 0.05
++ statistically significant for Dunnett's t-test at 0.01

*

** statistically significant for Mann-Whitney, and Dunnett's

*** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

@ sample mean was statistically different from control

- sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	BENZO(E)PYRENE		BENZO(A)PYRENE		PERYLENE		INDENO(1,2,3-C,D)PYRENE	
			GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg
94-216	Ref. No. Inlet									
94-217	E1,2-1	South Channel	4	<	5	8	<	5	<	5
94-218	E1,3-1	South Channel	3	<	5	42	<	5	<	5
94-219	E1-1	South Channel	54	95	33					61
94-220	C1	Back River	12		15	38				13 <
94-221	E2-2	South Channel	3	<	5	134	<	5	<	5
94-222	E2-3	South Channel	3	<	5	91	<	5	<	5
94-223	E2-1	South Channel	3	<	5	5	<	5	<	5
94-224	C3	Back River	4		6	9	<	5	<	5
94-225	C2	Back River	72		97	167	<	5	<	5
94-226	D1,1-1	Lower Channel	36		63	45				58
94-227	D1,2-2	Lower Channel	3	<	5	169	<	5	<	5
94-228	D1,3-1	Lower Channel	14		17	52	<	5	<	5
94-229	D2,1-1	Lower Channel	3	<	5	251	<	5	<	5
94-230	D2,2-2	Lower Channel	4	<	5	31	<	5	<	5
94-231	D2,3-1	Lower Channel	6		11	8				8 <
94-242	Ref. No. Inlet									
94-243	E3-2	South Channel	64		111	26				60
94-244	E3-5	South Channel	9		10	18	<	5	<	5
94-245	D3,1-4	Lower Channel	3	<	5	6	<	5	<	5
94-246	D3,2-1	Lower Channel	3	<	5	<	3	<	5	<
94-247	D3,3-2	Lower Channel	5		7	8	<	5	<	5
94-248	E3-9	South Channel	14		20	14	<	5	<	5
94-266	B5-1	City Channel	26		35	49				25 <
94-267	B5-2	City Channel	57		85	86				55
94-268	G1	Ocean Terminal	48		62	65				42
94-269	G2	Ocean Terminal	35		46	52				36 <
94-270	H1	Steven Terminal	196	216	150					152
94-271	H2	Steven Terminal	67	90	119					66

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	BENZO(E)PYRENE GC, ug/kg	BENZO(A)PYRENE GC, ug/kg	PERYLENE GC, ug/kg	INDENO(1,2,3-C,D)PYRENE GC, ug/kg
94-272	H3	Steven Terminal	72	98	61	62
94-273	Ref. No. Inlet					
94-274	B9-1	City Channel	28	34	69	30 <
94-275	B9-2	City Channel	19	23	71	16 <
94-276	B9-3	City Channel	19	22	67	5 <
94-277	F1	Dundee Canal	52	65	100	56
94-278	F2	Dundee Canal	3	<	3	5
94-279	B5-3	City Channel	3	<	5	5 <
94-280	B7-1	City Channel	12	16	42	12 <
94-281	B8-1	City Channel	35	42	221	28 <
94-282	B8-2	City Channel	50	60	452	44
94-283	B2-1	City Channel	15	19	41	13 <
94-284	B2-3	City Channel	29	34	223	5 <
94-285	B2-4	City Channel	71	81	300	59
94-286	B8-3	City Channel	48	62	133	5
94-287	A2-2	Upper River	23	20	841	23 <
94-288	A2-3	Upper River	53	73	104	52
94-289	A2-4	Upper River	28	27	548	23 <
94-290	B6-1	City Channel	6	<	5	5 <
94-291	B6-2	City Channel	42	54	97	5
94-292	B6-3	City Channel	54	67	80	5 <
94-293	A1-1	Upper River	6	7	1156	5 <
94-294	A1-2	Upper River	3	<	131	5 <
94-295	A1-3	Upper River	14	15.5	873	16 <
94-296	B7-2	City Channel	53	72	68	47 <
94-297	B7-3	City Channel	44	49	107	5 <
94-298	B1-1	City Channel	7	8	276	5 <
94-299	B1-2	City Channel	3	<	48	5 <
94-300	B1-5	City Channel	3	6	25	5 <
94-301	B3-3	City Channel	5	6	21	5 <
94-302	B3-5	City Channel	63	66	80	5
94-303	B3-6	City Channel	95	136	180	84
94-304	B4-4	City Channel	10	12	21	5 <
94-305	B4-5	City Channel	13	18	36	14 <
94-306	B4-6	City Channel	29	40	55	31
		Ref. Redfish Bay (upper Savannah)				
		Ref. No. Inlet (upper Savannah)				
		Ref. No. Inlet (Lower Savannah)				
		Ref. Redfish Bay (Lower Savannah)				

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id No. PRE-RUN 1-24-94	STATION Number	Location	BENZO(E)PYRENE		BENZO(A)PYRENE		PERYLENE		INDENO(1,2,3-C,D)PYRENE	
			GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg	GC, ug/kg
MSL	STATION				CL7(187)	CL6(128)	CL6(11 MIREX)			
94-295	A1-3		<	0.060	<	0.050	<	0.036	0.036	0.333
94-295R*	A1-3			0.661	<	0.050	<	0.036	0.036	0.873
94-293	A1-1									<
94-293	A1-1									
94-294	A1-2									
94-294	A1-2									
94-301	B3-3									
94-301	B3-3									
94-302	B3-5									
94-302	B3-5									
94-303	B3-6									
94-303	B3-6									
94-304	B4-4									
94-304	B4-4									
94-305	B4-5									
94-305	B4-5									
94-306	B4-6									
94-306	B4-6									
94-267	B5-2									
94-267	B5-2									
94-279	B5-3									
94-279	B5-3									
94-274	B9-1									
94-274	B9-1									
94-244	E3-5									
94-244	E3-5									
94-277	F1									
94-277	F1									
94-269	G2									
94-269	G2									
94-271	H2									
94-271	H2									

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.

MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Microtox Tests: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

@ sample mean was statistically different from control

~ sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	DIBENZ(A,H)ANTHRACENE GC, ug/kg	BENZO(G,H,I)PERYLENE GC, ug/kg	Total PAH GC, ug/kg	FLUORENE HPLC, ug/kg	PHENANTHRENE HPLC, ug/kg
94-216	Ref. No. Inlet						
94-217	E1,2-1	South Channel	1 <	3	13.8	16.6	
94-218	E1,3-1	South Channel	1 <	3	497	10.1	27.8 <
94-219	E1-1	South Channel	10	44	1185	172.5	51.1
94-220	C1	Back River	1	5	335	43.6	22 <
94-221	E2-2	South Channel	1 <	3	460	<	5.9
94-222	E2-3	South Channel	1 <	3	880	14	12.7 <
94-223	E2-1	South Channel	1 <	3	965	6.4	4.8 <
94-224	C3	Back River	1 <	3	134	20.6	32 <
94-225	C2	Back River	10	45	2326	365.5	362.2
94-226	D1,1-1	Lower Channel	11	53	806	50.4	29.2
94-227	D1,2-2	Lower Channel	1 <	3	333	<	5.9
94-228	D1,3-1	Lower Channel	3	14	365	47.1	11.5 <
94-229	D2,1-1	Lower Channel	1 <	3	302	<	5.9
94-230	D2,2-2	Lower Channel	1 <	3	127	9.7	7 <
94-231	D2,3-1	Lower Channel	1	5	205	16.4	8.3 <
94-242	Ref. No. Inlet						
94-243	E3-2	South Channel	12	45	1139	75.7 <	2.7 <
94-244	E3-5	South Channel	1	5	227	<	5.9
94-245	D3,1-4	Lower Channel	1 <	3	41	<	5.9
94-246	D3,2-1	Lower Channel	1 <	3	29	<	5.9
94-247	D3,3-2	Lower Channel	2 <	3	188	18.5	11.2 <
94-248	E3-9	South Channel	1	13	397	35.3	17.9 <
94-266	B5-1	City Channel	1	19	980	144.5	91
94-267	B5-2	City Channel	9	40	3372	485.5	581
94-268	G1	Ocean Terminal	7	32	2527	413.8	433.8
94-269	G2	Ocean Terminal	1	27	1610	227.3	269
94-270	H1	Steven Terminal	24	125	5375	1010.5	406.2
94-271	H2	Steven Terminal	9	53	2617	483	325.2

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	DIBENZ(A,H)ANTHRACENE GC, ug/kg	BENZO(G,H,I)PERYLENE GC, ug/kg	Total PAH GC, ug/kg	FLUORENE HPLC, ug/kg	PHENANTHRENE HPLC, ug/kg
94-272	H3	Steven Terminal	10		49	2392	406.4
94-273	Ref. No. Inlet						277.5
94-274	B9-1	City Channel	1		23	887	121.8
94-275	B9-2	City Channel	1		14	728	123.8
94-276	B9-3	City Channel	1		14	706	134.2
94-277	F1	Dundee Canal	8		40	1936	282.4
94-278	F2	Dundee Canal	2 <		3	1730	282.2
94-279	B5-3	City Channel	1 <		3	983	260
94-280	B7-1	City Channel	1		8	365	51.7
94-281	B8-1	City Channel	1		22	1349	197.6
94-282	B8-2	City Channel	10		30	3046	489.2
94-283	B2-1	City Channel	1		10	553	64.6
94-284	B2-3	City Channel	1		22	1279	167.6
94-285	B2-4	City Channel	9		46	4279	758.8
94-286	B8-3	City Channel	9		43	1778	249.9
94-287	A2-2	Upper River	1		19	1323	51.5
94-288	A2-3	Upper River	8		38	1798	301.7
94-289	A2-4	Upper River	1		14	1168	100.2
94-290	B6-1	City Channel	1 <		3	4190	< 5.9
94-291	B6-2	City Channel	8		34	1185	146.8
94-292	B6-3	City Channel	1		32	1294	181
94-293	A1-1	Upper River	1		4	1266	13.6 <
94-294	A1-2	Upper River	1 <		3	160	7.8 <
94-295	A1-3	Upper River	1		12	1103	21.8
94-296	B7-2	City Channel	1		39	1349	202.8
94-297	B7-3	City Channel	1		27	1547	259.6
94-298	B1-1	City Channel	1 <		3	416	25.4
94-299	B1-2	City Channel	1 <		3	107	8.9
94-300	B1-5	City Channel	1 <		3	77	16.2
94-301	B3-3	City Channel	1 <		3	159	36.2 <
94-302	B3-5	City Channel	7		45	1842	375.8
94-303	B3-6	City Channel	11		67	3608	629.6
94-304	B4-4	City Channel	1		7	246	46.4
94-305	B4-5	City Channel	1		12	504	79.3
94-306	B4-6	City Channel	5		25	1142	196
	Ref. Redfish Bay (upper Savannah)						
	Ref. No. Inlet (upper Savannah)						
	Ref. No. Inlet (Lower Savannah)						
	Ref. Redfish Bay (Lower Savannah)						

Appendix B2 continued Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL	STATION	DIBENZ(A,H)ANTHRACENE GC, ug/kg	BENZO(G,H,I)PYRELYNE GC, ug/kg	PHENANTHRENE HPLC, ug/kg
*RE-RUN 1-24-94				

MSL	STATION	CL7(170) ug/kg			CL8(195) ug/kg			CL10(209) ug/kg			Total of congeners ug/kg
		DIBENZ(A,H)ANTHRACENE GC, ug/kg	BENZO(G,H,I)PYRELYNE GC, ug/kg	Total PAH GC, ug/kg	FLUORENE HPLC, ug/kg	GC, ug/kg	HPLC, ug/kg	<	<	<	
94-295	A1-3										
94-295R*	A1-3										
94-293	A1-1										
94-293	A1-1										
94-294	A1-2										
94-294	A1-2										
94-301	B3-3										
94-301	B3-3										
94-302	B3-5										
94-302	B3-5										
94-303	B3-6										
94-303	B3-6										
94-304	B4-4										
94-304	B4-4										
94-305	B4-5										
94-305	B4-5										
94-306	B4-6										
94-306	B4-6										
94-267	B5-2										
94-267	B5-2										
94-279	B5-3										
94-279	B5-3										
94-274	B9-1										
94-274	B9-1										
94-244	E3-5										
94-244	E3-5										
94-277	F1										
94-277	F1										
94-269	G2										
94-269	G2										
94-271	H2										
94-271	H2										

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.
MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Microtox Tests: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242.

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney and Dunnett's

** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free

*** sample mean was statistically different from control

~ sample mean 80% or less than the control

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	ANTHRACENE HPLC, ug/kg		FLUORANTHENE HPLC, ug/kg		PYRENE HPLC, ug/kg		BENZA(A)ANTHRACENE HPLC, ug/kg		CHRYSENE HPLC, ug/kg		BENZO(B)FLUORANTHENE HPLC, ug/kg		BENZO(K)FLUORANTHENE HPLC, ug/kg	
			Ref. No. Inlet													
94-216	E1,2-1	South Channel	15.8	7	18.7	4.6 <	7.3	10.5 <	5.3 <	5.3 <	44.5 <	4.8 <	7.3	41.4	43.5 <	43.5 <
94-217	E1,3-1	South Channel	8.3 <	5.7	12.2 <	4.8 <	7.3	7.3	5.3 <	5.3 <	43.5 <	102.3	113	13.4	99.9	99.9
94-218	E1-1	South Channel	1.3 <	5.7	129.5	19.4	14.8	14.8	13.4	13.4	196 <	7.3	7.3	95.9 <	5.3 <	5.3 <
94-219	C1	Back River	8.3 <	5.7	53.2	4.8 <	7.3	7.3	5.3 <	5.3 <	7.3	16 <	4.8 <	7.3	8.8 <	5.3 <
94-220	E2-2	South Channel	8.3 <	5.7	7.3 <	4.8 <	7.3	7.3	5.3 <	5.3 <	95.9 <	16 <	4.8 <	7.3	8.8 <	5.3 <
94-221	E2-3	South Channel	8.3 <	5.7	16 <	4.8 <	7.3	7.3	5.3 <	5.3 <	95.9 <	7.3	7.3	8.8 <	5.3 <	5.3 <
94-222	E2-1	South Channel	8.3 <	5.7	7.9 <	4.8 <	7.3 <	7.3	5.3 <	5.3 <	8.8 <	7.9 <	7.9 <	8.8 <	5.3 <	5.3 <
94-223	C3	Back River	8.3 <	5.7	19.6	8.2	10.9	12.2	5.6	5.6	228.3	90.5	164	122	155	155
94-224	C2	Back River	20.5	35.9	340.1	99.5	164	228.3	155	155	45.5	23.3	23.3	45.5	31.6	31.6
94-225	D1,1-1	Lower Channel	8.8 <	5.7	52.3	26	23.3	153.1 <	5.3 <	5.3 <	153.1 <	7.3	7.3	153.1 <	5.3 <	5.3 <
94-226	D1,2-2	Lower Channel	8.3 <	5.7	11.6 <	4.8 <	7.3	153.1 <	5.3 <	5.3 <	153.1 <	11.6 <	11.6 <	153.1 <	5.3 <	5.3 <
94-227	D1,3-1	Lower Channel	8.3 <	5.7	51.2	18.1	11.4	51.1	14.1	14.1	350.2 <	7.3	7.3	350.2 <	5.3 <	5.3 <
94-228	D2,1-1	Lower Channel	8.3 <	5.7 <	7.3 <	4.8 <	7.3	7.3	5.3 <	5.3 <	47.5	7.3	7.3	47.5	5.9	5.9
94-229	D2,2-2	Lower Channel	8.3 <	5.7	7.7 <	4.8 <	7.3	7.3	5.3 <	5.3 <	14.8	10.8	11.1	14.8	14.2	14.2
94-230	D2,3-1	Lower Channel	8.3 <	5.7	15.7	10.8	11.1	11.1	11.1	11.1	10.8	10.8	10.8	10.8	10.8	10.8
94-231	Ref. No. Inlet	South Channel	8.3 <	5.7	91	180.4	199.5	44.4	136.3	136.3	199.5	199.5	199.5	199.5	10.5	10.5
94-232	E3-2	South Channel	8.3 <	5.7 <	7.3 <	4.8 <	7.3	21.3	21.3	21.3	7.3	7.3	7.3	7.3	12.2 <	12.2 <
94-244	E3-5	South Channel	8.3 <	5.7 <	7.3 <	4.8 <	7.3	12.2 <	5.3 <	5.3 <	8.8 <	7.3 <	7.3 <	8.8 <	5.3 <	5.3 <
94-245	D3,1-4	Lower Channel	8.3 <	5.7 <	7.3 <	4.8 <	7.3	12.2 <	5.3 <	5.3 <	8.8 <	7.3 <	7.3 <	8.8 <	12.2 <	12.2 <
94-246	D3,2-1	Lower Channel	8.3 <	5.7 <	7.3 <	4.8 <	7.3 <	12.2 <	5.3 <	5.3 <	8.8 <	7.3 <	7.3 <	8.8 <	12.2 <	12.2 <
94-247	D3,3-2	Lower Channel	8.3 <	5.7	19.7	10.9	14	19.1	12.1	12.1	27.3	21.8	21.8	27.3	30.4	30.4
94-248	E3-9	South Channel	8.3 <	5.7	35.6	66.3	68.8	74.2	53.3	53.3	383.7	330.5	330.5	383.7	139.4	139.4
94-266	B5-1	City Channel	25 <	5.7	126.9	66.3	68.8	74.2	53.3	53.3	383.7	330.5	330.5	383.7	125.5	125.5
94-267	B5-2	City Channel	160.7 <	5.7	380.8	182.7	174.7	174.7	139.4	139.4	383.7	330.5	330.5	383.7	93.1	93.1
94-288	G1	Ocean Terminal	56.7	69.3	314.4	117.4	113.2	113.2	97.1	97.1	383.7	330.5	330.5	383.7	41.2	41.2
94-289	G2	Ocean Terminal	40	35.5	179.9	92.1	140.2	140.2	59.4	59.4	383.7	330.5	330.5	383.7	312.8	312.8
94-270	H1	Steven Terminal	133.7 <	5.7	645.4	260.4	154.1	154.1	107.2	107.2	116.7	130.5	130.5	116.7	107.2	107.2
94-271	H2	Steven Terminal	62.9	50.5	50.5	260.4	154.1	154.1	107.2	107.2	116.7	130.5	130.5	116.7	107.2	107.2

Note: MSL samples 94-266 to 94-306 collected from Upper Savannah.

MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

^ Microtox Tests: Upper Savannah samples compared to reference 273. Lower Savannah samples compared to references 216 and 242

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

*

** statistically significant for Mann-Whitney and Dunnets

*** statistically significant for Mann-Whitney, Dunnets, and Distribution-free

@ sample mean was statistically different from control

- sample assayed and statistically compared to Redfish Bay (Lower Savannah)

sample assayed and statistically compared to Redfish Bay (Lower Savannah)

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	ANTHRACENE		FLUORANTHENE		PYRENE		BENZ(A)ANTHRAZENE		CHRYSENE		BENZO(B)FLUORANTHENE		BENZO(K)FLUORANTHENE	
			HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	
94-272	H3	Steven Terminal	68.2	52.1	282.2		189.3		218							113.7
94-273	Ref. No. Inlet															67.4
94-274	B9-1	City Channel	25.9	11.1	108		57.1		47.9							81.5
94-275	B9-2	City Channel	23.9	13.7	93.9		40.1		52							93
94-276	B9-3	City Channel	37.5	11.7	102.7		43.4		62.1							48.3
94-277	F1	Dundee Canal	82.6	21.5	226.3		139.2		194.8							28.8
94-278	F2	Dundee Canal	52.5	<	5.7		185.9		122.3							23.3
94-279	B5-3	City Channel	61.4	47.2	173		77.7		95.2							69.7
94-280	B7-1	City Channel	8.3	<	5.7		50.5		22.4							109.6
94-281	B8-1	City Channel	33.4	<	5.7		167.1		71.6							99.4
94-282	B8-2	City Channel	54.8	69.3	339.2		119.5		168.1							93.4
94-283	B2-1	City Channel	16.6	13	69.3		32.8		32.9							39.2
94-284	B2-3	City Channel	59.7	17	174.9		58.4		70.3							13.7
94-285	B2-4	City Channel	122.9	141	488.8		180.7		199.2							44.3
94-286	B8-3	City Channel	51.5	24.2	234.3		105.3		101							312.8
94-287	A2-2	Upper River	11.3	6.9	62.7		26.4		42.9							81.9
94-288	A2-3	Upper River	35.6	22.1	230.2		112.3		123.7							588.9
94-289	A2-4	Upper River	10.6	8.2	107		42.8		61.1							137.3
94-290	B6-1	City Channel	8.3	34	18.6		20.5		121.7							107.8
94-291	B6-2	City Channel	46.2	<	5.7		165.5		79.9							486.1
94-292	B6-3	City Channel	28.6	23.1	185.2		93.6		105.7							42.3
94-293	A1-1	Upper River	8.3	<	5.7		16.1		9.4	<						179
94-294	A1-2	Upper River	8.3	<	5.7		8.2	<	4.8	<						179
94-295	A1-3	Upper River	15.1	<	5.7		33.7		14.6							195.9
94-296	B7-2	City Channel	20.4	17.3	207.6		109.7		101.1							195.9
94-297	B7-3	City Channel	36.9	25.9	238		104.3		74.2							53.7
94-298	B1-1	City Channel	8.3	<	5.7		27.3		8.8	<						7.1
94-299	B1-2	City Channel	9.1	<	5.7	<	7.3		4.8	<						5.8
94-300	B1-5	City Channel	8.3	<	5.7		12.8	<	4.8	<						5.3
94-301	B3-3	City Channel	8.3	<	5.7		29.9		9	<						5.3
94-302	B3-5	City Channel	40.8	26	286.3				7.3							5.3
94-303	B3-6	City Channel	106.2	57	533.9				7.3							5.3
94-304	B4-4	City Channel	10.6	5.8	44.3		19.3		18.8							68.8
94-305	B4-5	City Channel	14.9	<	5.7		114		33.3							61.9
94-306	B4-6	City Channel	24.7	14	167.5		65.2		42.8							8.3
	Ref. Redfish Bay (upper Savannah)															89.6
	Ref. No. Inlet (upper Savannah)															147.3
	Ref. No. Inlet (Lower Savannah)															10.8
	Ref. Redfish Bay (Lower Savannah)															10.7
	Ref. Redfish Bay (Lower Savannah)															70

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	BENZO(A)PYRENE HPLC, ug/kg	PERYLENE HPLC, ug/kg	DIBENZ(A,H)ANTHRACENE HPLC, ug/kg	BENZO(G,H,I)PERYLENE HPLC, ug/kg	Total HPLC, ug/kg	Cl 2(08) ug/kg
			HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg	HPLC, ug/kg
94-216	Ref. No. Inlet							
94-217	E1,2-1	South Channel	7.4	6.4 <	7.3 <	8.2	0 <	0.274
94-218	E1,3-1	South Channel	7.4 <	5.4 <	7.3 <	8.2	74.70402853 <	0.274
94-219	E1-1	South Channel	144.3	70.2	8.7	74.7	93.3457842	0.322
94-220	C1	Back River	19.2	18.3 <	7.3	11.7	11.73703247 <	0.274
94-221	E2-2	South Channel	7.4 <	5.4 <	7.3 <	8.2	0 <	0.274 <
94-222	E2-3	South Channel	7.4	7.4 <	7.3 <	8.2	0 <	0.274 <
94-223	E2-1	South Channel	7.4 <	5.4 <	7.3 <	8.2	0 <	0.274
94-224	C3	Back River	7.4	10.5 <	7.3 <	8.2	0 <	0.274
94-225	C2	Back River	190.8	92.8	12.6	90.6	126.5196818	< 0.274 <
94-226	D1,1-1	Lower Channel	76.9	54.1	8.4	60.6	60.63696075	< 0.274
94-227	D1,2-2	Lower Channel	7.4	6.8 <	7.3 <	8.2	0 <	0.274
94-228	D1,3-1	Lower Channel	14.7	26.6 <	7.3	11.5	11.50237807	< 0.274 <
94-229	D2,1-1	Lower Channel	7.4	11.6 <	7.3 <	8.2	0 <	0.274
94-230	D2,2-2	Lower Channel	7.8	5.4 <	7.3 <	8.2	0 <	0.274
94-231	D2,3-1	Lower Channel	20.2	10 <	7.3	11.1	11.09509339	< 0.278 <
94-242	Ref. No. Inlet							
94-243	E3-2	South Channel	191.6	120.4	11.6	77.2	77.19833564	< 0.274
94-244	E3-5	South Channel	12.8	14.9 <	7.3	9.8	9.802390696	< 0.274 <
94-245	D3,1-4	Lower Channel	7.4 <	5.4 <	7.3 <	8.2	0 <	0.274 <
94-246	D3,2-1	Lower Channel	7.4 <	5.4 <	7.3 <	8.2	0 <	0.274 <
94-247	D3,3-2	Lower Channel	15	11.6 <	7.3 <	8.2	0 <	0.412 <
94-248	E3-9	South Channel	33.6	24.6 <	7.3	20.1	20.09127636	0.368 <
94-266	B5-1	City Channel	69.8	44.3 <	7.3	35.2	35.21346686	< 0.274
94-267	B5-2	City Channel	162.5	114.5	8.7	73.1	73.10540787	< 0.274
94-268	G1	Ocean Terminal	105	75.7 <	7.3	46	115.292258	< 0.274
94-269	G2	Ocean Terminal	56.9	73.9 <	7.3	33	68.50447439	< 0.274 <
94-270	H1	Steven Terminal	323.1	290.5	17.2	168.3	168.2649035	0.553
94-271	H2	Steven Terminal	134	132.1 <	7.3	76.2	126.6337518	0.354

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Ref. No. Inlet	Location	BENZO(A)PYRENE HPLC, ug/kg	PERYLENE HPLC, ug/kg	DIBENZ(A,H)ANTHRACENE HPLC, ug/kg	BENZO(G,H,I)PERYLENE HPLC, ug/kg	Total HPLC, ug/kg	CL2(08) ug/kg
94-272	H3	Steven Terminal	136.5	138.1 <	7.3	7.3	74.4	126.4523685 <
94-273	B9-1	City Channel	58.4	57.2 <	7.3	36.1	47.19480281 <	0.274
94-274	B9-2	City Channel	32.3	43.2 <	7.3	18.6	32.3208937 <	0.274
94-275	B9-3	City Channel	26.9	40.5 <	7.3	17.1	28.7787639 <	0.274
94-276	F1	Dundee Canal	80	130.3 <	7.3	45.8	67.3075505	0.434
94-277	F2	Dundee Canal	100.1	147.4	8.4	68	68.04704586 <	0.274
94-278	B5-3	City Channel	48.5	64.7 <	7.3	28.1	75.25657446	0.366
94-279	B5-4	City Channel	19	23.2 <	7.3	12.3	12.2656924 <	0.274
94-280	B7-1	City Channel	78.8	55.4 <	7.3	48.3	48.34549355	0.738
94-281	B8-1	City Channel	129.6	88.2	7.6	71.4	140.7081765	1.167
94-282	B8-2	City Channel	22.3	37.6 <	7.3	16.2	29.22030313 <	0.274
94-283	B2-1	City Channel	48.5	75.9 <	7.3	34.1	51.15371773	0.796
94-284	B2-3	City Channel	100.1	157.1 <	7.3	56.2	197.2084369 <	0.274
94-285	B2-4	City Channel	82.3	114.6 <	7.3	51.6	75.78501808	0.972
94-286	B8-3	City Channel	42.5	45.3 <	7.3	32.3	39.20609342 <	0.274
94-287	A2-2	Upper River	124.8	79.9 <	7.3	57.8	79.8875617	0.601
94-288	A2-3	Upper River	45.8	49.8 <	7.3	32.7	40.86491843 <	0.274
94-289	A2-4	Upper River	12.3	59.2	39.8	14	47.97504684	1.175
94-290	B6-1	City Channel	58.5	217.5 <	7.3	36.2	36.23795678	1.028
94-291	B6-2	City Channel	75.9	100 <	7.3	42.8	45.88067289	1.219
94-292	B6-3	City Channel	11.5	22 <	7.3 <	8.2	0	1.28
94-293	A1-1	Upper River	7.6	6.5 <	7.3 <	8.2	0	0.48
94-294	A1-2	Upper River	15	32.7 <	7.3	8.7	8.708284969	0.431
94-295	A1-3	Upper River	101	107.3	9.5	56.7	73.96946082	1.644
94-296	B7-2	City Channel	74.6	95.5 <	7.3	41.7	67.68225371	1.714
94-297	B7-3	City Channel	11.9	15.8 <	7.3 <	8.2	0	0.908
94-298	B1-1	City Channel	7.4 <	5.4 <	7.3 <	8.2	0	0.612
94-299	B1-2	City Channel	7.4	6.3 <	7.3 <	8.2	0	0.579
94-300	B1-5	City Channel	7.4	9.2 <	7.3 <	8.2	0	0.406
94-301	B3-3	City Channel	7.4	124.9	8.8	62.2	88.14076794	0.892
94-302	B3-5	City Channel	96.2	183.8	12.2	89.3	146.3592003	2.286
94-303	B3-6	City Channel	15.4	15.1 <	7.3	10.4	16.14645314	0.406
94-304	B4-4	City Channel	16.2	25.3 <	7.3	8.8	8.801542087	
94-305	B4-5	City Channel	76.6	52.1 <	7.3	40.7	54.63977922	0.334
94-306	B4-6	City Channel						
		Ref. Redfish Bay (upper Savannah)						
		Ref. No. Inlet (upper Savannah)						
		Ref. No. Inlet (Lower Savannah)						
		Ref. Redfish Bay (Lower Savannah)						

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	HEXACHLOROBENZENE ug/kg	LINDANE ug/kg	CL3(18) ug/kg	HEPTACHLOR ug/kg	CL4(28) ug/kg	ALDRIN ug/kg	CL4(44) ug/kg	HEPTACHLOREPOXIDE ug/kg
94-216	Ref. No. Inlet									
94-217	E1,2-1	South Channel	0.487 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-218	E1,3-1	South Channel	0.324 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-219	E1-1	South Channel	0.469 <	0.145 <	0.253 <	0.454 <	0.167 <	0.669 <	0.22	0.229 <
94-220	C1	Back River	0.228	0.17 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.144 <
94-221	E2-2	South Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-222	E2-3	South Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-223	E2-1	South Channel	0.146 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-224	C3	Back River	0.09 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-225	C2	Back River	0.084	0.399 <	0.253 <	0.454 <	0.167	1.332 <	0.22	0.321 <
94-226	D1,1-1	Lower Channel	0.313	0.174 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.155 <
94-227	D1,2-2	Lower Channel	0.201 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-228	D1,3-1	Lower Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.083 <
94-229	D2,1-1	Lower Channel	0.105 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-230	D2,2-2	Lower Channel	0.094 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.068 <
94-231	D2,3-1	Lower Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.274 <
94-242	Ref. No. Inlet									
94-243	E3-2	South Channel	0.144 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.06 <
94-244	E3-5	South Channel	0.084	0.203 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.246 <
94-245	D3,1-4	Lower Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-246	D3,2-1	Lower Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.058 <
94-247	D3,3-2	Lower Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.133 <
94-248	E3-9	South Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.173 <
94-266	B5-1	City Channel	0.185	0.259 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-267	B5-2	City Channel	0.255	0.36 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-268	G1	Ocean Terminal	0.219 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-269	G2	Ocean Terminal	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22 <	0.042 <
94-270	H1	Steven Terminal	0.262	0.64 <	0.253 <	0.454 <	0.167 <	0.204 <	0.28 <	0.21 <
94-271	H2	Steven Terminal	0.186 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22	0.205 <

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.									
MSL Lab. Id. No.	STATION Number	Location	HEXACHLOROBENZENE ug/kg	LINDANE ug/kg	CL3(18) ug/kg	HEPTACHLOR ug/kg	CL4(52) ug/kg	ALDRIN ug/kg	CL4(44) ug/kg
94-272	H3	Steven Terminal	0.156 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-273	Ref. No. Inlet								0.21
94-274	B9-1	City Channel	0.084	0.255 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-275	B9-2	City Channel	0.131	0.326 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-276	B9-3	City Channel	0.136	0.28 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-277	F1	Dundee Canal	0.553 <	0.145 <	0.253 <	0.454 <	0.167	0.629 <	0.22
94-278	F2	Dundee Canal	0.345	0.575 <	0.253	0.585 <	0.167	0.858 <	0.22
94-279	B5-3	City Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167	0.473 <	0.22
94-280	B7-1	City Channel	0.084	0.227 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-281	B8-1	City Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-282	B8-2	City Channel	0.084 <	0.145 <	0.253	0.785 <	0.167 <	0.204 <	0.22
94-283	B2-1	City Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-284	B2-3	City Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-285	B2-4	City Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-286	B8-3	City Channel	0.084 <	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-287	A2-2	Upper River	0.141	0.205 <	0.253	0.454 <	0.167 <	0.204 <	0.22
94-288	A2-3	Upper River	0.155	0.269 <	0.253	0.454 <	0.167 <	0.204 <	0.22
94-289	A2-4	Upper River	0.181 <	0.145 <	0.253	0.454 <	0.167 <	0.204 <	0.22
94-290	B6-1	City Channel	0.474	0.338	0.547	3.148 <	0.167 <	0.204 <	0.22
94-291	B6-2	City Channel	0.236	0.432 <	0.253	0.454 <	0.167 <	0.204 <	0.22
94-292	B6-3	City Channel	0.084 <	0.145 <	0.253	0.454 <	0.167 <	0.204 <	0.22
94-293	A1-1	Upper River	0.098	0.145 <	0.253	0.454 <	0.333 <	0.204 <	0.22
94-294	A1-2	Upper River	0.084 <	0.145 <	0.253	0.454 <	0.948 <	0.204 <	0.22
94-295	A1-3	Upper River	0.084	0.225	0.321 <	0.454 <	0.502	0.32 <	0.418 <
94-296	B7-2	City Channel	0.326 <	0.145 <	0.253	0.454 <	2.765 <	0.204 <	0.22
94-297	B7-3	City Channel	0.23 <	0.145 <	0.253	0.454 <	1.545 <	0.708	0.319 <
94-298	B1-1	City Channel	0.084	0.145 <	0.253	0.454 <	1.471	0.218 <	0.22
94-299	B1-2	City Channel	0.084 <	0.145 <	0.253	0.454 <	0.925 <	0.204 <	0.22
94-300	B1-5	City Channel	0.084 <	0.145 <	0.253	0.454 <	2.607 <	0.204 <	0.22
94-301	B3-3	City Channel	0.084 <	0.145 <	0.402 <	0.454 <	0.584 <	0.204 <	0.22
94-302	B3-5	City Channel	0.084	0.843	0.869 <	0.454 <	0.707	0.7 <	0.22
94-303	B3-6	City Channel	0.084 <	0.145 <	0.601 <	0.454 <	0.426	0.892 <	0.22
94-304	B4-4	City Channel	0.084	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-305	B4-5	City Channel	0.139	0.278 <	0.253 <	0.454 <	0.167 <	0.204 <	0.22
94-306	B4-6	City Channel							0.096 <

Ref. Redfish Bay (upper Savannah)

Ref. No. Inlet (upper Savannah)

Ref. No. Inlet (Lower Savannah)

Ref. Redfish Bay (Lower Savannah)

2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

STATION Number	Location	CL4(66) ug/kg	2,4-DDE ug/kg	CL5(101) ug/kg	Alpha Chlordane ug/kg	TRANS-NONACHLOR ug/kg	DiELDRIN ug/kg	4,4-DDE ug/kg	CL4(77) ug/kg	2,4-DDD ug/kg
Ref. No. Inlet E1.2-1	South Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
E1.3-1	South Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
E1-1	South Channel	0.11 <	0.217	0.33 <	0.359 <	0.345 <	0.403 <	0.371 <	0.524 <	0.794 <
C1	Back River	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.465 <	0.794 <
E2-2	South Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
E2-3	South Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
E2-1	South Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
C3	Back River	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
C2	Back River	0.039 <	0.217	0.227 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
D1,1-1	Lower Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.33 <	0.794 <
D1,2-2	Lower Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
D1,3-1	Lower Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.251 <	0.794 <
D2,1-1	Lower Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
D2,2-2	Lower Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
D2,3-1	Lower Channel	0.419 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
Ref. No. Inlet E3-2	South Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
E3-5	South Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
D3,1-4	Lower Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
D3,2-1	Lower Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
D3,3-2	Lower Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
E3-9	South Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
B5-1	City Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.675 <	0.403 <	0.371 <	0.19 <	0.794 <
B5-2	City Channel	0.039 <	0.217	0.435 <	0.435 <	1.025 <	0.403 <	0.371 <	0.19 <	0.794 <
G1	Ocean Terminal	0.039 <	0.217	0.241 <	0.373 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
G2	Ocean Terminal	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.19 <	0.794 <
H1	Steven Terminal	0.309 <	0.217	0.375 <	0.375 <	0.502 <	0.403 <	0.371 <	0.19 <	0.794 <
H2	Steven Terminal	0.039 <	0.217	0.262 <	0.359 <	0.345 <	0.403 <	0.371 <	1.157 <	0.794 <

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Ref. No.	Location	CL4(66) ug/kg	2,4-DDE ug/kg	CL5(101) ug/kg	Alpha Chlordane ug/kg	TRANS-NONACHLOR ug/kg	DIELDRIN ug/kg	4,4-DDE ug/kg	CL4(77) ug/kg	2,4-DDD ug/kg
94-272	H3	Steven Terminal	0.183 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371	0.439 <	0.794 <
94-273											
94-274	B9-1	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.79 <	0.403 <	0.371	< 0.19	< 0.794
94-275	B9-2	City Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-276	B9-3	City Channel	0.039 <	0.217	0.171 <	0.359 <	0.345 <	0.403 <	0.371	0.692 <	0.794 <
94-277	F1	Dundee Canal	0.039 <	0.217	0.561	0.561	0.422 <	0.403 <	0.371	1.671	0.794 <
94-278	F2	Dundee Canal	0.757 <	0.217	1.619	0.764	0.387 <	0.403 <	0.371	5.062 <	0.794 <
94-279	B5-3	City Channel	0.33 <	0.217	0.273 <	0.359 <	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-280	B7-1	City Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-281	B8-1	City Channel	0.039 <	0.217	0.154 <	0.359	1.418 <	0.403 <	0.371	< 0.19	< 0.794
94-282	B8-2	City Channel	0.039 <	0.217	0.243 <	0.359	1.678 <	0.403 <	0.371	< 0.19	< 0.794
94-283	B2-1	City Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-284	B2-3	City Channel	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-285	B2-4	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.996 <	0.403 <	0.371	< 0.19	< 0.794
94-286	B8-3	City Channel	0.039 <	0.217	0.17 <	0.359	1.181 <	0.403 <	0.371	0.936 <	0.794 <
94-287	A2-2	Upper River	0.039 <	0.217	0.555 <	0.359 <	0.345 <	0.403 <	0.371	1.732 <	0.794 <
94-288	A2-3	Upper River	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371	0.318 <	0.794 <
94-289	A2-4	Upper River	0.039 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-290	B6-1	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-291	B6-2	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.481 <	0.403 <	0.371	0.615 <	0.794 <
94-292	B6-3	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-293	A1-1	Upper River	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-294	A1-2	Upper River	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-295	A1-3	Upper River	2.3075 <	0.217	1.144	0.896	0.345 <	0.562	2.229	4.755 <	0.794 <
94-296	B7-2	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-297	B7-3	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-298	B1-1	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-299	B1-2	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-300	B1-5	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-301	B3-3	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-302	B3-5	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-303	B3-6	City Channel	0.039 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-304	B4-4	City Channel	0.148 <	0.217 <	0.092 <	0.359	0.345 <	0.403 <	0.371	< 0.19	< 0.794
94-305	B4-5	City Channel									
94-306	B4-6	City Channel	0.039 <	0.217	0.669	0.504	0.513 <	0.403 <	0.371	0.309 <	0.794
		Ref. Redfish Bay (upper Savannah)									
		Ref. No. Inlet (upper Savannah)									
		Ref. No. Inlet (lower Savannah)									
		Ref. Redfish Bay (lower Savannah)									

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Ref. No.	Location	CL5(118) ug/kg	4,4-DDD ug/kg	2,4-DDT ug/kg	CL6(153) ug/kg	CL5(105) ug/kg	4,4-DDT ug/kg	CL6(138) ug/kg	CL5(126) ug/kg	CL7(187) ug/kg	CL6(128) ug/kg
94-216	Ref. No. Inlet											
94-217	E1,2-1	South Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-218	E1,3-1	South Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-219	E1-1	South Channel	0.287 <	0.416 <	0.382 <	0.562 <	0.086 <	0.367 <	0.318 <	0.215 <	0.06 <	0.089 <
94-220	C1	Back River	0.18 <	0.416 <	0.382 <	0.599 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-221	E2-2	South Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-222	E2-3	South Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-223	E2-1	South Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-224	C3	Back River	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-225	C2	Back River	0.314 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-226	D1,1-1	Lower Channel	0.086 <	0.416 <	0.382 <	0.47 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-227	D1,2-2	Lower Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-228	D1,3-1	Lower Channel	0.086 <	0.416 <	0.382 <	0.314 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-229	D2,1-1	Lower Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-230	D2,2-2	Lower Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-231	D2,3-1	Lower Channel	0.092 <	0.416 <	0.382 <	0.204 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-242	Ref. No. Inlet											
94-243	E3-2	South Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-244	E3-5	South Channel	0.086 <	0.416 <	0.382 <	0.605 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-245	D3,1-4	Lower Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-246	D3,2-1	Lower Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-247	D3,3-2	Lower Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215 <	0.06 <	0.05 <
94-248	E3-9	South Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.286 <	0.215 <	0.06 <	0.05 <
94-266	B5-1	City Channel	0.086 <	0.416 <	0.382 <	0.722 <	0.086 <	0.367 <	0.248 <	0.215 <	0.06 <	0.05 <
94-267	B5-2	City Channel	0.086 <	0.416 <	0.382 <	1.103 <	0.086 <	0.367 <	0.405 <	0.215 <	0.06 <	0.05 <
94-268	G1	Ocean Terminal	0.086 <	0.416 <	0.382 <	0.32 <	0.086 <	0.367 <	0.443 <	0.215 <	0.06 <	0.05 <
94-269	G2	Ocean Terminal	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.454 <	0.06 <	0.05 <
94-270	H1	Steven Terminal	0.466	0.742 <	0.382 <	1.422 <	0.086 <	0.367 <	0.509 <	0.215 <	0.06 <	0.05 <
94-271	H2	Steven Terminal	0.216 <	0.416 <	0.382 <	1.249 <	0.086 <	0.367 <	0.343 <	0.215 <	0.06 <	0.05 <

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Number	Location	CL5(118) ug/kg	4,4-DDD ug/kg	2,4-DDT ug/kg	CL6(153) ug/kg	CL5(105) ug/kg	4,4-DDT ug/kg	CL6(138) ug/kg	CL5(126) ug/kg	CL7(187) ug/kg	CL6(128) ug/kg
94-272	H3	Steven Terminal	0.086 <	0.416 <	0.382	0.355 <	0.086 <	0.367	0.239 <	0.215 <	0.06 <	0.05
94-273	Ref. No. Inlet											
94-274	B9-1	City Channel	0.086 <	0.416 <	0.382	0.32 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05 <
94-275	B9-2	City Channel	0.086 <	0.416 <	0.382	0.443 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05 <
94-276	B9-3	City Channel	0.086 <	0.416 <	0.382	0.997 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05
94-277	F1	Dundee Canal	0.183	0.497 <	0.382	2.69 <	0.086 <	0.367	0.748 <	0.215 <	0.06 <	0.05 <
94-278	F2	Dundee Canal	1.382	2.399 <	0.382	3.833	0.728	1.843	3.183 <	0.215	0.517 <	0.05 <
94-279	B5-3	City Channel	0.086 <	0.416 <	0.382	0.956 <	0.086 <	0.367	0.288 <	0.215 <	0.06 <	0.05 <
94-280	B7-1	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.309 <	0.215 <	0.06 <	0.05 <
94-281	B8-1	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.632	0.632	1.185 <	0.06 <	0.05 <
94-282	B8-2	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	1.229	1.229	4.162 <	0.06 <	0.05 <
94-283	B2-1	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05 <
94-284	B2-3	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.353 <	0.215 <	0.06 <	0.05 <
94-285	B2-4	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.665	1.572 <	0.06 <	0.05 <
94-286	B8-3	City Channel	0.086 <	0.416 <	0.382	1.46 <	0.086 <	0.367	0.329 <	0.215 <	0.06 <	0.05 <
94-287	A2-2	Upper River	0.596	0.995 <	0.382	1.693 <	0.086 <	0.367	1.352 <	0.215 <	0.231	0.308
94-288	A2-3	Upper River	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.398 <	0.215 <	0.06 <	0.05 <
94-289	A2-4	Upper River	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.721	0.721	3.688 <	0.06 <	0.05 <
94-290	B6-1	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.232	2.573 <	0.06 <	0.05 <
94-291	B6-2	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.354 <	0.215 <	0.06 <	0.05 <
94-292	B6-3	City Channel	0.086 <	0.416 <	0.382	0.509 <	0.086 <	0.367	0.362 <	0.215 <	0.06 <	0.05 <
94-293	A1-1	Upper River	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.232	0.385 <	0.06 <	0.05 <
94-294	A1-2	Upper River	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05 <
94-295	A1-3	Upper River	1.082	2.976 <	0.382	2.436 <	0.086	2.0145 <	0.215 <	0.661 <	0.05 <	
94-296	B7-2	City Channel	0.086 <	0.416 <	0.382	0.524 <	0.086	0.772	0.753 <	0.215 <	0.06 <	0.05 <
94-297	B7-3	City Channel	0.086 <	0.416 <	0.382	2.186 <	0.086	0.742	0.742	0.506 <	0.06 <	0.05 <
94-298	B1-1	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086	0.819	0.508 <	0.215 <	0.06 <	0.05 <
94-299	B1-2	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05 <
94-300	B1-5	City Channel	0.086 <	0.416 <	0.382 <	0.124 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05 <
94-301	B3-3	City Channel	0.086 <	0.416 <	0.382	0.183 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05 <
94-302	B3-5	City Channel	0.086 <	0.416 <	0.382	2.122 <	0.086	1.073	0.937	0.501 <	0.06 <	0.05 <
94-303	B3-6	City Channel	0.086 <	0.416 <	0.382	3.501 <	0.086	0.735	0.735 <	0.215 <	0.06 <	0.05 <
94-304	B4-4	City Channel	0.086 <	0.416 <	0.382	0.371 <	0.086 <	0.367	0.232 <	0.215 <	0.06 <	0.05 <
94-305	B4-5	City Channel	0.443	0.994 <	0.382	2.082	0.305 <	0.367	1.493 <	0.215	0.262	0.208
94-306	B4-6	City Channel										
		Ref. Redfish Bay (upper Savannah)										
		Ref. No. Inlet (upper Savannah)										
		Ref. No. Inlet (Lower Savannah)										
		Ref. Redfish Bay (Lower Savannah)										

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Ref. No. Inlet	Location	CL7(180) ug/kg	MIREX ug/kg	CL7(170) ug/kg	CL8(195) ug/kg	CL9(206) ug/kg	CL10(209) ug/kg	Total of congeners ug/kg	X2 total PCBs ug/kg	Pesticide Total : DDT Total : ug/kg
94-216											
94-217	E1,2-1	South Channel	0.036 <	0.212 <	0.127 <	0.088	0.212 <	0.069	0.212	2	0.424731
94-218	E1,3-1	South Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.000	2	2.385
94-219	E1-1	South Channel	0.036 <	0.212 <	0.127 <	0.088	0.363	0.077	3.880	2	7.760415
94-220	C1	Back River	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	1.388	2	2.776119
94-221	E2-2	South Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.000	2	0
94-222	E2-3	South Channel	0.036 <	0.212 <	0.127 <	0.088	0.48 <	0.069	0.480	2	0.959233
94-223	E2-1	South Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.000	2	0
94-224	C3	Back River	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.000	2	0
94-225	C2	Back River	0.036 <	0.212 <	0.127 <	0.088	0.472 <	0.069	2.666	2	5.332078
94-226	D1,1-1	Lower Channel	0.036 <	0.212 <	0.127 <	0.088	0.215 <	0.069	1.170	2	2.340191
94-227	D1,2-2	Lower Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.000	2	0
94-228	D1,3-1	Lower Channel	0.036 <	0.212 <	0.127 <	0.088	0.265 <	0.069	0.912	2	1.824521
94-229	D2,1-1	Lower Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.000	2	0
94-230	D2,2-2	Lower Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.068	2	0.136599
94-231	D2,3-1	Lower Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	1.266	2	2.532251
94-242	Ref. No. Inlet										
94-243	E3-2	South Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.060	2	0.119696
94-244	E3-5	South Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.851	2	1.701516
94-245	D3-1-4	Lower Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.000	2	0
94-246	D3-2-1	Lower Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.058	2	0.1115323
94-247	D3-3-2	Lower Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.545	2	1.090501
94-248	E3-9	South Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.828	2	1.655559
94-266	B5-1	City Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.969	2	1.938717
94-267	B5-2	City Channel	0.036	1.756 <	0.127 <	0.088 <	0.158 <	0.069	1.944	2	3.887884
94-268	G1	Ocean Terminal	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	1.004	2	2.007643
94-269	G2	Ocean Terminal	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.454	2	0.908081
94-270	H1	Steven Terminal	0.036 <	0.212 <	0.127 <	0.088 <	0.269 <	0.069	4.183	2	8.365163
94-271	H2	Steven Terminal	0.092 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	3.879	2	7.757221

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

MSL Lab. Id. No.	STATION Ref. No. Inlet	Location	CL7(180) ug/kg	MIREX ug/kg	CL7(170) ug/kg	CL8(195) ug/kg	CL9(206) ug/kg	CL10(209) ug/kg	Total of congeners X2 ug/kg	total PCBs ug/kg	Pesticide Total : ug/kg	DDT Total : ug/kg
94-272	H3	Steven Terminal	0.059 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	1.383	2	2.766366	2.217
94-273	B9-1	City Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.320	2	0.641	2.7
94-274	B9-2	City Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.443	2	0.886639	2.373
94-275	B9-3	City Channel	0.104 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	2.063	2	4.126619	2.332
94-276	F1	Dundee Canal	0.036 <	0.235 <	0.127 <	0.088 <	0.158 <	0.069	6.916	2	13.83289	2.916
94-277	F2	Dundee Canal	0.036 <	0.263 <	0.127 <	0.12	0.423	0.179	19.666	2	39.33115	3.334
94-278	B5-3	City Channel	0.036 <	0.212 <	0.127 <	0.088	0.321	0.312	3.631	2	7.262793	2.145
94-279	B7-1	City Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.309	2	0.61769	2.227
94-280	B8-1	City Channel	0.036 <	0.281 <	0.127 <	0.088	0.426 <	0.069	3.135	2	6.269469	3.287
94-281	B8-2	City Channel	0.036 <	0.537 <	0.127 <	0.088	0.506 <	0.069	8.091	2	16.18238	3.803
94-282	B2-1	City Channel	0.036 <	0.212 <	0.127 <	0.088	0.193 <	0.069	0.193	2	0.386976	2.145
94-283	B2-3	City Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	1.149	2	2.298706	2.145
94-284	B2-4	City Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	2.236	2	4.472303	2.796
94-285	B8-3	City Channel	0.036 <	0.212 <	0.127 <	0.088	0.256 <	0.069	4.259	2	8.517541	2.981
94-286	A2-2	Upper River	0.755	0.944 <	0.127 <	0.088	0.277 <	0.069	7.718	2	15.43569	2.994
94-287	A2-3	Upper River	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	1.317	2	2.633506	2.34
94-288	A2-4	Upper River	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	4.409	2	8.818139	2.242
94-289	B6-1	City Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	7.443	2	14.88662	2.728
94-290	B6-2	City Channel	0.036 <	0.212 <	0.127 <	0.088	0.208 <	0.069	2.205	2	4.410428	2.72
94-291	B6-3	City Channel	0.036 <	0.212 <	1.344 <	0.088 <	0.158 <	0.069	4.420	2	8.840862	2.244
94-292	A1-1	Upper River	0.254 <	0.212 <	0.127 <	0.112 <	0.158 <	0.069	2.032	2	4.064022	2.325
94-293	A1-2	Upper River	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.480	2	0.959174	2.926
94-294	A1-3	Upper River	0.036 <	0.603	0.233 <	0.088	0.565 <	0.069	14.641	2	29.28298	3.647
94-295	B7-2	City Channel	0.036 <	0.212	0.831 <	0.088 <	0.158 <	0.069	3.752	2	7.503946	4.985
94-296	B7-3	City Channel	0.036 <	0.212	1.326 <	0.088	0.163 <	0.069	7.698	2	15.39593	3.669
94-297	B1-1	City Channel	0.036 <	0.212	0.182 <	0.088 <	0.158 <	0.069	1.940	2	3.888056	3.449
94-298	B1-2	City Channel	0.036 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.612	2	1.223036	2.903
94-299	B1-5	City Channel	0.068 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	0.647	2	1.293402	4.585
94-300	B3-3	City Channel	0.045 <	0.212	0.148 <	0.088 <	0.158 <	0.069	1.238	2	2.475429	2.562
94-301	B3-5	City Channel	0.418 <	0.212	0.884 <	0.088 <	0.158 <	0.109	8.016	2	16.03106	3.428
94-302	B3-6	City Channel	0.571 <	0.212	0.684 <	0.088 <	0.158 <	0.069	9.551	2	19.10134	2.412
94-303	B4-4	City Channel	0.068 <	0.212 <	0.127 <	0.088 <	0.158 <	0.069	1.042	2	2.084594	2.145
94-304	B4-5	City Channel	0.687 <	0.212 <	0.127 <	0.088	0.578	0.096	7.564	2	15.12722	2.646
94-305	B4-6	City Channel										3.125
Ref. Redfish Bay (upper Savannah)												
Ref. No. Inlet (upper Savannah)												
Ref. No. Inlet (Lower Savannah)												
Ref. Redfish Bay (Lower Savannah)												

Appendix B2 continued. Toxicity and chemistry data from upper and lower Savannah River sediments.

94-271 H2

MSI samples 94-266 to 94-306 collected from Upper Savannah.

MSL samples 94-217 to 94-248 samples collected from Lower Savannah.

Upper Savannah samples compared to those from significant

non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test

* Statistically significant for Mann-Whitney

*** statistically significant for Mann-Whitney and Dunn

**** statistically significant for Mann-Whitney, Dunnett's

sample mean was statistically different from control

Appendix B3. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	Location	% Amph surv. as % of control	Amph Surv Signif. (% of sample)	Microtox EC50 (mg/ml)	Microtox EC50 %ctl.	Signif. (216 & 242)	% urch fert. in 100% pw.	Signif.
94-191	A1-1	Upper Turtle R.	89	95	37.79	2.68	323.74	ns	98.8
94-192	A2-2	Upper Turtle R.	90	96	51	3.70	446.37	ns	99.0
94-193	A3-1	Upper Turtle R.	89	95	8.55	0.63	76.41	ns	100.0
94-194	D1-1	Blythe Isl. Range	92	98	46.71	3.45	416.80	ns	99.4
94-195	D2-1	Blythe Isl. Range	92	98	26.76	2.00	237.72	ns	99.8
94-196	P1	Purvis Cr.	92	98	21.49	1.61	194.38	ns	99.4
94-197	B2-1	Academy Cr.	94	100	7.8791	0.58	70.48	ns	93.4
94-198	C1-1	East River Harbor	95	101	3.3155	0.25	29.98	ns	23.0
94-199	B1-1	Academy Cr.	92	98	3.6969	0.28	33.02	ns	14.8
94-200	C3-1	East River Harbor	95	101	4.2	3.27	394.23	ns	75.4
94-201	C2-1	East River Harbor	99	105	2.43	0.17	20.94	**	82.6
94-202	D3-2	Blythe Isl. Range	90	96	51	3.80	458.29	ns	99.6
94-203	E1-1	Brunswick Pt. Range	89	95	6.3034	0.46	55.88	ns	99.0
94-204	E2-2	Brunswick Pt. Range	91	97	2.3216	0.17	20.42	**	99.0
94-210	E3-1	Brunswick Pt. Range	80	85	22.28	1.68	203.18	ns	99.8
94-211	F1-1	Terry Cr.	81	86	@	0.1223	0.01	1.11	**
94-212	F2-1	Dupree Cr.	0	0	@~	1.528	0.12	14.05	**
94-213	G1-1	Back River	96	103	ns	0.325	0.02	2.79	**
94-214	G2-1	Back River	92	99	73.9363	5.32	641.96	ns	99.8
94-215	G3-1	Back River	89	96	13.064	1.00	121.16	ns	99.8
94-216	Ref. No. Inlet				11.2882	0.83	100.84	ns	
94-242	Ref. No. Inlet				10.7696	0.82	0.99	ns	99.3
	Ref. Redfish Bay							99.6	ns
	Ref. No. Inlet								
Duplicates for averaged result represented above									
94-204	E2-2				AVS				
94-204	E2-2				6.34				
					5.8				

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

*

statistically significant for Mann-Whitney and Dunnets

*** statistically significant for Mann-Whitney, Dunnets, and Distribution-free

@ sample mean was statistically different from control

- sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	% urch fert. in 50% pw.	Signif. in 25% pw.	% urch fert. in 100% pw.	Signif. in 50% pw.	% urchdevp in 100% pw.	Signif. in 50% pw.	% urchdevp in 25% pw.	Signif. in 50% pw.	% urchdevp in 25% pw.	Signif. in 50% pw.	UAN, amph start mg/l	UAN, amph end mg/l	UAN, amph ave. mg/l
94-191	A1 -1	99.6	ns	99.6	ns	0	++	52.8	++	98.6	ns	.028		
94-192	A2 -2	99.4	ns	99.0	ns	96.6	ns	97.80	ns	98.2	ns	.011		
94-193	A3 -1	100.0	ns	99.4	ns	63.4	++	96.4	ns	97.4	ns	.010		
94-194	D1 -1	99.2	ns	99.6	ns	58	++	96.6	ns	97.6	ns	.012		
94-195	D2 -1	99.2	ns	99.4	ns	37.25	++	95.6	ns	96.4	ns	.014		
94-196	P1	99.6	ns	99.8	ns	98	ns	99	ns	98.4	ns	.011		
94-197	B2 -1	99.2	ns	99.6	ns	0	++	85.4	ns	98.4	ns	.019		
94-198	C1 -1	93.2	ns	99.2	ns	3.25	++	0.004	++	7.4	++	.044		
94-199	B1 -1	65.4	++	99.0	ns	2.6	++	0	++	61.6	++	.031		
94-200	C3-1	98.2	ns	99.6	ns	1	++	0	++	56.6	++	.076		
94-201	C2 -1	98.6	ns	99.6	ns	0	++	0	++	5.75	++	.070		
94-202	D3 -2	99.2	ns	99.0	ns	96.4	ns	97.4	ns	98.4	ns	.009		
94-203	E1 -1	100.0	ns	99.8	ns	3.8	++	98.2	ns	98.4	ns	.024		
94-204	E2 -2	99.8	ns	98.8	ns	87.8	ns	97.4	ns	98.5	ns	.012		
94-210	E3-1	100.0	ns	99.2	ns	98.6	ns	98.2	ns	98.6	ns	.016		
94-211	F1-1	99.4	ns	99.4	ns	1.6	++	60.6	++	98.4	ns	.046		
94-212	F2-1	99.8	ns	99.2	ns	18.5	++	97.6	ns	98.4	ns	.038		
94-213	G1-1	98.4	ns	100.0	ns	5.4	++	0	++	95.8	ns	.077		
94-214	G2-1	100.0	ns	99.6	ns	96.6	ns	97.2	ns	99	ns	.031		
94-215	G3-1	99.6	ns	97.2	ns	98.6	ns	98.6	ns	98.4	ns	.006		
94-216	Ref. No. Inlet													
94-242	Ref. No. Inlet													
	Ref. Redfish Bay	99.0	ns	99.3	ns	99.2	ns	98.4	ns	98.3	ns			
	Ref. No. Inlet	100.0	ns	99.8	ns	31.2	++	99	ns	99.4	ns	.131		
	Duplicates for averaged result represented above													
94-204	E2-2													
94-204	E2-2													

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnets, and Distribution-free

@ sample mean was statistically different from control

~ sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	Pwater UAN ug/l	Ag ($\mu\text{g/g}$)	Al (%)	As ($\mu\text{g/g}$)	Cd ($\mu\text{g/g}$)	Cr ($\mu\text{g/g}$)	Cu ($\mu\text{g/g}$)	Fe (%)	Hg ($\mu\text{g/g}$)	Mn ($\mu\text{g/g}$)	Ni ($\mu\text{g/g}$)	Pb ($\mu\text{g/g}$)	Se ($\mu\text{g/g}$)						
94-191	A1 -1	37.34	0.04	0.35	2.1	0.110773751	10.0	1.4	0.28	.056	117	2.810989638	2.8	0.161606268						
94-192	A2 -2	8.93	0.03	1.57	3.3	<	0.03	31.6	2.7	1.11	.064	112	6.097079716	6.4	0.125295975					
94-193	A3 -1	17.67	0.03	0.49	1.5	<	0.03	17.7	1.4	0.29	.150	61	<	1.9	4.5	0.127668063				
94-194	D1 -1	17.55	0.04	0.33	1.0	0.035749851	13.8	0.9	0.21	.086	30	2.280422227	2.4	0.06970723	<					
94-195	D2 -1	26.77	0.03	0.27	5.7	0.064015905	9.5	0.9	0.87	.046	67	7.614314115	2.2	0.040755467	<					
94-196	P1	10.40	0.05	3.30	3.6	0.099000999	45.4	10.7	1.66	.188	11.968803197	20.6	0.365634366							
94-197	B2 -1	26.54	0.33	4.97	9.2	0.283630311	61.6	24.6	2.46	.280	278	17.9067946	40.1	0.231055414						
94-198	C1 -1	92.38	0.13	5.00	10.2	0.150338511	84.0	27.5	2.53	.180	361	19.57387495	28.7	0.318598168						
94-199	B1 -1	99.49	0.13	5.66	9.1	0.219976219	89.5	35.2	2.89	.140	242	20.63020214	32.5	0.268529528						
94-200	C3-1	20.71	0.06	5.43	15.4	<	0.03	83.1	14.4	2.72	.260	1269	19.58651773	9.6	0.531060161					
94-201	C2 -1	24.71	0.08	5.55	10.7	<	0.03	83.6	14.8	2.82	.120	896	20.49732514	10.2	0.518129582					
94-202	D3 -2	11.59	<	0.02	0.24	1.4	<	0.03	5.0	0.7	0.15	<	.010	49	<	1.9	1.6	<	0.03	<
94-203	E1 -1	48.68	<	0.02	2.27	7.7	<	0.03	37.3	6.1	1.26	.080	264	8.4772027271	8.3	0.32985763				
94-204	E2 -2	27.52	<	0.02	2.27	6.2	<	0.03	38.3	5.8	1.29	.100	187	7.989183874	8.4	0.355495046				
94-210	E3-1	14.34	<	0.02	1.27	4.97	0.052464858	15.5	2.4	0.79	.041	165	3.920015838	4.9	0.204909919					
94-211	F1-1	69.08	<	0.02	6.09	16.40	<	0.32	74.9	16.4	3.11	.130	539	22.44639376	23.0	<	0.03			
94-212	F2-1	34.38	0.48	5.12	12.62	0.06441069	66.8	45.0	2.52	.160	275	21.833233993	31.7	<	0.03					
94-213	G1-1	61.55	0.04	5.74	15.20	0.044010716	71.7	10.1	2.88	.140	532	21.02946804	20.4	<	0.03					
94-214	G2-1	30.73	0.02	0.67	3.46	0.040919719	9.6	0.8	0.54	.018	87	<	1.9	3.2	<	0.03				
94-215	G3-1	4.09	<	0.02	0.75	14.81	<	0.32	9.0	0.8	1.61	.237	2.382330247	3.8	<	0.03				
94-216	Ref. No. Inlet																			
94-242	Ref. No. Inlet																			
94-204	Ref. Redfish Bay	14.43																		
94-204	Ref. No. Inlet	44.9																		

Duplicates for averaged result represented above

94-204 E2-2
94-204 E2-2

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

* statistically significant for Mann-Whitney

** statistically significant for Mann-Whitney and Dunnets

*** statistically significant for Dunnets, and Distribution-free

@ sample mean was statistically different from control

~ sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	Sn ($\mu\text{g/g}$)	Zn ($\mu\text{g/g}$)	% sand			% clay	% fines	% TOC	Cd ($\mu\text{mole/g}$)	Cu ($\mu\text{mole/g}$)	Ni ($\mu\text{mole/g}$)	Pb ($\mu\text{mole/g}$)
				% silt	% clay	% fines							
94-191	A1 -1	9.794319295	6.8	94.08	1.75	4.17	5.92	0.40	0.001	*	0.004	*	<0.010 *
94-192	A2 -2	15.390686666	22.9	76.71	5.59	17.7	23.29	0.47	<0.002	*	0.009	*	<0.033 *
94-193	A3 -1	9.375623379	8.3	91.7	0.74	7.56	8.3	0.32	0.007	*	<0.010	*	<0.034 *
94-194	D1 -1	7.8	8.0	97.36	0.37	2.27	2.64	0.16	0.001	*	<0.002	*	<0.065 *
94-195	D2 -1	7.8	8.4	94.41	0.95	4.64	5.59	0.17	0.001	*	0.028	0.015	<0.013 *
94-196	P1	31.26873127	44.3	84.48	2.61	12.92	15.53	1.46	0.002	*	<0.004	*	<0.011 *
94-197	B2 -1	44.93831995	98.1	17.68	15.78	66.54	82.32	2.52	<0.006	*	0.114	<0.088 *	<0.090 *
94-198	C1 -1	43.70768618	94.5	4.66	13.97	81.37	95.34	2.81	0.018	*	<0.030	*	<0.187 *
94-199	B1 -1	47.95877923	111.8	10.31	18.64	71.04	89.68	3.82	<0.007	*	<0.019	*	<0.181 *
94-200	C3 -1	33.21575544	81.8	0.2	15.25	84.55	99.8	3.58	<	0.004	*	<0.005	*
94-201	C2 -1	32.29641371	84.3	7.23	21.57	71.2	92.77	3.36	0.012	*	<0.023	*	<0.121 *
94-202	D3 -2	7.8	4.4	94.73	1.12	4.15	5.27	0.15	<0.001	*	<0.003	*	<0.016 *
94-203	E1 -1	13.03388811	34.4	61.01	12.33	26.66	38.99	1.91	0.015	*	<0.023	*	<0.141 *
94-204	E2 -2	15.12744876	39.5	63.18	11.42	25.39	36.81	1.40	<0.013	*	<0.033	*	<0.207 *
94-210	E3 -1	9.800039596	16.6	78.3	5.9	15.8	21.72	0.86	0.003	*	<0.003	*	0.028 *
94-211	F1 -1	27.77777778	83.9	1.6	25.0	73.4	98.39	4.22	<0.028	*	1.216	<0.446	<0.459 *
94-212	F2 -1	29.41741973	109.1	22.7	22.5	54.9	77.34	7.24	<0.021	*	<0.053	*	<0.335 *
94-213	G1 -1	39.89667049	76.0	2.1	26.8	71.2	97.92	3.42	<0.019	*	<0.050	*	<0.313 *
94-214	G2 -1	7.8	10.1	89.4	1.9	8.7	10.64	0.63	<0.001	*	0.005	*	<0.017 *
94-215	G3 -1	7.8	14.9	94.5	1.8	3.7	5.52	0.28	<0.002	*	<0.006	*	<0.042 *
94-216	Ref. No. Inlet								<0.003	*	0.054	*	<0.054 *
94-242	Ref. No. Inlet								0.003	*	<0.007	*	<0.044 *
	Ref. Redfish Bay												
	Ref. No. Inlet												
	Duplicates for averaged result represented above												
94-204	E2 -2												
94-204	E2 -2												

- ns non-significant
+ statistically significant for Dunnett's t-test at 0.05
++ statistically significant for Dunnett's t-test at 0.01
* statistically significant for Mann-Whitney
** statistically significant for Mann-Whitney and Dunnett's
*** statistically significant for Mann-Whitney, Dunnets, and Distribution-free
@ Sample mean was statistically different from control
~ Sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	Zn (μ mole/g)	Hg (μ mole/g)	SEM (μ mole/g)	AVS (μ mole/g)	SEM:AVS ratio	% TOC	NAPHTHALENE (GC, ug/kg)	2-METHYLNAPHTHALENE (GC, ug/kg)
94-191	A1 -1	0.067	.00004	0.093	0.227	0.410	0.40	9	13
94-192	A2 -2	0.169	.00032	0.246	0.155	1.587 t	0.47	10	15
94-193	A3 -1	<0.005 *	.00010 *	0.150	1.284	0.117	0.32	19	25
94-194	D1 -1	0.073	.00002 *	0.105	0.466	0.226	0.16	14	19
94-195	D2 -1	0.026	.00002 *	0.081	0.268	0.303	0.17 <	5	5
94-196	P1	0.166	.00004 *	0.228	0.343	0.665	1.46	26	30
94-197	B2 -1	0.506	.00014 *	0.804	5.846	0.138	2.52	106	113
94-198	C1 -1	<0.014 *	.00028 *	0.429	6.752	0.064	2.81	46	70
94-199	B1 -1	<0.009 *	.00018 *	0.275	7.118	0.039	3.82	49	83
94-200	C3 -1	0.064	.00004 *	0.131	0.003	42.842 t	3.58	21	35
94-201	C2 -1	0.209	.00021 *	0.532	15.881	0.034	3.36 <	5	18
94-202	D3 -2	0.017	.00002 *	0.054	0.092	0.582	0.15	5	9
94-203	E1 -1	0.177	.00022 *	0.578	5.061	0.114	1.91	16	21
94-204	E2 -2	<0.015 *	.00031 *	0.468	5.796	0.081	1.40	63	60
94-210	E3 -1	0.245	.00003 *	0.322	0.212	1.521 t	0.86	16	15
94-211	F1 -1	1.483	.00069 *	3.604	14.439	0.250	4.22	39	26
94-212	F2 -1	0.437	.00050 *	1.170	42.999	0.027	7.24	1074	178
94-213	G1 -1	0.380	.00620	1.071	15.482	0.069	3.42	35	42
94-214	G2 -1	0.126	.00023	0.166	0.177	0.936	0.63	7	11
94-215	G3 -1	0.052	.00086	0.141	1.348	0.105	0.28	6	8
94-216	Ref. No. Inlet	0.108	.00014 *	0.268	0.599	0.447			
94-242	Ref. No. Inlet	0.157	.00009 *	0.253	0.478	0.531			
	Ref. Redfish Bay								
	Ref. No. Inlet								
Duplicates for averaged result represented above									
94-204		E2-2							
94-204		E2-2							

- ns non-significant
- + statistically significant for Dunnett's t-test at 0.05
- ++ statistically significant for Dunnett's t-test at 0.01
- * statistically significant for Mann-Whitney
- ** statistically significant for Mann-Whitney and Dunnett's
- *** statistically significant for Mann-Whitney, Dunnets, and Distribution-free
- @ sample mean was statistically different from control
- ~ sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

Duplicates for averaged result represented above

E2-2
E2-2

ns	non-significant
+	statistically significant for Dunnett's t-test at 0.05
++	statistically significant for Dunnett's t-test at 0.01
*	statistically significant for Mann-Whitney
**	statistically significant for Mann-Whitney and Dunnett's
***	statistically significant for Mann-Whitney, Dunnets, and Distribution-free
@	Sample mean was statistically different from control
-	Sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	2,3,5-trime nap (GC, ug/kg)	FLUORENE (GC, ug/kg)	PHENANTHRENE (GC, ug/kg)	1-METHYLPHENANTHRENE (GC, ug/kg)	ANTHRACENE (GC, ug/kg)	FLUORANTHENE (GC, ug/kg)
94-191	A1 -1	1	<	1	3	11	2
94-192	A2 -2	1	1	4	2	2	10
94-193	A3 -1	1	2	15	2	6	33
94-194	D1 -1	1	<	1	3	2	9
94-195	D2 -1	1	<	1	2	2	5
94-196	P1	3	7	28	6	21	102
94-197	B2 -1	16	112	468	108	318	3599
94-198	C1 -1	2	12	102	<	2	383
94-199	B1 -1	7	49	268	34	196	1004
94-200	C3 -1	3	4	20	5	34	58
94-201	C2 -1	1	3	18	<	20	50
94-202	D3 -2	1	<	1	3	2	3
94-203	E1 -1	1	2	6	2	3	17
94-204	E2 -2	1	5	15	2	5	17
94-210	E3 -1	1	2	10	2	5	16
94-211	F1 -1	1	11	45	2	2	57
94-212	F2 -1	62	177	890	2	245	742
94-213	G1 -1	2	3	10	2	4	16
94-214	G2 -1	2	2	4	2	2	4
94-215	G3 -1	1	<	1	2	2	3
94-216	Ref. No. Inlet						
94-242	Ref. No. Inlet						
	Ref. Redfish Bay						
	Ref. No. Inlet						
Duplicates for averaged result represented above							
94-204	E2 -2						
94-204	E2 -2						

- ns non-significant
- + statistically significant for Dunnett's t-test at 0.05
- ++ statistically significant for Dunnett's t-test at 0.01
- *
- ** statistically significant for Mann-Whitney and Dunnets
- *** statistically significant for Mann-Whitney, Dunnets, and Distribution-free
- @ sample mean was statistically different from control
- sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	PYRENE (GC, ug/kg)	BENZ(A)ANTHRACENE (GC, ug/kg)	CHRYSENE (GC, ug/kg)	BENZO(B)FLUORANTHENE (GC, ug/kg)	BENZO(K)FLUORANTHENE (GC, ug/kg)
94-191	A1 -1	7	<	4	3	6
94-192	A2 -2	8	<	4	5	<
94-193	A3 -1	28	10	8	14	8
94-194	D1 -1	7	<	4	2	6
94-195	D2 -1	5	<	4	1	3
94-196	P1	125	50	57	152	163
94-197	B2 -1	3203	1020	1269	2074	1668
94-198	C1 -1	417	172	202	386	120
94-199	B1 -1	1112	433	524	854	390
94-200	C3 -1	49	32	56	58	32
94-201	C2 -1	47	24	37	52	28
94-202	D3 -2	4	<	4	<	<
94-203	E1 -1	16	4	5	1	3
94-204	E2 -2	14	5	5	11	4
94-210	E3 -1	13	17	21	35	15
94-211	F1 -1	7	16	21	34	15
94-212	F2 -1	1028	187	209	324	127
94-213	G1 -1	13	7	11	22	6
94-214	G2 -1	3	<	4	<	<
94-215	G3 -1	2	<	4	1	3
94-216	Ref. No. Inlet					
94-242	Ref. No. Inlet					
	Ref. Redfish Bay					
	Ref. No. Inlet					
Duplicates for averaged result represented above						
94-204	E2 -2					
94-204	E2 -2					

- ns non-significant
+ statistically significant for Dunnnett's t-test at 0.05
++ statistically significant for Dunnnett's t-test at 0.01
* statistically significant for Mann-Whitney
** statistically significant for Mann-Whitney, Dunnnett's, and Distribution-free
*** sample mean was statistically different from control
@ sample mean was statistically different from control
~ sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	BENZO(E)PYRENE (GC, ug/kg)	BENZO(A)PYRENE (GC, ug/kg)	PERYLENE (GC, ug/kg)	INDENO(1,2,3-C,D)PYRENE (GC, ug/kg)	DIBENZ(A,H)ANTHRACENE (GC, ug/kg)
94-191	A1 -1	3	<	5	<	3
94-192	A2 -2	3	<	5	<	5
94-193	A3 -1	5	<	7	<	5
94-194	D1 -1	3	<	5	<	5
94-195	D2 -1	3	<	5	<	5
94-196	P1	72	<	69	20	5
94-197	B2 -1	726	1021	248	564	103
94-198	C1 -1	120	164	49	125	21
94-199	B1 -1	328	441	123	273	49
94-200	C3 -1	22	30	17	23	1
94-201	C2 -1	20	25	9	24	1
94-202	D3 -2	3	<	5	<	1
94-203	E1 -1	5	<	6	44	5
94-204	E2 -2	5	7	5	6	1
94-210	E3 -1	11	18	32	15	3
94-211	F1 -1	13	15	33	5	1
94-212	F2 -1	120	144	100	138	1
94-213	G1 -1	9	13	9	10	1
94-214	G2 -1	3	<	5	5	1
94-215	G3 -1	3	<	5	5	1
94-216	Ref. No. Inlet					
94-242	Ref. No. Inlet					
	Ref. Redfish Bay					
	Ref. No. Inlet					
Duplicates for averaged result represented above						
94-204	E2 -2					
94-204	E2 -2					

ns non-significant

+ statistically significant for Dunnett's t-test at 0.05

++ statistically significant for Dunnett's t-test at 0.01

*

** statistically significant for Mann-Whitney and Dunnett's

*** statistically significant for Mann-Whitney, Dunnotts, and Distribution-free

@ sample mean was statistically different from control

- sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	BENZO(G,H,I)PERYLENE (GC, ug/kg)	Total PAH (GC, ug/kg)	FLUORENE (HPLC, ug/kg)	PHENANTHRENE (HPLC, ug/kg)	ANTHRACENE (HPLC, ug/kg)	FLUORANTHENE (HPLC, ug/kg)	PYRENE (HPLC, ug/kg)
94-191	A1 -1	3	85	9	6	<	8.3	<
94-192	A2 -2	3	73	11	5	<	8.3	<
94-193	A3 -1	3	207	34	17	<	8.3	5.7
94-194	D1 -1	3	84	9	5	<	8.3	5.7
94-195	D2 -1	3	27	6	<	2.7	8.3	5.7
94-196	P1	45	1042	96	32	19	<	5.7
94-197	B2 -1	431	17544	5690	479	485	153	3032
94-198	C1 -1	103	2692	374	108	42	<	5.7
94-199	B1 -1	212	6717	1032	270	195	<	5.7
94-200	C3 -1	15	562	60	25	34	5.7	33
94-201	C2 -1	14	398	57	26	24	5.7	34
94-202	D3 -2	3	34	<	5.9	<	2.7	5.7
94-203	E1 -1	3	187	17	7	<	8.3	5.7
94-204	E2 -2	3	310	18	17	<	8.3	7
94-210	E3 -1	8	280	17	11	<	8.3	5.7
94-211	F1 -1	9	524	55	94	26	<	5.7
94-212	F2 -1	134	8371	785	4999	271	184	542
94-213	G1 -1	4	269	22	15	<	8.3	5.7
94-214	G2 -1	3	57	6	7	<	8.3	5.7
94-215	G3 -1	3	24	<	5.9	<	2.7	5.7
94-216	Ref. No. Inlet						<	7.3
94-242	Ref. No. Inlet						<	7.3
	Ref. Redfish Bay							
	Ref. No. Inlet							
	Duplicates for averaged result represented above							
94-204	E2-2							
94-204	E2-2							

ns non-significant
+ statistically significant for Dunnert's t-test at 0.05
++ statistically significant for Dunnert's t-test at 0.01
* statistically significant for Mann-Whitney
** statistically significant for Mann-Whitney and Dunnett's
*** statistically significant for Mann-Whitney, Dunnnett's, and Distribution-free
@ sample mean was statistically different from control
~ sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	BENZ(A)ANTHRACENE (HPLC, ug/kg)	CHRYSENE (HPLC, ug/kg)	BENZO(B)FLUORANTHENE (HPLC, ug/kg)	BENZO(K)FLUORANTHENE (HPLC, ug/kg)	BENZO(A)PYRENE (HPLC, ug/kg)
94-191	A1 -1	4.8	13	<	8.8	<
94-192	A2 -2	4.8	<	7.3	8.8	<
94-193	A3 -1	12	8	<	8.8	5.3
94-194	D1 -1	4.8	<	7.3	<	5.3
94-195	D2 -1	4.8	<	7.3	8.8	8
94-196	P1	44	54	28	53	72
94-197	B2 -1	890	1160	399	1380	1443
94-198	C1 -1	148	134	71	162	197
94-199	B1 -1	266	446	142	204	274
94-200	C3 -1	31	54	29	40	39
94-201	C2 -1	31	51	19	35	36
94-202	D3 -2	4.8	<	7.3	8.8	<
94-203	E1 -1	5	<	7.3	50	7
94-204	E2 -2	7	<	7.3	10	12
94-210	E3 -1	19	22	38	26	31
94-211	F1 -1	14	10	54	6	19
94-212	F2 -1	88	152	211	1115	242
94-213	G1 -1	9	13	16	17	17
94-214	G2 -1	4.8	<	7.3	8.8	5.3
94-215	G3 -1	4.8	<	7.3	8.8	5.3
94-216	Ref. No. Inlet					
94-242	Ref. No. Inlet					
	Ref. Redfish Bay					
	Ref. No. Inlet					

Duplicates for averaged result represented above

94-204 E2-2
94-204 E2-2

- ns non-significant
- + statistically significant for Dunnett's t-test at 0.05
- ++ statistically significant for Dunnett's t-test at 0.01
- * statistically significant for Mann-Whitney and Dunnets
- ** statistically significant for Mann-Whitney, Dunnets, and Distribution-free
- @ sample mean was statistically different from control
- sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	PERYLENE (HPLC, ug/kg)	DIBENZ(A,H)ANTHRACENE (HPLC, ug/kg)	BENZO(G,H,I)PERYLENE (HPLC, ug/kg)	Total (HPLC, ug/kg)	CL2(08) ug/kg	HEXACHLOROBENZENE ug/kg
94-191	A1 -1	6	<	7.3	<	8.2	0.084
94-192	A2 -2	5.4	<	7.3	<	8.2	0.168
94-193	A3 -1	10	<	7.3	<	8.2	0.677
94-194	D1 -1	5.4	<	7.3	<	8.2	0.084
94-195	D2 -1	5.4	<	7.3	<	8.2	0.084
94-196	P1	82	<	7.3	47	637	9.077
94-197	B2 -1	704	102		778	16695	0.084
94-198	C1 -1	180	13		106	1853	0.084
94-199	B1 -1	455		30	211	3936	0.437
94-200	C3 -1	39	<	7.3	18	403	0.201
94-201	C2 -1	40	<	7.3	17	370	0.084
94-202	D3 -2	5.4	<	7.3	<	8.2	0.084
94-203	E1 -1	10	<	7.3	<	8.2	0.084
94-204	E2 -2	13	<	7.3	<	8.2	0.084
94-210	E3 -1	26	<	7.3	17	221	0.342
94-211	F1 -1	45	<	7.3	12	367	0.084
94-212	F2 -1	591	33		177	9389	0.084
94-213	G1 -1	24	<	7.3	13	162	0.084
94-214	G2 -1	5.4	<	7.3	<	8.2	0.21
94-215	G3 -1	5.4	<	7.3	<	8.2	0.084
94-216	Ref. No. Inlet						
94-242	Ref. No. Inlet						
	Ref. Redfish Bay						
	Ref. No. Inlet						
	Duplicates for averaged result represented above						
94-204	E2-2						
94-204	E2-2						

- ns non-significant
+ statistically significant for Dunnett's t-test at 0.05
++ statistically significant for Dunnett's t-test at 0.01
* statistically significant for Mann-Whitney
** statistically significant for Mann-Whitney, Dunnets, and Distribution-free
*** statistically significant for Mann-Whitney, Dunnets, and Distribution-free
@ sample mean was statistically different from control
~ sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	LINDANE ug/kg	CL3(18) ug/kg	HEPTACHLOR ug/kg	CL4(52) ug/kg	ALDRIN ug/kg	CL4(44) ug/kg	HEPTACHLOREPOXIDE ug/kg	CL4(66) ug/kg
94-191	A1 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.042 <	0.210 < 0.039 <
94-192	A2 -2	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.164 <	0.210 < 0.039 <
94-193	A3 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.185 <	0.210 < 0.292 <
94-194	D1 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.042 <	0.210 < 0.039 <
94-195	D2 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.042 <	0.210 < 0.039 <
94-196	P1	0.145 <	0.253 <	0.454 <	0.307	1.616 <	0.220 <	0.888 <	0.210 < 1.064 <
94-197	B2 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.434 <	0.210 < 0.967 <
94-198	C1 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.901 <	0.220 <	0.574 <	0.210 < 0.741 <
94-199	B1 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.35 <	0.210 < 1.171 <
94-200	C3 -1	0.201 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.361 <	0.210 < 0.039 <
94-201	C2 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.042 <	0.210 < 0.039 <
94-202	D3 -2	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.042 <	0.210 < 0.039 <
94-203	E1 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.141 <	0.210 < 0.039 <
94-204	E2 -2	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.106 <	0.210 < 0.039 <
94-210	E3 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.344 <	0.220 <	0.042 <	0.210 < 0.039 <
94-211	F1 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.042 <	0.210 < 0.039 <
94-212	F2 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.042 <	0.210 < 0.039 <
94-213	G1 -1	0.596 <	0.253 <	0.454 <	0.167 <	0.735 <	0.220 <	0.367 <	0.210 < 0.039 <
94-214	G2 -1	0.145 <	0.253 <	0.454 <	0.167 <	1.234 <	0.220 <	0.042 <	0.210 < 0.039 <
94-215	G3 -1	0.145 <	0.253 <	0.454 <	0.167 <	0.204 <	0.220 <	0.042 <	0.210 < 0.039 <
94-216	Ref. No. Inlet								
94-242	Ref. No. Inlet								
	Ref. Redfish Bay								
	Ref. No. Inlet								

Duplicates for averaged result represented above

94-204 E2-2
94-204 E2-2

- ns non-significant
- + statistically significant for Dunnett's t-test at 0.05
- ++ statistically significant for Dunnett's t-test at 0.01
- * statistically significant for Mann-Whitney
- ** statistically significant for Mann-Whitney and Dunnett's
- *** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free
- @ sample mean was statistically different from control
- sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	2,4-DDE		CL5(101)		alpha-Chlordane		TRANS-NONACHLOR		DIELDRIN		4,4-DDE		CL4(77)		2,4-DDD		CL5(118)	
		ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	
94-191	A1 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.190 <	0.371 <	0.403 <	0.345 <	0.190 <	0.794 <	0.086 <	0.086 <		
94-192	A2 -2	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.190 <	0.371 <	0.403 <	0.345 <	0.190 <	0.794 <	0.103 <	0.103 <		
94-193	A3 -1	0.217	0.166 <	0.359 <	0.345 <	0.403 <	0.371 <	0.546 <	0.794 <	0.359 <	0.403 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.258 <	0.258 <	
94-194	D1 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.359 <	0.403 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086 <	0.086 <	
94-195	D2 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.359 <	0.403 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086 <	0.086 <	
94-196	P1	0.217	1.827 <	0.359 <	0.345 <	0.938	1.577	4.167	11.962	1.827 <	0.359 <	0.345 <	0.938	1.577	4.167	11.962	1.513	1.513	
94-197	B2 -1	0.217	0.619	0.619	0.555 <	0.403 <	0.371 <	0.190 <	0.794 <	0.619	0.619	0.555 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086 <	0.086 <	
94-198	C1 -1	0.217	0.442 <	0.442 <	0.345 <	0.403 <	0.562 <	0.190 <	0.794 <	0.442 <	0.442 <	0.345 <	0.403 <	0.562 <	0.190 <	0.794 <	0.653	0.653	
94-199	B1 -1	0.217	0.627	0.627 <	0.345 <	0.403 <	0.371 <	1.019 <	0.794 <	0.627	0.627 <	0.345 <	0.403 <	0.371 <	1.019 <	0.794 <	0.710	0.710	
94-200	C3 -1	0.217	0.273 <	0.359 <	0.345 <	0.403 <	0.371 <	0.865 <	0.794 <	0.273 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.387	0.387	
94-201	C2 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086	0.086
94-202	D3 -2	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086	0.086
94-203	E1 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086	0.086
94-204	E2 -2	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.096	0.096
94-210	E3 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.453	0.424	0.950	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.453	0.424	0.950	0.149	0.149
94-211	F1 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086	0.086
94-212	F2 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086	0.086
94-213	G1 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086	0.086
94-214	G2 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086	0.086
94-215	G3 -1	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.217 <	0.092 <	0.359 <	0.345 <	0.403 <	0.371 <	0.190 <	0.794 <	0.086	0.086
94-216	Ref. No. Inlet																		
94-242	Ref. No. Inlet																		
94-204	Ref. Redfish Bay																		
94-204	Ref. No. Inlet																		

Duplicates for averaged result represented above

94-204 E2-2
94-204 E2-2ns non-significant
+ statistically significant for Dunnett's t-test at 0.05
++ statistically significant for Dunnnett's t-test at 0.01
* statistically significant for Mann-Whitney and Dunnets
** statistically significant for Mann-Whitney and Dunnets, and Distribution-free
@ sample mean was statistically different from control
~ sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	4,4-DDD ug/kg	2,4-DDT ug/kg	CL6(153) ug/kg	CL5(105) ug/kg	4,4-DDT ug/kg	CL6(138) ug/kg	CL5(126) ug/kg	CL7(187) ug/kg	CL6(128) ug/kg	CL7(180) ug/kg
94-191	A1 -1	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215	1.104 <	0.050	0.286 <
94-192	A2 -2	0.416 <	0.382	0.158 <	0.086 <	0.367 <	0.232 <	0.215	1.500 <	0.050	0.448 <
94-193	A3 -1	0.416 <	0.382	0.374 <	0.086 <	0.367	0.264 <	0.215	6.601 <	0.050	2.052 <
94-194	D1 -1	0.416 <	0.382	0.159 <	0.086 <	0.367 <	0.232 <	0.215	1.528 <	0.050	0.508 <
94-195	D2 -1	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215	1.133 <	0.050	0.358 <
94-196	P1	1.400 <	0.382	11.266 <	0.086 <	0.367	0.744 <	0.215	56.760 <	0.050	18.518 <
94-197	B2 -1	0.416 <	0.382	1.553 <	0.086	1.750	2.332 <	0.215	2.772 <	0.050	0.764 <
94-198	C1 -1	1.674 <	0.382	1.183 <	0.086	0.410	0.578 <	0.215	4.610 <	0.050	1.323 <
94-199	B1 -1	3.267 <	0.382	1.570 <	0.086 <	0.367	0.870 <	0.215	6.607 <	0.050	2.280 <
94-200	C3 -1	0.416 <	0.382	1.363 <	0.086 <	0.367	0.338 <	0.215	3.587 <	0.050	0.991 <
94-201	C2 -1	0.416 <	0.382	0.321 <	0.086 <	0.367 <	0.232 <	0.215	4.535 <	0.050	1.443 <
94-202	D3 -2	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215	0.470 <	0.050	0.116 <
94-203	E1 -1	0.416 <	0.382	0.475 <	0.086 <	0.367 <	0.232 <	0.215	1.613 <	0.050	0.494 <
94-204	E2 -2	0.416 <	0.382	0.430 <	0.086 <	0.367 <	0.232 <	0.215	2.325 <	0.050	0.730 <
94-210	E3 -1	0.818 <	0.382	0.582 <	0.086 <	0.367 <	0.232 <	0.215	3.214 <	0.050	0.680 <
94-211	F1 -1	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215	0.060 <	0.050	0.036 <
94-212	F2 -1	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215	0.060 <	0.050	0.036 <
94-213	G1 -1	3.354 <	0.382	1.367 <	0.086 <	0.367 <	0.232	1.265	1.589 <	0.050	0.036 <
94-214	G2 -1	0.416 <	0.382	0.421 <	0.086 <	0.367 <	0.232 <	0.215	0.060 <	0.050	0.036 <
94-215	G3 -1	0.416 <	0.382 <	0.124 <	0.086 <	0.367 <	0.232 <	0.215	0.060 <	0.050	0.036 <
94-216	Ref. No. Inlet										
94-242	Ref. No. Inlet										
	Ref. Redfish Bay										
	Ref. No. Inlet										

Duplicates for averaged result represented above

94-204 E2-2
94-204 E2-2

- ns non-significant
- + statistically significant for Dunnett's t-test at 0.05
- ++ statistically significant for Dunnett's t-test at 0.01
- * statistically significant for Mann-Whitney and Dunnett's
- ** statistically significant for Mann-Whitney, Dunnets, and Distribution-free
- @ sample mean was statistically different from control
- ~ sample mean 80% or less than the control

Appendix B3 continued. Toxicity and chemistry data from St. Simons Sound.

MSL Lab Id No.	STATION No.	MIREX	CL7(170)	CL8(195)	CL9(206)	CL10(209)	sum 20PCBs	X2	total PCBs	Pesticide Total :	DDT Total :
		ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg	ug/kg
94-191	A1 -1	0.212 <	0.127 <	0.088	8.850	1.933	14.729	2	29.5	2.145	2.547
94-192	A2 -2	0.212 <	0.127	0.147	17.508	3.984	26.781	2	53.6	2.229	2.547
94-193	A3 -1	0.212 <	0.127	0.857	83.118	19.766	116.142	2	232.3	2.738	2.547
94-194	D1 -1	0.212 <	0.127	0.246	23.723	5.695	34.203	2	68.4	2.145	2.547
94-195	D2 -1	0.212 <	0.127	0.358	16.005	3.210	23.532	2	47.1	2.145	2.547
94-196	P1	0.212 <	0.127	5.922	612.639	169.425	887.940	2	1775.9	11.813	15.905
94-197	B2 -1	0.212 <	0.127 <	0.088	22.379	5.541	39.927	2	79.9	2.615	3.930
94-198	C1 -1	0.212 <	0.127	0.371	63.819	14.935	91.779	2	183.6	2.228	4.039
94-199	B1 -1	0.212 <	0.127	0.308	94.122	21.330	133.205	2	266.4	2.766	5.398
94-200	C3-1	0.212 <	0.127	0.539	46.037	9.397	66.002	2	132.0	2.318	2.547
94-201	C2 -1	0.212 <	0.127	1.234	67.571	14.381	91.829	2	183.7	2.145	2.547
94-202	D3 -2	0.212 <	0.127 <	0.088	8.007	1.572	12.741	2	25.5	2.145	2.547
94-203	E1 -1	0.212 <	0.127	0.324	19.281	3.891	28.521	2	57.0	2.145	2.547
94-204	E2 -2	0.212 <	0.127	0.658	32.074	7.417	46.052	2	92.1	2.145	2.547
94-210	E3-1	0.212 <	0.127	0.350	24.705	5.190	37.502	2	75.0	2.403	3.187
94-211	F1-1	0.212 <	0.127 <	0.088 <	0.158 <	0.069	2.879	2	5.8	2.145	2.547
94-212	F2-1	0.212 <	0.127 <	0.088 <	0.158 <	0.069	2.879	2	5.8	2.145	2.547
94-213	G1-1	0.212 <	0.127	0.857	10.081	2.321	20.522	2	41.0	2.596	5.485
94-214	G2-1	0.212 <	0.127 <	0.088	1.387	0.228	5.594	2	11.2	2.271	2.547
94-215	G3-1	0.212 <	0.127 <	0.088	0.335 <	0.069	3.056	2	6.1	2.145	2.547
94-216	Ref. No. Inlet										
94-242	Ref. No. Inlet										
	Ref. Redfish Bay										
	Ref. No. Inlet										
	Duplicates for averaged result represented above										
94-204	E2-2										
94-204	E2-2										

ns non-significant
+ statistically significant for Dunnett's t-test at 0.05
++ statistically significant for Dunnett's t-test at 0.01
* statistically significant for Mann-Whitney
** statistically significant for Mann-Whitney and Dunnett's
*** statistically significant for Mann-Whitney, Dunnett's, and Distribution-free
@ sample mean was statistically different from control
~ sample mean 80% or less than the control

Appendix C. Method detection limits (MDLs) for organics and trace metals.

Substance	Method of Analysis	MDL*
Fluorene	HPLC+GC-MS	5.7 ng/g
Phenanthrene	HPLC+GC-MS	2.7 ng/g
Anthracene	HPLC+GC-MS	8.3 ng/g
Fluoranthene	HPLC+GC-MS	5.9 ng/g
Pyrene	HPLC+GC-MS	7.3 ng/g
Benz(a)anthracene	HPLC+GC-MS	4.8 ng/g
Chrysene	HPLC+GC-MS	7.3 ng/g
Perylene	HPLC+GC-MS	5.4 ng/g
Benzo(b)fluoranthene	HPLC+GC-MS	8.8 ng/g
Benzo(k)fluoranthene	HPLC+GC-MS	5.3 ng/g
Benzo(a)pyrene	HPLC+GC-MS	7.4 ng/g
Dibenzo(a,h)anthracene	HPLC+GC-MS	7.3 ng/g
Benzo(g,h,i)perylene	HPLC+GC-MS	8.2 ng/g
HCB	GC-ECD	0.5 ng/g
y - HCH	GC-ECD	0.5 ng/g
Heptachlor	GC-ECD	0.7 ng/g
Aldrin	GC-ECD	0.5 ng/g
Heptachlor epoxide	GC-ECD	0.8 ng/g
2, 4'-DDE	GC-ECD	0.6 ng/g
cis-Chlordane	GC-ECD	0.8 ng/g
trans-Chlordane	GC-ECD	0.8 ng/g
Die�drin	GC-ECD	1.0 ng/g
4, 4'-DDE	GC-ECD	0.9 ng/g
2, 4'-DDD	GC-ECD	2.3 ng/g
4, 4'-DDD	GC-ECD	0.9 ng/g
2, 4'-DDT	GC-ECD	0.8 ng/g
4, 4'-DDT	GC-ECD	0.9 ng/g
Mirex	GC-ECD	0.7 ng/g
PCB congeners (21)	GC-ECD	0.29-2.27 ng/g
Cadmium	ICP	0.3 ug/g
Copper	ICP	0.2 ug/g
Lead	ICP	0.8 ug/g
Nickel	ICP	1.4 ug/g
Chromium	ICP	0.4 ug/g
Zinc	ICP	0.3 ug/g
Aluminum	ICP	882 g/kg
Iron	ICP	297 g/kg
Silver	GF-AA	0.1 ug/g
Manganese	ICP	0.3 ug/g
Arsenic	ICP	5.7 ug/g
Selenium	GF-AA	0.1 ug/g
Tin	ICP	7.8 ug/g
Lead	GF-AA	0.4 ug/g
Arsenic	GF-AA	0.1 ug/g

*MDLs established for PAHs from the standard deviations of the analyses of the spikes, for organochlorines as three times the standard deviation of repeated measures, and for metals as the mean blank plus 3 standard deviations (where mean blank is negative, MDL set as 0.0).